

Are your **MRI contrast agents** cost-effective?

Learn more about generic **Gadolinium-Based Contrast Agents**.



FRESENIUS  
KABI

caring for life

# AJNR

## **The influence of volumetric tumor doubling time, DNA ploidy, and histologic grade on the survival of patients with intracranial astrocytomas.**

F G Blankenberg, R L Teplitz, W Ellis, M S Salamat, B H Min, L Hall, D B Boothroyd, I M Johnstone and D R Enzmann

This information is current as of April 20, 2024.

*AJNR Am J Neuroradiol* 1995, 16 (5) 1001-1012  
<http://www.ajnr.org/content/16/5/1001>

# The Influence of Volumetric Tumor Doubling Time, DNA Ploidy, and Histologic Grade on the Survival of Patients with Intracranial Astrocytomas

Francis G. Blankenberg, Raymond L. Teplitz, William Ellis, M. Shahriar Salamat, Byung Hee Min, Lisa Hall, Derek B. Boothroyd, Iain M. Johnstone, and Dieter R. Enzmann

**PURPOSE:** To improve the prediction of individual survival in patients with intracranial astrocytomas through the analysis of volumetric tumor doubling time ( $VD_t$ ) and DNA ploidy. **METHODS:** A pilot study was retrospectively conducted on a group of 25 patients with intracranial astrocytomas in whom recurrent and/or progressive disease was observed on serial contrast-enhanced CT or MR examinations.  $VD_t$  was computed using two or more data points from a semilogarithmic plot of tumor volume versus time. Size-adjusted survival was calculated using a method based on  $VD_t$  and initial tumor volume to decrease the lead time bias attributable to differing tumor sizes at presentation. **RESULTS:** Slower  $VD_t$  was associated with significantly longer survival and size-adjusted survival as determined by a univariate Cox proportional hazard analysis. Aneuploidy was a significant indicator of poor survival. Aneuploid and multiclonal astrocytomas had poor size-adjusted survivals compared with diploid astrocytomas. Grade IV astrocytomas had significantly poorer survival and size-adjusted survival compared with lower grades (I to III), which individually were not significantly correlated. However, grade IV histology was not a significant independent predictor of size-adjusted survival in a multivariate Cox model, whereas  $VD_t$  and DNA ploidy remained significant.  $VD_t$  also had a significant direct linear correlation to survival and size-adjusted survival. **CONCLUSIONS:**  $VD_t$  and DNA ploidy were more sensitive than histologic grading as indicators of individual survival. Initial tumor size needs to be considered when staging and assessing survival in patients with intracranial astrocytomas.

**Index terms:** Astrocytoma; Brain, neoplasms

*AJNR Am J Neuroradiol* 16:1001–1012, May 1995

The survival of patients with intracranial astrocytomas has been studied extensively with respect to histologic grade, DNA ploidy, proliferative index (as measured by bromodeoxyuridine and thymidine labeling indices and flow cytometry), and more recently by oncogene analysis (1–21) in an attempt to improve prog-

nostic accuracy. Despite improved methods for determining prognosis from histologic grading of astrocytomas in groups of patients (1, 2), the outlook for an individual patient has remained uncertain. The use of DNA ploidy as measured by flow cytometry and image cytometry in conjunction with histologic grade to predict individual survival is a subject of controversy in the literature (3–11), and other studies have shown an inconsistent relationship between labeling indices of cellular proliferation and survival (12–17).

Two studies have attempted to correlate volumetric tumor doubling time ( $VD_t$ ) with histologic grade, with limited success (22, 23). Our study focused on a group of 25 patients with intracranial astrocytomas that radiographically demonstrated growth of the primary and/or recurrent tumor after initial resection

---

Received February 17, 1994; accepted after revision December 14.

Sponsored by a grant from the Office of Technology and Licensing, Stanford University, June 1990.

From the Department of Diagnostic Radiology, Stanford (Calif) University School of Medicine (F.G.B., D.R.E.); Departments of Statistics and Health Research and Policy, Stanford University (D.B.B., I.M.J.); and Department of Medical Pathology, University of California at Davis, Sacramento (R.L.T., W.E., M.S.S., B.H.M., L.H.).

Address reprint requests to Francis G. Blankenberg, MD, Lucile Packard Children's Hospital at Stanford, 725 Welch Rd, Stanford, CA 94304.

*AJNR* 16:1001–1012, May 1995 0195-6108/95/1605–1001

© American Society of Neuroradiology

and adjuvant therapy. Our purpose was to correlate survival, DNA ploidy, and histologic grade with  $VD_t$  in an effort to improve prognostication and understanding of the cellular dynamics in patients with intracranial astrocytomas.

## Materials and Methods

### *Patient and Specimen Selection*

Twenty-five patients with intracranial astrocytomas who had serial computed tomography (CT) or magnetic resonance (MR) examinations demonstrating residual and/or recurrent tumor were retrospectively studied in cooperation with the Division of Neuroradiology at Stanford University Hospital and the Department of Pathology (Neuropathology Section) of the University of California at Davis. Twenty-four patients underwent primary resection and/or diagnostic biopsy of their brain tumors from July 1979 to April 1991. One additional patient (patient 4) did not undergo biopsy or therapy but was included, because the clinical presentation, course, and radiographic appearance were consistent with a high-grade astrocytoma (glioblastoma multiforme). The average age was 43.5 years with a range of 5 months to 76 years; there were a total of 15 female and 10 male patients; 23 patients had supratentorial astrocytomas, one a cerebellar astrocytoma, and one a pontine astrocytoma.

### *Histology and DNA Image Cytometry*

Eighteen patients had their primary histologic material available for review by W.E. and S.S., who were blinded to the results of previous pathologic interpretation, DNA ploidy,  $VD_t$ , and survival. Six patients were initially evaluated by the Division of Neuropathology at Stanford University Hospital; however, no paraffin-embedded tissue from their primary biopsy or resection was available for review. One patient (patient 4) did not undergo biopsy but was included in the study as mentioned above. Histologic grading followed the criteria of Dumas-Duport et al (1), which emphasize the presence of mitoses, necrosis, and endothelial proliferation. In this study, pilocytic astrocytomas were classified as grade I along with fibrillary astrocytomas that lacked all criteria for anaplasia. The designation *gemistocytic astrocytoma* indicated that the majority of the neoplastic astrocytes had a gemistocytic form. The reference histologic diagnosis used for each case was based on the primary surgical biopsy and/or resection. Seventeen patients had sufficient paraffin-embedded material from their primary tumors for determination of DNA ploidy. DNA image cytometry was used for the estimation of DNA ploidy in a technique described by Teplitz et al (24). The DNA histograms were interpreted in a blinded fashion by R.L.T. Studies were performed on 7- $\mu$ m sections of suitable paraffin blocks, stained specifically for DNA by the Cell Analysis (CAS, Elmhurst, Ill) procedure and examined with the CAS model 200 image

cytometer. A 280-nm filter was used with a 20-nm band pass and the CAS thickness correction program. To avoid the possibility of detecting only clonal expansion within these tumors, a minimum of 25 nuclei were analyzed on four separate quadrants of each tumor. No cases were included when the minimum number of determinations (100 determinations) could not be obtained. In most cases several hundred nuclei were quantitated. DNA ploidy results were classified in the following manner: D, diploid, that is, normal somatic cell DNA content (7.18 pg/cell); E, euploid, that is, diploid or an even multiple of diploid; A, aneuploid, that is, abnormal somatic cell DNA content; and MC, multiclonal, that is, two or more major clones.

### *Radiographic Tumor Volume Estimation and Calculation of $VD_t$*

Eleven patients had contrast-enhanced serial CT examinations to determine residual and/or recurrent tumor growth. Nine patients had serial MR examinations, five with serial T1-weighted images (600–800/20/2 [repetition time/echo time/excitations]) with or without gadolinium enhancement and four with serial T2-weighted images, (2000–2500/30, 80/2), and five patients had a combination of contrast enhanced CT and/or MR T1-weighted images with or without gadopentetate dimeglumine. The margins of only enhancing solid tumor tissue as seen on CT or MR imaging were used for determination of tumor volume. When only T2-weighted images were available, vasogenic edema (ie, white matter edema) was not included in the volume determinations. Although the actual margins were more difficult to determine on nonenhanced T2-weighted MR images, we were able to define the bulk of a tumor based on its abnormal signal and mass effect. A similar set of methods for determination of tumor margins for radiation therapy planning has been described by Galoway et al (25) and others (26, 27).

The calculation of the volume of gross tumor was done retrospectively from the images of different scanners using the method described by Breiman et al (28). Briefly, the cross-sectional area of a lesion was calculated by planimetry. The absolute cross-sectional area of each square (in centimeters) was determined from the centimeter scale on the filmed image. Tumor volumes were calculated by multiplying the absolute cross-sectional area on each MR or CT section by section thickness and summing the resultant products. To adjust for intersection gaps on MR scans, the average cross-sectional area of two contiguous sections was multiplied by the section gap thickness; the product was then added to the total volume. Tumor volumes were performed by F.B., who was blinded to the histologic grade, DNA ploidy, and survival data.

Volumetric tumor doubling time of the residual primary or recurrent disease was calculated using two or more data points separated by at least 1 month from a semilogarithmic plot of tumor volume versus time as described by Collins (29) and others (30–32). Only periods in which there was clear tumor progression demonstrated on serial imaging studies before or between tumor resections were

used for the calculation of  $VD_t$ . Patients with three or more data points had  $VD_t$  calculated from the slope of the best-fit line generated by a simple linear regression. Ten patients had two serial volume determinations, eight had three ( $r \geq .789$ ), and two had six ( $r \geq .972$ ). Five patients had two or more periods of clinical progression or disease recurrence separated by surgical resection and/or radiation therapy in which the resultant  $VD_t$  of the recurrent and/or residual disease of each time period was averaged. The intervals in which it was possible to measure  $VD_t$  were as follows: 11 patients after primary resection and radiation therapy, three after a second resection, one after the third resection, three after primary radiation therapy without resection, two before and after radiation therapy without resection, two after the primary and third resections, one after the primary and second resections, and two patients after radiation therapy only.

Size-adjusted survival was computed using a method previously described by Collins et al (29) and others (30–32) to eliminate the lead-time bias for primary tumors of differing sizes at diagnosis. The volume of each tumor at diagnosis was converted into the approximate number of tumor doublings from its theoretical origin as a single neoplastic cell (assuming an average cell diameter of 10  $\mu\text{m}$ ). An arbitrary number of 30 tumor doublings (a 1-cm-diameter tumor) was subtracted from the observed volume of each tumor at diagnosis (also expressed as the number of tumor doublings). A volume of 30 doublings was chosen, because this usually represents the smallest size at which a central nervous system tumor can be detected clinically (33). This was then multiplied by  $VD_t$ , and the resultant product (ie, the extrapolated number of months that were needed for a tumor to reach the observed size at diagnosis from an initial size of 1 cm) was then added to the observed survival to obtain size-adjusted survival. The volume of tumor at death was extrapolated in a similar fashion from the last radiographically determined tumor volume after completion of all therapy to the time of death. The formulas for the calculation of the number of tumor doublings, size-adjusted survival, and estimated tumor volume at death are given in the Appendix.

#### Statistical Analysis

Cox proportional hazard models were fitted for survival and size-adjusted survival using various subsets of the possible predictor variables as covariates. Likelihood ratio tests were performed for assessing the significance of models, whereas Wald statistics and coefficient/SE were used to evaluate the significance of particular coefficients in models involving more than one covariate by comparison with standard normal quantities. Survival analyses were performed using S-PLUS software (Statistical Sciences). All  $P$  values are based on asymptotic approximations. Survival, size-adjusted survival,  $VD_t$ , and tumor volume were all converted to natural logarithms before computation of their respective averages mentioned in the text assuming log normal distributions for each of the above quantities, according to Spratt et al (34, 35). The average

TABLE 1: Analysis of survival and size-adjusted survival with respect to  $VD_t$ , histologic grade, and DNA ploidy

Univariate Models	Survival, $P$	Size-adjusted Survival, $P$
Tumor growth rate		
$VD_t$	.02	<.001
Histology		
Grades I–IV	.007	.03
Grades I–III (excluding IV)	.77, NS	.47, NS
Grade IV (vs non–Grade IV)	.007	.006
DNA ploidy		
Diploid, euploid, aneuploid, and multiclonal	.14, NS	.007
Aneuploid (coeff)*	.05	.003
Multiclonal (coeff)	NS	.02
Best-fit multivariate models		
$VD_t$ and Grade IV	.001	...
$VD_t$ (coeff)	.04	...
Grade IV (coeff)	.01	...
$VD_t$ and diploid	...	<.001
$VD_t$ (coeff)	...	<.001
Diploid (coeff)	...	.03

\* Coefficient of significance of a single variable in models involving more than one covariate. NS indicates not significant.

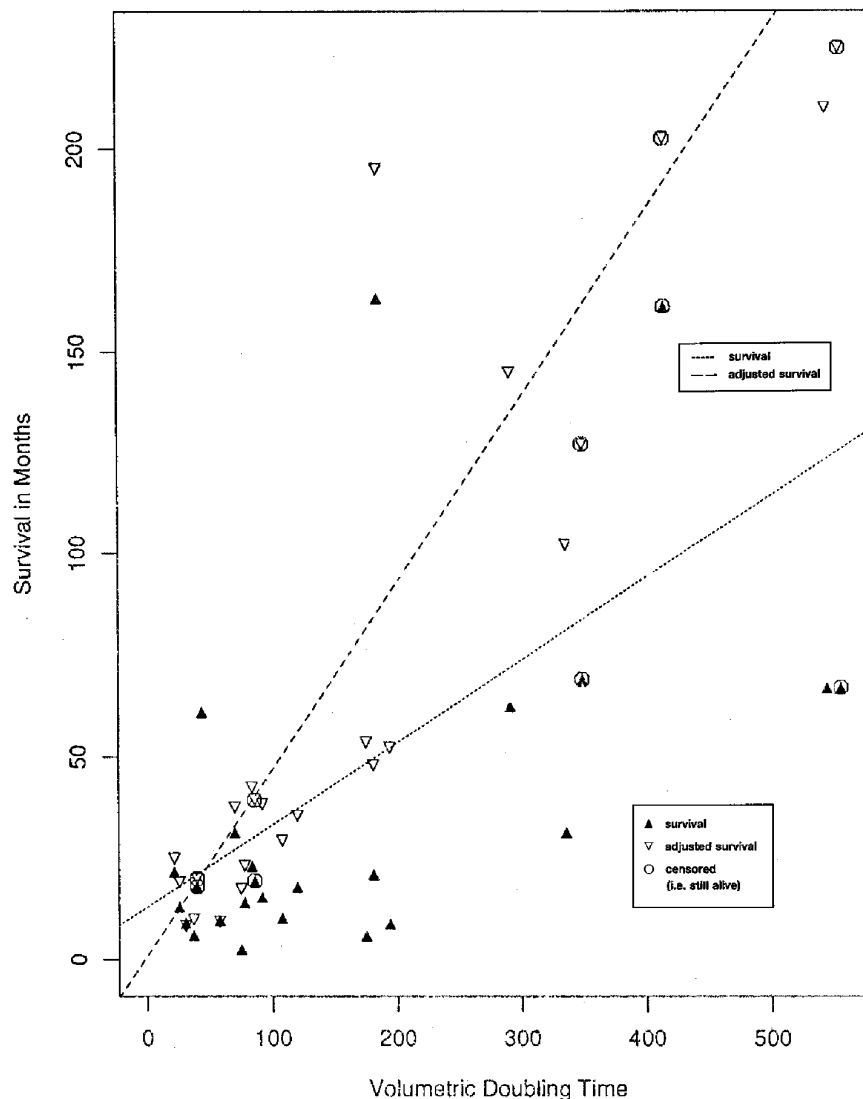
values mentioned in the text represent the geometric means of survival, size-adjusted survival, and  $VD_t$ . Comparisons of averages (ie, geometric means) were performed using a two-tailed Student's  $t$  test. Linear regressions of survival and size-adjusted survival on  $VD_t$  were calculated by the method of Buckley and James for regression with censored data, that is, patients still alive at the completion of the study (36), using a function written in S-PLUS by one of the authors (D.B.B.).

## Results

### Survival

Overall,  $VD_t$  and the presence or absence of grade IV histology were the most important predictors of (unadjusted) survival in the univariate and as multivariate Cox models (Table 1). Longer survival was seen in patients with longer  $VD_t$  (ie, slowly growing tumors), and shorter survival was seen in patients with grade IV tumors. A univariate Cox model including only grade I to III tumors, excluding grade IV astrocytomas, did not show any significant differences between the survival of each grade (nb, patients evaluated as grades I–II were considered grade II in our analysis). The presence of aneuploidy was of borderline significance with respect to survival in a univariate model, but this did not persist after the adjustment of the other covariates in a multivariate model.

Fig 1. Relationship of survival time (*solid triangle*) and adjusted survival (*inverted triangle*) in months, to  $VD_t$  in days for each patient. Also shown are fitted (Buckley-James) regression lines for both survival responses. Censored patients (ie, patients who were still alive at the end of the study) are *circled*.



### Size-Adjusted Survival

The analysis of size-adjusted survival was performed on 24 of 25 patients, because one patient (patient 6) did not have sufficient documentation of the size of the primary tumor available.

Overall,  $VD_t$  and DNA ploidy were the most important predictors of size-adjusted survival both in the univariate and multivariate Cox models; long  $VD_t$  and tumor diploidy were correlated with a reduced hazard (see Table 1). The presence or absence of grade IV histology was of significance in a univariate model, but this did not persist after adjusting for the other covariates in a multivariate model. In addition, a univariate model including only grade I to III astrocytomas did not reveal any significant dif-

ferences among the size-adjusted survivals of each grade.

### $VD_t$ and Tumor Volume

There was a significant linear relationship between  $VD_t$  and survival ( $P = .0011$ ) and also with size-adjusted survival ( $P < .001$ ) as shown in Figure 1. The five patients who were alive at the end of the study (censored data) were included in the analysis. The slopes of survival and size-adjusted survival lines obtained using Buckley and James' method were  $6.09 \pm 1.98$  and  $13.8 \pm 1.89$  tumor doublings, respectively (ie, number of tumor doublings = survival [or size-adjusted survival]/ $VD_t$ ). More than one  $VD_t$  was able to be calculated in five patients at

TABLE 2: Measurement of  $VD_t$  during different clinical periods

Patient	Average $VD_t$ , d	$VD_t$	Clinical Period
8	70	75 d (2 points)	pre-radiation therapy (5/84-12/84)
		79 d (3 points, $r = .878$ )	7 mo after 1st radiation therapy (8/85-9/85)
		56 d (5 points, $r = .789$ )	2 mo after 2nd radiation therapy (12/85-5/86)
13	92	102 d (3 points, $r = .971$ )	6 mo after 1st resection/radiation therapy (12/88-4/89)
		84 d (4 points, $r = .936$ )	1 mo after 2nd resection/radiation therapy (4/89-8/89)
17	181	182 d (2 points)	during and immediately after radiation therapy (7/86-10/86)*
		181 d (2 points)	4 months after radiation therapy (1/87-8/87)
22	349	327 d (2 points)	4 mo after 1st resection/radiation therapy (7/88-7/89)
		370 d (4 points, $r = .977$ )	5 mo after 3rd resection (7/90-12/91)
25	556	530 d (4 points, $r = .88$ )	2 mo after 1st resection/radiation therapy (8/87-6/88)
		582 d (3 points, $r = .863$ )	1 mo after 3rd resection (7/89-4/91)

Note.—Patient 17 did not undergo a primary resection of tumor.

different periods of tumor progression (primary or secondary tumor) as listed in Table 2. In these patients,  $VD_t$  of the primary or recurrent disease changed little despite intervening resection and/or radiation therapy over a time interval ranging from 8 to 44 months as shown in Figure 2.

Diploid and euploid tumors had a significantly longer average  $VD_t$  ( $P < .001$ ) compared with aneuploid and multiclonal astrocytomas (Table 3). No tumor in the diploid group had a  $VD_t$  of less than 84 days. Conversely, no tumor of the aneuploid and multiclonal groups had a  $VD_t$  greater than 92 days (Table 4). Grade IV

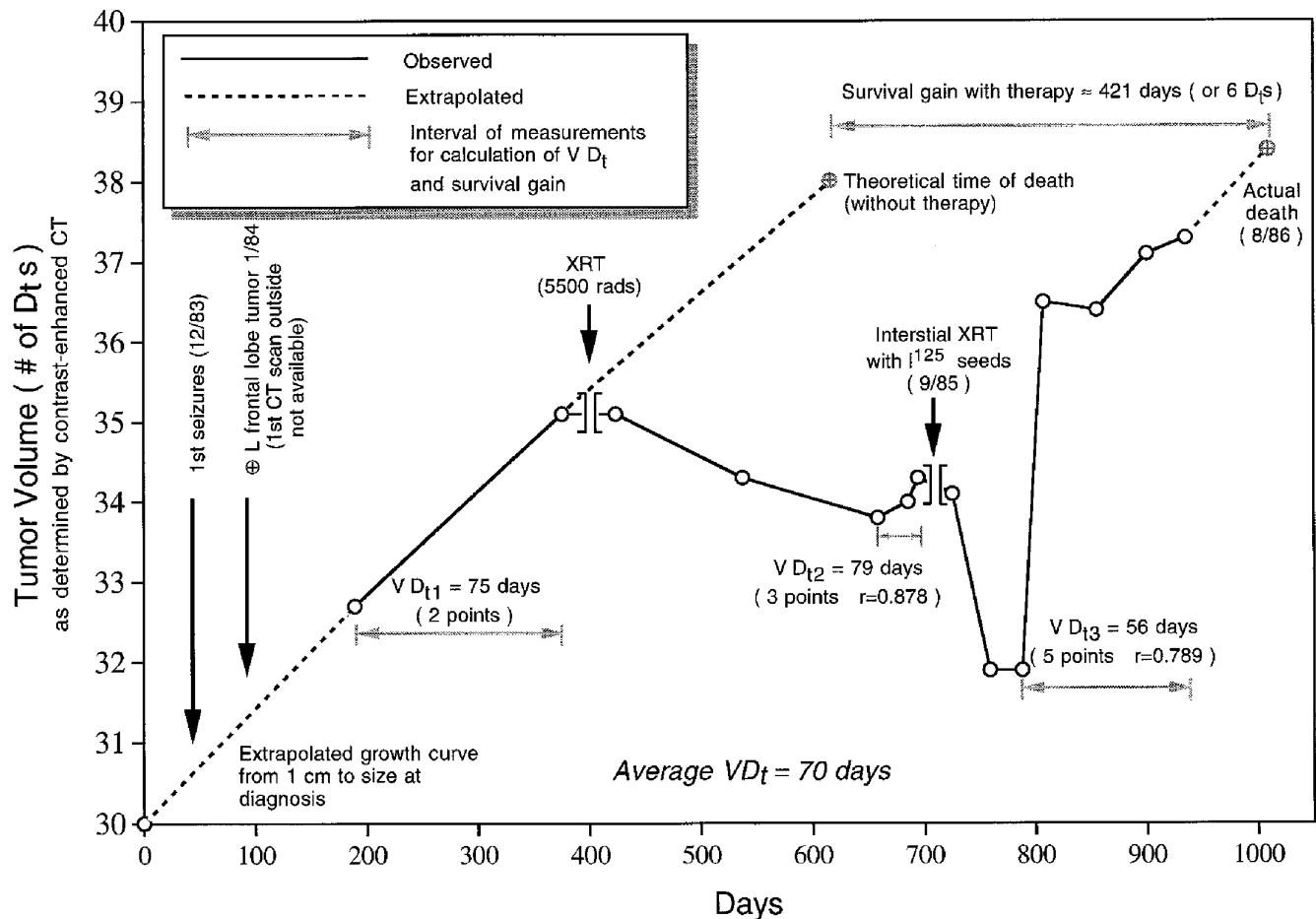


Fig 2. Tumor volume measurements of patient 8 from diagnosis to death. XRT indicates radiation therapy.

TABLE 3: Histologic grade and DNA ploidy correlated to  $VD_t$  and tumor volume

Group	Average $VD_t$ , d	Average, in $VD_t$
All patients	110.3 (105)*	$4.7 \pm 0.94$ , n = 25 ( $4.65 \pm 0.82$ , n = 16)*
Grades I-II	145 (108.3)	$4.98 \pm 0.93$ , n = 9 ( $4.69 \pm 0.45$ , n = 4)
Grade III	140 (173.7)	$4.94 \pm 1.25$ , n = 7 ( $5.16 \pm 1.23$ , n = 4)
Grade IV	69.7 (80.4)†	$4.24 \pm 0.77$ , n = 9 ( $4.38 \pm 0.68$ , n = 8)
Diploid and euploid	179 (156)‡	$5.19 \pm 0.59$ , n = 11 ( $5.05 \pm 0.39$ , n = 8)
Aneuploid and multiclonal	44.3 (48.9)	$3.79 \pm 0.60$ , n = 6 ( $3.89 \pm 0.63$ , n = 4)

Group	Average Tumor Volume, $cm^3$	Average No. of Tumor Doublings
At diagnosis		
Supratentorial astrocytomas	20.5 (22.1)*	$35.3 \pm 2.4$ , n = 22 ( $35.4 \pm 2.47$ , n = 16)*
Grades I-II	19.2 (15.6)	$35.2 \pm 2.29$ , n = 8 ( $34.9 \pm 2.26$ , n = 4)
Grade III	19.2 (19.2)	$35.2 \pm 3.48$ , n = 6 ( $35.2 \pm 4.2$ , n = 4)
Grade IV	26.6 (29.1)	$35.7 \pm 1.66$ , n = 8 ( $35.8 \pm 1.71$ , n = 8)
At death		
Supratentorial astrocytomas	181 (165)*	$38.4 \pm 0.87$ , n = 17 ( $38.3 \pm 1.08$ , n = 16)*
Grades I-II	233 (165)	$38.8 \pm 1.31$ , n = 5 ( $38.3 \pm 0.37$ , n = 4)
Grade III	203 (203)	$38.6 \pm 1.34$ , n = 4 ( $38.6 \pm 1.34$ , n = 4)
Grade IV	143.5 (143)	$38.1 \pm 1.26$ , n = 8 ( $38.1 \pm 1.26$ , n = 8)

Note.—Numbers in parentheses are averages of the 16 patients with supratentorial gliomas with both preoperative and antemortem scans.

\* Only 16 patients with supratentorial astrocytomas had scans available after last therapy before death in addition to scans before resection and/or radiation therapy of the primary tumor.

† Not significantly different,  $P = .08$  ( $P < .10$ ) from grades I and II.

‡ Significantly different,  $P < .001$  ( $P < .005$ ) from the aneuploid and multiclonal group of astrocytomas.

tumors tended to have a shorter average  $VD_t$  than grade I and II astrocytomas, but this difference was not significant. In addition, there were no significant differences between the average  $VD_t$  of grade IV tumors compared with grade III tumors or non-grade IV tumors. The two infratentorial tumors in our study demonstrated similar sizes at diagnosis compared with cerebral astrocytomas of approximately 35 tumor doublings. However, the two infratentorial astrocytomas demonstrated a markedly smaller final tumor size at death of 35.3 tumor doublings compared with the final volume of 38.4 tumor doublings for the cerebral tumor group (see Table 3).

#### Histologic Grade and DNA Ploidy

Of the 17 patients with DNA ploidy data, 6 patients (35.3%) had aneuploid or multiclonal tumors, 2 (11.8%) had euploid tumors, and 9 (52.9%) had diploid tumors (see Table 4). Of the seven patients with grade IV tumors, four (57%) had aneuploid or multiclonal tumors with an average  $VD_t$  of 45.1 days, and three (43%) were diploid with an average  $VD_t$  of 151 days. Of the four grade III patients, one (25%) had an aneuploid tumor with a rapid  $VD_t$  of 31 days, one (25%) had a euploid tumor with a relatively slow  $VD_t$  of 291 days, and two (50%) were dip-

loid with an average  $VD_t$  of 126.5 days. Of the six grade I and II patients, one (17%) had a multiclonal tumor with a relatively rapid  $VD_t$  of 58 days, one (17%) had an euploid tumor with an extremely slow  $VD_t$  of 556 days, and four (66%) were diploid with an average  $VD_t$  of 161.6 days.

In the 12 patients in whom one or more biopsies were performed after initial resection, the relationship of DNA ploidy and histology with respect to time was unclear (Table 5). Generally when tumors recurred and were subsequently rebiopsied, they had progressed in histologic grade and in the degree of aneuploidy and multiclonality. One patient (patient 2), who initially had a grade IV tumor with an initially aneuploid DNA histogram, converted to a grade III diploid tumor after resection, radiation therapy, and systemic chemotherapy.

#### Discussion

To gauge therapy and advise patients with intracranial astrocytomas, an accurate measure of prognosis is needed. Histologic grading has not been adequate to determine individual outcome. Our study demonstrates that  $VD_t$  and DNA ploidy may be better prognosticators than histologic grade. The importance of the growth rate of residual or recurrent tumors in patients

TABLE 4: Patients with intracranial astrocytomas

Patient (Age at Diagnosis)	Initial Histology before Therapy	Primary Tumor Location	Survival, mo	Size-adjusted Survival, mo	VD <sub>t</sub> , d	DNA Ploidy
1(58 y)	Grade IV	P	21.6	24.9	22	MC D/T <sub>3</sub> T <sub>4</sub>
2(28 y)	Grade IV (gemistocytic)	P	13.0	19.1	26	A Hypodiploid
3(77 y)	Grade III	T	9	8.3	31	A SubD/necrosis
4(72 y)	Grade IV*	T	6.0	9.9	37	...
5(40 y)	Grade III	P	>18	>20	40	...
6(36 y)	Grade II	F/T/P	61	61+	44	...
7(7 y)	Grade I-II	Pontine	9.7	9.4	58	MC D/T <sub>3</sub> Sub T <sub>3</sub>
8(52 y)	Grade I, oligodendro/astrocytoma	F	31.3	37.6	70	...
9(76 y)	Grade IV	T	2.5	17.5	75	...
10(43 y)	Grade IV	P	14.1	23.2	78	MC D/A/H
11(30 y)	Grade II	F	23	42.6	84	D D/5.72% T <sub>4</sub>
12(8 mo)	Grade III	T/SS	>19.4	>39.5	86	D D/9/10% supra D
13(50 y)	Grade IV	F	15.4	38.4	92	MC D/A/H
14(51 y)	Grade IV	T/P	10.3	29.4	108	D D/11% - A
15(67 y)	Grade I	P/O	17.9	35.5	120	D D/6% supra T <sub>4</sub>
16(61 y)	Grade IV	T	5.8	53.6	175	D D/a few >4N clones
17(71 y)	Grade IV	F/P	21.0	48.0	181	D D/sub T <sub>3</sub>
18(52 y)	Grade III (gemistocytic)	P	163	195	185	D D/10% A
19(62 y)	Grade I	T	8.9	52.3	194	D D
20(32 y)	Grade III	F	62.4	144.9	291	E D/T <sub>4</sub> /6.79% hyper T <sub>4</sub>
21(35 y)	Grade III	Cerebellar	31.3	102	336	...
22(5 mo)	Grade I, pilocytic	SS	>69	>127	349	D D
23(45 y)	Grade I-II (fibrillary)	F	>161	>202.5	415	...
24(23 y)	Grade III	F(Bifrontal)	67	210	545	...
25(11 y)	Grade II	F	>67	>225	556	E E/<3% H

Note.—> indicates patient still alive; +, unknown size of primary tumor; %, percentage of a particular subclone in the total population of nuclei studied; MC, multiclonal tumor; A, aneuploid; D, diploid; T<sub>3</sub>, triploid; T<sub>4</sub>, tetraploid; E, euploid; H, hyperploid; F, frontal; P, parietal; O, occipital; T, temporal; and SS, suprasellar.

\* Probable histology based on radiographic appearance.

with intracranial astrocytomas is underscored by the significance of VD<sub>t</sub> with respect to survival and size-adjusted survival. Furthermore, there seems to be a significant direct relationship between VD<sub>t</sub> and survival as well as size-adjusted survival based on our study. It is important to note, however, that our study dealt with patients who were not cured by primary therapy and who subsequently progressed or developed recurrent disease, the imaging of which allowed for the computation of VD<sub>t</sub>. Therefore, our results cannot necessarily be generalized to all patients with intracranial astrocytomas, particularly the lower-grade astrocytomas, which may be cured by simple excision and postoperative radiation therapy.

Hoshino et al (15) and others (16), although concluding that both the size and the growth rate of astrocytomas were important determinants of survival, thought that radiographically determined VD<sub>t</sub> did not truly represent tumor proliferative potential and, therefore, was not prognostic. Several reasons were provided, one of which was that astrocytomas tend to be

poorly marginated and that the entire extent of a tumor cannot be reliably measured from CT and/or MR. However, others suggest that the determination of the location and volume of gross tumor, however, can be reliably defined by CT and/or MR (25–27). Furthermore, Yamashita and Kuwabara (22) found that the ratio of tumor volumes of gross tumor estimated from CT scanning was consistently similar to the ratio of tumor volumes determined at surgery. Estimation of microscopic infiltration and extension, manifested usually as nonspecific peritumoral white matter edema, however, can be challenging and, therefore, was not included in our volume determinations.

A number of prior studies of radiographically determined VD<sub>t</sub> (of primary and metastatic breast and lung carcinoma) have demonstrated the following: (a) that VD<sub>t</sub> is directly related to survival, time to recurrence, and the percentage of patients with recurrent disease at clinical follow-up; and (b) that VD<sub>t</sub> and survival are distributed in an exponential fashion in a patient population (ie, both VD<sub>t</sub> and survival have log-



TABLE 5: Histology and DNA ploidy in patients with one or more biopsies after initial resection

Patient	Pathologic Specimen	Date	Histology	DNA Ploidy
2	1st resection	12/87	Grade IV	A hypodiploid
	2nd resection	4/88	Grade III	D D
6	1st resection	1981	Grade II	...
	2nd resection	9/85	Grade IV	MC D/T4/A
8	1st resection	1/84	Grade I, oligodendro/astrocytoma	...
	2nd resection	8/85	Grade III	...
10	1st resection	6/86	Grade IV (gemistocytic)	MC D/A/H
	2nd resection	2/87	Grade IV (gemistocytic)	...
11	1st resection	2/89	Grade II	D D/5.72% T <sub>4</sub>
	2nd resection	11/90	Grade IV	MC D/4% A and H
13	1st resection	6/88	Grade IV	MC D/A/H
	2nd resection	4/89	Grade IV	...
16	1st resection	12/86	Grade IV	D D/A few > 4N clones
	2nd resection	3/87	Grade IV	MC D/T <sub>3</sub> /T <sub>4</sub> ? octaploids
20	1st resection	4/85	Grade III	E D/T4/6.79% Hyper T <sub>4</sub>
	2nd resection	7/89	Grade IV	MC D/T <sub>3</sub> /T <sub>4</sub>
21	1st resection	5/87	Grade III	...
	2nd resection	3/89	Grade III	...
22	1st resection	3/87	Grade I, pilocytic	D D
	2nd resection	8/89	Grade I, pilocytic	D D
	3rd resection	2/90	Grade I, pilocytic	...
23	1st resection	9/79	Grades I-II (fibrillary)	...
	2nd resection	8/84	Grade III (fibrillary)	D D/0.75% A
	3rd resection	7/87	Grade III	MC D/4.8% A and H
	4th resection	11/88	Grade III	MC D/T <sub>4</sub> /A
25	1st resection	6/87	Grade II	E E (<3% H)
	2nd resection	6/88	Grade III	MC T <sub>3</sub> /T <sub>4</sub> /small supra T <sub>4</sub> population

Note.—Abbreviations are as in Table 4.

normal frequency distributions) (37–53). Because tumor growth rate is also an exponential function with respect to time, several authors have concluded that survival is primarily dictated by  $VD_t$  in patients in whom therapy is not curative (50, 51). This hypothesis is bolstered by the observation that  $VD_t$  is directly proportional to survival and the time to recurrence as noted in a recent critical review (54). Our current study establishes a link between radiographically determined  $VD_t$  and survival in patients with intracranial astrocytomas.

A point worth emphasizing is that our data both directly and indirectly demonstrate that  $VD_t$  must be relatively constant during the periods of clinical observation. When  $VD_t$  was able to be measured directly, over two or more periods of disease progression, it changed little despite intervals between estimates of  $VD_t$  of 8 or more months during which there was surgical resection and/or radiation therapy. The slope analysis of survival versus  $VD_t$  as depicted in Figure 1 provides an indirect confirmation of the relative constancy of  $VD_t$  over the clinical period of observation. If survival were linearly in-

dependent of  $VD_t$ , then the slope of the best-fit line generated by a linear regression analysis would be zero. However, the slope of survival versus  $VD_t$  is  $6.09 \pm 1.98$  tumor doublings. Stated slightly differently, the slope is greater than 3 SD ( $P = .0011$ ) above zero (ie, no linear association between  $VD_t$  and survival). The slope of adjusted survival,  $13.8 \pm 1.89$  tumor doublings (ie, greater than 6 SD from zero), further underscores the fact that the linear relationship between  $VD_t$  and survival in our study is not coincidental.

The exceptions to the above were patients 6 and 18, who had unusually long survivals. Patient 6 initially had a grade II tumor, which recurred 5 years later as a rapidly enlarging mass ( $VD_t$  of 44 days), of which the patient quickly died. Presumably the recurrent tumor was a distinctly different (ie, dedifferentiated) clone with a rapid  $VD_t$  compared with the original tumor. Patient 18 had a relatively slowly growing grade III astrocytoma ( $VD_t$  of 185 days) who survived for an exceptionally long period. Presumably the initial therapy for this slowly growing tumor was markedly more cytoreductive compared

with other cases in this series. A marked degree of cytoreduction coupled with a relatively slow  $VD_t$  could have accounted for a long survival.

At first glance our analysis seems to ignore the effect of therapy (surgery, radiation therapy, or chemotherapy); however, this is not the case. In fact, the cumulative amount of cytoreduction can be directly inferred when the  $VD_t$ , tumor size at diagnosis and at death, and survival are known. For example, survival, as expressed as the number of tumor doublings, should be equal to the difference between the volume of tumor at death and diagnosis, also expressed as the number of tumor doublings, if no therapy was undertaken. If these quantities are different (ie, the patient survived more tumor doublings than expected), then there must have been a reduction of tumor volume due to therapy in which the patient gained  $x$  number of tumor doubling times ( $x = \text{survival [number of doublings]} - \text{tumor volume at death [number of doublings]} - \text{tumor volume at diagnosis [number of doublings]}$ ). Therefore, the average gain in number of tumor doubling times in our group was three tumor doublings (3.0 doublings = 6.09 doublings [slope] - 38.4 doublings [volume at death] - 35.3 doublings [volume at diagnosis]). This gain is equivalent to a 90% reduction in tumor volume. Because the average  $VD_t$  in our study was 110 days, the therapeutic gain of three doubling times is equal to 1 year of extended survival. The gain, in terms of time, however, would be greater for patients with longer  $VD_t$ s and far less for the rapidly growing (shorter- $VD_t$ ) tumors.

These arguments apply only to cerebral astrocytomas in which mass effect is usually the cause of death. The two infratentorial tumors in our group were similar to each other in size at the time of diagnosis and at the time of death, which may indicate that the invasion of vital brain stem structures by recurrent disease, not gross mass effect, was the cause of death.

In an individual patient, the benefit of a single therapeutic intervention can be estimated from the following example regardless of its anatomic location. For example, if a patient has an intracranial astrocytoma with an initial tumor volume of 30  $VD_t$ s ( $0.523 \text{ cm}^3$ ) growing at a  $VD_t$  of 1 month, which recurs with a volume of 30  $VD_t$ s 12 months after resection, then the reduction of tumor burden must have been equivalent to 12  $VD_t$ s. A 12- $VD_t$  decrease in tumor burden is equivalent to an approximately

0.999% reduction in tumor volume (ie, less than  $0.0005 \text{ cm}^3$  of residual tumor after resection), which would be undetectable when the patient is scanned in the immediate postoperative period.

A surprising result of our study was the relative insensitivity of histologic grade with respect to survival. In a multivariate analysis, histology only became a significant independent predictor of survival if grade IV was compared with non-grade IV tumors. In contrast,  $VD_t$  (a continuous variable) based on a multivariate analysis was a highly significant independent predictor of survival. In addition, histology ceased to be a significant independent predictor as shown by the multivariate analysis of histology, DNA ploidy, and  $VD_t$  with respect to size-adjusted survival. Histologic grade is clearly significant in large registry-based studies (1, 9, 10). Based on our study, however, histology is a relatively insensitive predictor of survival with respect to  $VD_t$  and requires relatively large patient populations to see significant differences between individual grades.

One possible bias not explored in our study was the use of the histologic grade of the primary tumor as the reference diagnosis for each case.  $VD_t$ , on the other hand, was determined from a period(s) of progression of the primary tumor or from the growth of recurrent tumor sometime after initial diagnosis. As stated previously,  $VD_t$  in our patient population was relatively constant from diagnosis to death. In contrast, in the 12 patients with two or more follow-up biopsies, 6 increased in histologic grade, and 1 decreased. Furthermore, in two of the five patients with two or more measurements of  $VD_t$  at different clinical periods (patients 8 and 25), histologic grade increased, whereas  $VD_t$  remained constant. Because of the changes of histologic grade over time and with therapy, it is unlikely that the grade of recurrent disease would be as helpful as  $VD_t$  in the prediction of survival, although this issue was not directly addressed in our study.

The relationship between DNA ploidy and survival has been a subject of controversy in the literature. One large study (10, 11) claims a significantly longer survival in patients with hypertriploid DNA histograms, whereas another large series (9) claims a significantly poorer survival in patients with aneuploid and multiclonal tumors. Our study showed that aneuploid and multiclonal tumors had a significantly

shorter survival than the remainder of the group only in the univariate analysis (see Table 1). An aneuploid and multiclonal DNA histogram was not a significant independent predictor of survival as shown in a multivariate analysis. The differences in the various studies of DNA ploidy may in part be attributable to the lack of correction for the lead time bias because of differing tumor sizes at diagnosis. We, therefore, attempted to decrease lead time bias by the calculation of size-adjusted survival, thereby establishing a baseline tumor volume (a 1-cm-diameter tumor) from which the survival of patients could be more reasonably compared. When size-adjusted survival was used in place of survival, DNA ploidy (diploid and euploid versus aneuploid and multiclonal) became a highly significant independent variable as shown by a multivariate analysis. The identification of patients destined to have poor survival within each histologic grade was also possible through the analysis of DNA ploidy (as well as  $VD_t$ ). Furthermore, we found that aneuploid and multiclonal tumors in our study grew significantly faster than the diploid and euploid tumors. This conceivably could be another confounding factor in correlating DNA ploidy with survival if the issue of lead time bias because of differing tumor sizes at diagnosis is not addressed. The lead time bias in our study ranged from 0 to 12.7 years with an average lead time of 2.3 years.

DNA ploidy and histologic grade, however, progressed with time as seen with multiple sequential biopsies with one exception. The exception may have been attributable to sampling or to radiation therapy and chemotherapy, which can be expected to select against the aneuploid clones of the tumor, leaving the diploid clones as in other malignancies (55). Timing, therefore, seems to be a critical factor in obtaining the most predictive value from estimates of DNA ploidy. Analysis of the primary tumor tissue before therapy correlates best with survival. The frequency of aneuploid tumors also increased with higher histologic grades as noted by Zaprianov and Christov (6), Ahayi (3), and others (7–11). In addition, these authors also noted the same relationships of DNA ploidy, histologic grade, timing of biopsied material, and survival as were found in our study.

With the exception of the study by Zaprianov and Christov (6), another problem with previous studies is that flow cytometry or image cytometry

(after tumor disaggregation and preparation of nuclei) were used to estimate DNA ploidy. When performing flow cytometry, tumor cells are not separated from normal cells, and the estimates of DNA ploidy may, therefore, be unreliable, as noted in two critical reviews (56, 57). A similar problem may also occur when nuclei analyzed by image cytometry are prepared from suspensions (11). In this type of preparation, distinction between normal and pathologic isolated nuclei may be uncertain.

### Conclusions

$VD_t$  and tumor size are important prognostic variables in patients with intracranial astrocytomas who have recurrent or progressive disease.  $VD_t$  demonstrated a significant direct linear relationship to survival as well as size-adjusted survival and, therefore, is generally predictive of an individual astrocytoma patient's prognosis. DNA ploidy also becomes significant when survival is adjusted for tumor size at presentation. Histologic grade is a relatively insensitive predictor of individual survival compared with *in vivo* tumor growth rate ( $VD_t$ ). Based on our results, we recommend that  $VD_t$  and DNA ploidy as well as primary tumor size be further studied to refine current staging as well as the assessment of therapy in patients with intracranial astrocytomas.

### Appendix

The method of Collins et al (29) was used to calculate tumor volume in terms of the number of tumor doublings from a single neoplastic cell origin (with an average diameter of 10  $\mu\text{m}$ ) assuming exponential tumor growth. Briefly, a 1-cm-diameter tumor has a volume of 0.523  $\text{cm}^3$ , which corresponds to  $2^{30}$  cells or 30 tumor doublings. Therefore:

$$\frac{2^{30} \text{ cells}}{0.523 \text{ cm}^3} = \frac{2^x \text{ cells}}{\text{observed tumor volume (cm}^3\text{)}},$$

where  $x$  represents the number of tumor doublings. Solving for  $x$ :

$$1) \quad x = 30 + \frac{\ln(\text{observed tumor volume (cm}^3\text{)})}{\ln 2},$$

where  $\ln$  is a natural logarithm, and  $\ln 2 \approx 0.693$ .

Adjusted survival was calculated using the formula:

$$2) \quad \text{adjusted survival} = (x - 30) \times (VD_t) + \text{observed survival},$$

where  $x$  represents the observed tumor volume at diagnosis expressed as the number of tumor doublings as calculated from equation 1.

Tumor volume at death was determined by first calculating the number of months from the last imaging study until death and dividing by  $VD_t$  (in months). This was then added to the last observed tumor volume (also as expressed as the number of tumor doublings) to obtain the estimated tumor volume at death. The estimated tumor volume at death was then converted into units of cubic centimeters by the following relationship:

$$3) \text{ tumor volume(cm}^3\text{)} = \frac{2^z \text{ cells}}{2^{30} \text{ cells}} \times (0.523 \text{ cm}^3),$$

where  $z$  is the estimated tumor volume at death in terms of the number of tumor doublings.

## Acknowledgments

We thank Jerry Halpern, PhD, for helpful discussions concerning Buckley and James censored regression. We also thank Lela Terrazas for preparation of the tables and figures used in the manuscript.

## References

1. Daumas-Duport C, Scheithaver B, O'Fallon J, Kelly P. Grading of astrocytomas: a simple and reproducible method. *Cancer* 1988; 62:2152-2165
2. Ullen H, Mattsson B, Collins VP. Long-term survival after malignant glioma. *Acta Oncol* 1990;29:875-878
3. Ahyai A. Flow cytometric analysis of cellular DNA content in human astrocytomas and oligodendrogliomas. *Neurosurg Rev* 1988;11:177-187
4. Fitzgibbons PL, Turner RR, Appley AJ, et al. Flow cytometric DNA and nuclear antigen content in astrocytic neoplasms. *Am J Clin Pathol* 1989;88:640-644
5. Cho KS, Nagashima T, Barnwell S, Hoshino T. Flow cytometric determinations of modal DNA populations in relation to proliferative potential of human intracranial neoplasms. *J Neurosurg* 1988;69:588-592
6. Zaprianov Z, Christov K. Histological grading, DNA content, cell proliferation and survival of patients with astroglial tumors. *Cytometry* 1988;9:380-386
7. Coons SW, Davis JR, Way DL. Correlation of DNA content and histology in prognosis of astrocytomas. *Am J Clin Pathol* 1988; 90:289-293
8. Jimenez O, Timms A, Quirke P, et al. Prognosis in malignant glioma: a retrospective study of biopsy specimens by flow cytometry. *Neuropathol Appl Neurobiol* 1989;15:331-338
9. Danova M, Giaretti W, Merlo F, et al. Prognostic significance of nuclear DNA content in human neuroepithelial tumors. *Int J Cancer* 1991;48:663-667
10. Salmon I, Kiss R, Dewitte O, et al. Histopathologic grading and DNA ploidy in relation to survival among 206 adult astrocytic tumor patients. *Cancer* 1992;70:538-546
11. Salmon I, Kiss R. Relationship between proliferative activity and ploidy level in a series of 530 human brain tumors, including astrocytomas, meningiomas, schwannomas, and metastases. *Hum Pathol* 1993;24:329-335
12. Hoshino T. A commentary on the biology and growth kinetics of low-grade and high-grade gliomas. *J Neurosurg* 1984;61: 895-900
13. Bookwalter JW III, Selker RG, Schiffer L, Randall M, Iannuzzi D, Kristofik M. Brain-tumor cell kinetics correlated with survival. *J Neurosurg* 1986;65:95-98
14. Germano IM, Ito M, Cho KG, et al. Correlations of histopathological features and proliferative potential of gliomas. *J Neurosurg* 1989;70:701-706
15. Hoshino T, Prados M, Wilson CB, Cho KG, Lee KS, Davis RL. Prognostic implications of the bromodeoxyuridine labeling index of human gliomas. *J Neurosurg* 1989;71:335-341
16. Labrousse F, Daumas-Duport C, Batorski L, Hoshino T. Histological grading and bromodeoxyuridine labeling index of astrocytomas: comparative study in a series of 60 cases. *J Neurosurg* 1991;75:202-205
17. Ito S, Hoshino T, Shibuya M, et al. Proliferative characteristics of juvenile pilocytic astrocytomas determined by bromodeoxyuridine labeling. *Neurosurgery* 1992;31:413-419
18. Wong AJ, Bigner SH, Bigner DD, et al. Increased expression of the epidermal growth factor receptor gene in malignant gliomas is invariably associated with gene amplification. *Proc Natl Acad Sci USA* 1987;84:6899-6903
19. Diedrich V, Eckerman O, Schmidtke J. Rare Ha-ras and c-mos alleles in patients with intracranial tumors. *Neurology* 1988;38: 587-589
20. Fujimoto M, Weaker FJ, Herbert DC, Sharp ZD, Sheridan PJ, Shorty JL. Expression of three viral oncogenes (*v-sis*, *v-myc*, *v-fos*) in primary human brain tumors of neuroectodermal origin. *Neurology* 1988;38:289-293
21. Saucedo R, Ocadiz R, Gutierrez AL, Salcedo, Ortega V, Figueroa HH. Novel combination of *c-myc*, *N-myc*, and *N-ras* oncogene alternation in brain tumors. *Mol Brain Res* 1988;3:123-132
22. Yamashita T, Kuwabara T. Estimation of rate of growth of malignant brain tumors by computed tomography scanning. *Surg Neurol* 1983;20:464-470
23. Tsuboi K, Yoshii Y, Nakagawa K, Maki Y. Regrowth patterns of supratentorial gliomas: estimation from computed tomographic scans. *Neurosurgery* 1986;19:946-951
24. Teplitz RL, Butler B, Tesluk H, Min B, Russell L, et al. Quantitative DNA patterns in human preneoplastic breast lesions. *Anal Quant Cytol Histol* 1990;12:98-102
25. Galloway RL Jr, Maginnas RJ, Failing AL, et al. Volumetric measurement of canine gliomas using MRI. *Magn Reson Imaging* 1990;8:161-165
26. Myrianthopoulos LC, Vijayakumar S, Spelbring DR, et al. Quantitation of treatment volumes from CT and MRI in high-grade gliomas: implications for radiotherapy. *Magn Reson Imaging* 1992;10:375-383
27. TenHaken RK, Thornton AF, Sandler HM, et al. A quantitative assessment of the addition of MRI to CT-based, 3-D treatment planning of brain tumors. *Radiother Oncol* 1992;25:121-133
28. Breiman RS, Beck JW, Korobkin M, et al. Volume determinations using computed tomography. *AJR Am J Roentgenol* 1982;138: 329-333
29. Collins VP, Loeffler K, Tivey H. Observation on growth rates of human tumors. *AJR Am J Roentgenol* 1956;76:988-1000
30. Israel L, Chahinian P, Accard J, et al. Growth curve modification of measurable tumors by 75 mg/m<sup>2</sup> of CCNU every three weeks. *Eur J Cancer* 1973;9:789-797
31. Israel L, Chahinian P. Evaluation of the survival gain in 22 measurable lung tumors treated with chemotherapy. *Eur J Cancer* 1969;5:631-637

32. Van Peperziel HA, Breur K, Broerse J, Barandsen G. RBE values of 15 MeV neutrons for response of pulmonary metastases in patients. *Eur J Cancer* 1974;10:349-355
33. Alvord EC Jr. Why do gliomas not metastasize? *Arch Neurology* 1976;33:73-75
34. Spratt JS. The lognormal frequency distribution and human cancer. *J Surg Res* 1969;9:151-157
35. Spratt JS, Spratt JA. Growth rates. In: Donegan WL, Spratt JS, eds. *Cancer of the Breast*. 3rd ed. Philadelphia: WB Saunders, 1988:270-302
36. Buckley J, James I. Linear regression with censored data. *Biometrika* 1979;66:429-436
37. Philippe E, Le Gal Y. Growth of seventy-eight recurrent mammary cancers. Quantitative study. *Cancer* 1968;21:461-467
38. Lee YTN, Spratt JR. Rate of growth of soft tissue metastases of breast cancer. *Cancer* 1972;29:344-348
39. Kusama S, Spratt JS, Donegan WL, Watson FR, Cunningham C. The gross rates of growth of human mammary carcinoma. *Cancer* 1972;30:594-599
40. Pearlman AW. Breast cancer: influence of growth rate on prognosis and treatment evaluation. *Cancer* 1976;38:1826-1833
41. Heuser L, Spratt JS, Polk HC. Growth rates of primary breast cancers. *Cancer* 1979;43:1888-1894
42. Fournier DV, Schiller V, Junkermann H, Legler V, Baver M. Natural growth rate in 300 primary breast carcinomas need correlation to hormone factors. *Ann NY Acad Sci* 1986;464:563-556
43. Galante E, Gallus G, Guzzon A, Bono A, Bandieramonte G, DiPietro S. Growth rate of primary breast cancer and prognosis: observations on a 3 to 7 year follow-up in 180 breast cancers. *Br J Cancer* 1986;54:833-836
44. Spratt JS, Spratt JA. Growth rates and the cytokinetic behavior of breast cancer. In: Kubli F, Fournier DV, et al. *Breast Diseases*. Berlin: Springer-Verlag, 1989:97-110
45. Kuroishi T, Tominga S, Morimoto T, et al. Tumor growth rate and prognosis of breast cancer mainly detected by mass screening. *Jpn J Cancer Res* 1990;81:454-462
46. Spratt JS, Spratt TS. Rates of growth of pulmonary metastases and host survival. *Ann Surg* 1964;159:161-171
47. Joseph WL, Morton DL, Adkins PC. Prognostic significance of tumor doubling time in evaluating operability in pulmonary metastatic disease. *J Thorac Cardiovasc Surg* 1971;61:23-31
48. Weiss W. Tumor doubling time and survival of men with bronchogenic carcinoma. *Chest* 1974;65:3-8
49. Band PR, Kocandrlc C. Growth rate of pulmonary metastases in human sarcomas. *Cancer* 1975;36:471-474
50. Boag JW. Maximum likelihood estimates of the proportion of patients cured by cancer therapy. *J R Stat Soc* 1949;11:15-53
51. Rutquist LE, Wallgren A. Long term survival of 458 young breast cancer patients. *Cancer* 1985;55:658-665
52. Berg JW, Robbins GF. Factors influencing short and long term survival of breast cancer patients. *Surg Gynecol J Obstet* 1966;122:1311-1316
53. Fournier DV, Weber E, Hoeffken N, Baver M, Kubli F, Barth V. Growth rate of 147 mammary carcinomas. *Cancer* 1980;45:2198-2207
54. Plotkin D, Blankenberg F. Breast cancer—biology and malpractice. *Am J Clin Oncol* 1991;14:254-266
55. Dobashi K, Stratton JA, Teplitz RL, Liao SY, Braly PS, Disma PJ. Quantitative nuclear DNA analysis of human ovarian adenocarcinoma: compared before and after chemotherapy and correlated with clinical response. *Gynecol Oncol* 1986;24:81-90
56. Teplitz RL, Hammond W. Measurement of total DNA to assess lung cancer. *Ann Thorac Surg* 1993;55:576-577
57. Wersto RP, Liblit RL, Koss LG. Flow cytometric analysis of human solid tumors. *Hum Pathol* 1991;22:1095-1098

Please see the commentary on page 1013 in this issue.