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**ORIGINAL
RESEARCH**

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Malignant Tumors and Chronic Infections in the Masticator Space: Preliminary Assessment with In Vivo Single-Voxel ^1H -MR Spectroscopy

BACKGROUND AND PURPOSE: Differential diagnosis between malignant tumors and chronic infections in the masticator space remains challenging. The purpose of the study was to categorize various changes of both lesions by using single-voxel ^1H -MR spectroscopy.

MATERIALS AND METHODS: Twenty-four masticator space lesions, 7 chronic infections, and 17 malignant tumors were assessed with ^1H -MR spectroscopy before treatment procedures. The MR spectroscopic findings were compared with surgical and histopathologic results. Localization technique for ^1H -MR spectroscopy was made by a point-resolved spectroscopy sequence at an echo time of 144 ms. Choline (Cho) signals (identified at 3.2 ppm) and Cho signal intensity-to-noise ratio (Cho/noise) were considered as evaluating criteria.

RESULTS: All the lesions were found with 3 ^1H -MR spectroscopic types: type 1, lesions without Cho signals (3 chronic infections); type 2, lesions with Cho signals and Cho/noise ratio <3 (4 chronic infections and 4 malignant tumors); and type 3, lesions with Cho signals and Cho/noise ratio >3 (13 malignant tumors). The mean \pm SD of the Cho/noise ratio between chronic infections and malignant tumors was 2.31 ± 0.19 and 5.76 ± 3.29 ($P < .01$), respectively.

CONCLUSION: In vivo single-voxel ^1H -MR spectroscopy may be helpful in the assessment of masticator space lesions. Differences of Cho signals and Cho/noise ratios between malignant tumors and chronic infections provide valuable information in the differentiation of these 2 lesions.

Differentiation of chronic infections versus malignant tumors in the masticator space with CT and MR imaging is often difficult. To make an accurate diagnosis, one needs to explore new imaging techniques to analyze possible chemical or biochemical components of the lesions. Although MR spectroscopy was initially described with potential uses for head and neck lesions more than 20 years ago,¹⁻⁴ it was not until recent years that the technology was widely used clinically.⁵ With rapid development and improvement of hardware and software, in vivo point-resolved MR spectroscopy (^1H -MR spectroscopy) has started to be used in the evaluation of extracranial head and neck lesions of the tongue,⁶⁻⁷ nasopharynx,⁸ parotid gland,⁹ thyroid,¹⁰ and metastatic lymph nodes of the neck.^{6-8,10-12} In addition, the technology was used to detect recurrences of malignant tumors and to monitor treated or untreated tumors.^{5,11-13} However, to our knowledge, the ^1H -MR spectroscopic appearance of chronic infections and malignant tumors in the masticator space has not been reported.

As one of the latest techniques for imaging analysis, single-voxel ^1H -MR spectroscopy measures living tissue metabolism at the molecular level. Increased choline (Cho) signals were generally thought to be an important criterion in assessing various tumors. However, whether the Cho metabolites could play a role in the differentiation of chronic infections and ma-

lignant tumors in the masticator space was unknown. The aims of this preliminary study were to evaluate in vivo the single-voxel ^1H -MR spectroscopic appearance of chronic infections and malignant tumors in the masticator space and to determine if differences exist between both lesions in terms of the levels of Cho metabolites.

Materials and Methods

Subjects

From 2003 to 2006, 47 patients with lesions in the masticator spaces were referred to the Department of Radiology. MR imaging and ^1H -MR spectroscopy were performed for all patients. Three of the studies were unsuccessful because of uninterpretable spectra and unacceptable line width at half maximum of the unsuppressed water peak. Among the remaining 44 lesions, 24 were confirmed histopathologically to be either inflammatory lesions or malignant tumors. These 24 patients consisted of 18 men and 6 women, ranging in age from 3 to 82 years, with a mean age of 40.6 years. Pathologic outcomes of 24 lesions included 7 chronic infections, 4 squamous cell carcinomas, 7 salivary carcinomas, and 6 sarcomas. The demographic distribution of the subjects and lesions is listed in the Table.

MR Imaging and MR Spectroscopy

MR imaging and ^1H -MR spectroscopy were performed on a 1.5T Signa TwinSpeed system (GE Healthcare, Milwaukee Wis). A neurovascular array coil or a quadrature transmit/receive head coil was used. Axial MR imaging was performed by using T1-weighted spin-echo and T2-weighted fast spin-echo sequences. Coronal MR imaging was performed by using a T2-weighted fast spin-echo sequence with fat suppression. The parameters of the T1-weighted spin-echo sequence were listed as follows: TR/TE, 600/11 ms; 3 acquisitions; FOV, 24 cm; bandwidth, 31.25 Hz; matrix, 256 \times 192; section thickness, 5 mm; spacing, 1 mm. The parameters of the T2-weighted fast spin-

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Single-voxel ^1H -MR spectroscopic findings of chronic infections and malignant tumors in the masticator space						
Case No./Sex/Age	Origin	Pathology	VOI (cm^3)	Cho Signal	Cho	Signal-to-Noise Ratio
1/F/29	Unknown	Chronic infection	8.00	—	N/A	
2/M/42	Unknown	Chronic infection	8.39	—	N/A	
3/M/26	Odontogenic	Chronic infection	11.73	+	2.50	
4/M/16	Mandible	Chronic infection	13.29	—	N/A	
5/F/71	Mandible	Chronic infection	6.62	+	2.46	
6/M/3	Odontogenic	Chronic infection	10.48	+	2.13	
7/F/11	Mandible	Chronic infection	5.52	+	2.15	
8/M/49	Unknown	Adenoid cystic carcinoma	8.00	+	6.49	
9/F/49	Maxillary sinus	Adenoid cystic carcinoma	6.74	+	2.50	
10/M/39	Maxillary sinus	Adenoid cystic carcinoma	6.11	+	4.88	
11/M/22	Unknown	Adenoid cystic carcinoma	7.74	+	11.84	
12/M/70	Unknown	Adenocarcinoma	12.48	+	3.47	
13/M/82	Metastasis	Adenocarcinoma	4.17	+	3.40	
14/F/49	Parotid gland	Mucop epithelial carcinoma	10.11	+	7.20	
15/M/29	Parotid gland	Rhabdomyosarcoma	18.35	+	11.08	
16/M/33	Unknown	Rhabdomyosarcoma	5.12	+	4.95	
17/M/57	Unknown	Fibrosarcoma	7.83	+	2.22	
18/M/46	Unknown	Myxofibrosarcoma	7.63	+	3.96	
19/M/32	Mandible	Malignant fibrous histiocytoma	7.99	+	2.30	
20/M/20	Unknown	Synovial sarcoma	7.76	+	10.25	
21/F/34	Unknown	Squamous cell carcinoma	8.00	+	10.64	
22/M/51	Oropharynx	Squamous cell carcinoma	10.58	+	5.44	
23/M/49	Maxillary sinus	Squamous cell carcinoma	17.62	+	2.74	
24/M/70	Soft palate	Squamous cell carcinoma	7.88	+	4.56	

Note:—N/A indicates not available for measurement; VOI, volume of interest.

echo sequence were the following: axial TR/TE, 4700/85 ms and coronal TR/TE, 3200/100 ms; 3 acquisitions; echo-train length, 15; FOV, 24 cm (axial) and 20 cm (coronal); bandwidth, 19.23 Hz (axial) and 25 Hz (coronal); matrix, 256 \times 192 (axial) and 320 \times 160 (coronal); section thickness, 5 mm; spacing, 1 mm.

For lesion localization of ^1H -MR spectroscopy, an axial MR imaging with a T2-weighted spin-echo sequence (TR/TE, 3780/80 ms) or T1 fluid-attenuated inversion recovery (TR/TE/TI, 2130/12/750 ms) was performed with a matrix of 256 \times 160, FOV of 24 cm, and a section thickness of 5 mm with no intersection gap. A point-resolved spectroscopy sequence (PRESS, TR/TE/NEX, 1500/144/8; 64 signal-intensity acquisitions) was used as the localizing technique. On the basis of the size of each lesion, the volume of interest (VOI) was adjusted to cover the depicted lesion area shown on the localizer images. The mean volume for all the lesions in this series was 9.09 cm^3 with a range of 4.17–18.35 cm^3 .

The prescanning for ^1H -MR spectroscopy automatically adjusted the received gains, the transmit gain, the center frequency, a 3-plane autoshim for prescribed VOI, and the water-suppression pulse. The full width at half maximum of the unsuppressed water peak in the PRESS sequence was <17 Hz. After the acquisition of MR spectra, all of the data were automatically processed by the PROBE/SV (Single-voxel Proton Brain Examination, an image-guided clinical spectroscopy package provided by GE Healthcare, Milwaukee, Wis). The procedures of PROBE/SV include internal water-referenced phase-correction, apodization with a gaussian filter (2.5-Hz width), 1-time zero-filling, fast Fourier transformation, baseline correction, registration of metabolic resonance peaks (Cho at 3.2 ppm and creatine [Cr] at 3.0 ppm) and root-mean-square (RMS) noise, and integration of metabolite intensity based on the peak area. The RMS noise was measured by computing the value over two 300-Hz-wide regions of the spectrum (the range of frequencies [−570.0 Hz, −275.0 Hz] and [365.0 Hz, 665.0 Hz]) and taking the greater of the 2 levels. Subsequently, Cho signal intensity-to-noise ratio (Cho/noise) was mea-

sured by a radiologist. The overall time of the examination was approximately 9 minutes, including neoplasm-localization sequences (4 minutes), prescanning (1 minute), and acquisition of PRESS sequence (TE of 144 ms) and MR spectrum (4 minutes).

Criteria and Statistical Analysis

Cho signals and Cho/noise were regarded as the criteria in this study to determine if there were differences between chronic infections and malignant tumors. The Wilcoxon sign test, a nonparametric test, was used to compare differences of Cho/noise ratio between chronic infections and malignant tumors. A *P* value $<.05$ was considered as a statistically significant difference.

Results

Of 24 masticator space lesions, 21 lesions (4 chronic infections and 17 malignant tumors) were identified with Cho signals (Figs 1 and 2), and the remaining 3 chronic infections had no Cho signals (Fig 3). Cho/noise ratios were measured in these 21 lesions. The detailed distribution of Cho signals and Cho/noise ratio data for each lesion is listed in the Table. In addition, creatine (Cr) signals were found in 6 of 24 lesions (25%), including 5 malignant tumors (Fig 2) and 1 chronic infection.

All the lesions could be classified into 3 types on the basis of Cho signals and Cho/noise ratios: type 1 (Fig 3), lesions with absence of Cho signals (3 chronic infections); type 2, lesions with Cho signals and a Cho/noise ratio <3 (4 chronic infections and 4 malignant tumors); and type 3 (Figs 1 and 2), lesions with Cho signals and a Cho/noise ratio >3 (13 malignant tumors). If type 3 lesions were considered as malignant tumors, the true-positive detection rate of malignant tumors was 13 of 17; the true-negative rate, 7 of 7; the false-positive lesion, none of 7; and the false-negative rate, 4 of 17. Therefore, in vivo single-voxel ^1H -MR spectroscopy had a sensitivity of 76.5%, specificity of 100%, positive predictive value of

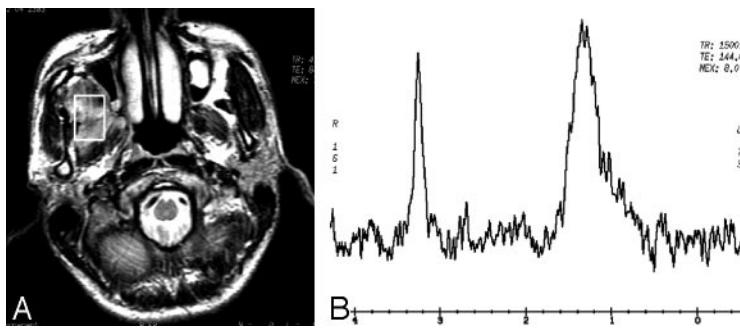


Fig 1. Case 11 with adenoid cystic carcinoma in the right masticator space. Axial T2-weighted image (*A*) demonstrates that the lesion with high signal intensity is located in the right medial masticator space. The MR spectrum (*B*) shows a distinct Cho peak at 3.2 ppm. The lipid signals are visible at 0.9–1.5 ppm.

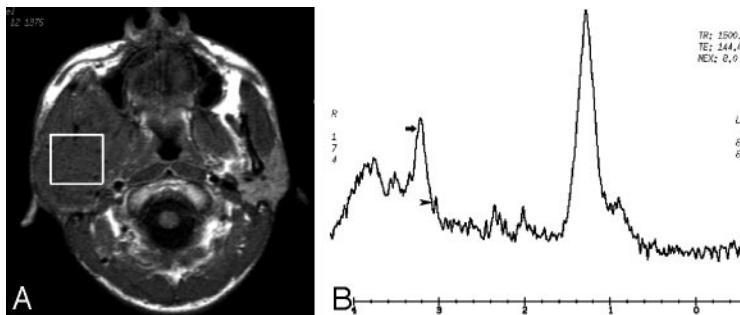


Fig 2. Case 15 with rhabdomyosarcoma in the right parotid gland and masticator space. Axial T1-weighted image (*A*) demonstrates the lesion with intermediate signal intensity in the right parotid gland and masticator space. On the MR spectrum (*B*), both Cho peak (arrow) and Cr peak (arrowhead) present at 3.2 and 3.0 ppm, respectively.

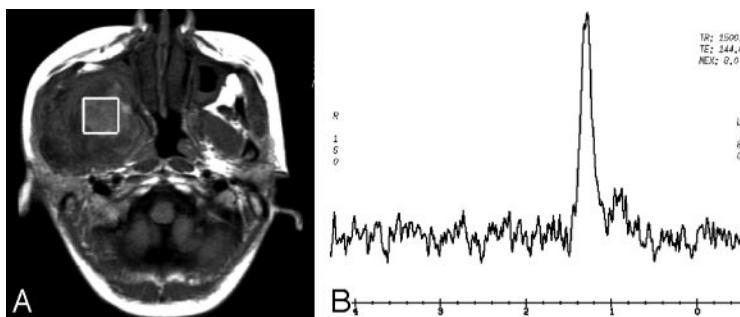


Fig 3. Case 1 with a chronic inflammatory lesion in the right masticator space. Axial T1-weighted image (*A*) demonstrates that the lesion with iso-intense signal intensity is located in the right medial masticator space. The box of VOI is positioned in the lesion of the right medial masticator space, and there are no Cho signals present on the spectrum (*B*). The lipid signals are shown at 0.9–1.5 ppm.

100%, negative predictive value of 63.6%, and accuracy of 83.3%.

The mean \pm SD of Cho/noise ratios for chronic infections and malignant tumors was 2.31 ± 0.19 and 5.76 ± 3.29 , respectively. Statistically, there was a significant difference between both lesions ($P = .009$).

Discussion

An early and correct differentiated diagnosis between an infection and a malignant tumor in the masticator space is pivotal for evaluating the prognosis of a patient. However, some chronic infections or inflammatory pseudotumors in the masticator spaces may be without any acute inflammatory signs, such as facial pain and swelling and pus formation, and are similar to the malignant tumors clinically. The inflammatory lesions in the extraorbital head and neck spaces can also mimic the malignant tumors on conventional CT and MR imaging.^{14–15} These lead to difficulty in indiscriminating between both kinds of lesions.

In vivo single-voxel ^1H -MR spectroscopy is one of the latest techniques for analyzing molecular structures of living organisms. The height of Cho peaks shown on in vivo single-voxel ^1H -MR spectroscopic images is believed to be related to increased cellular membrane phospholipid biosynthesis and is, therefore, considered as an active marker for cellular proliferation.¹⁶ A number of studies with in vivo ^1H -MR spectroscopy

have shown that Cho signals could be reliably found in most malignant tumors of the extracranial head and neck, such as squamous cell carcinomas,^{6–8,11–13,17} salivary carcinomas,^{9,12} and extracranial malignant lymphomas.^{11,18} Soft-tissue and bone sarcomas (outside the head and neck) marked with Cho signals have also been reported.¹⁹ However, there were few studies describing the in vivo ^1H -MR spectroscopic appearances of inflammatory lesions. Shah et al⁵ reported that ^1H -MR spectroscopy could be used to differentiate recurrent head and neck tumors from posttreatment changes. The inflammatory responses with formation of granulation tissue in the tumor bed might result from radiation therapy. The reactive changes after radiation therapy had a low Cho/Cr ratio or absence of Cho and its derivatives.⁵ The current study also showed that the chronic infections in the masticator space had either absence of Cho signals or low Cho/noise ratios. On the contrary, all malignant lesions in the space presented with Cho signals and most of them with high Cho/noise ratios. The different MR spectroscopic appearances between the chronic infections and malignant tumors might be due to the fact that there were hypervascularity, hypercellularity, and higher levels of Cho metabolites and its derivatives.

In the current study, Cho/noise ratio was used to evaluate masticator space lesions. Previous reports^{20,21} indicated that if the Cho/noise ratio was ≥ 2 , the lesion was most likely a malignant tumor. The data of this study demonstrated similar

results in the masticator space. The chronic infections were either without Cho signals or had a Cho/noise ratio <3, whereas most of malignant tumors had a Cho signal intensity and a Cho/noise ratio >3. Also, there was a statistically significant difference between the mean Cho/noise ratios of chronic inflammatory lesions and malignant tumors. On the basis of these differences, *in vivo* single-voxel ¹H-MR spectroscopy might be useful in differentiating chronic infections from malignant tumors in the masticator space.

On the basis of the literature review, the most common semiquantitative criterion used for evaluation of extracranial head and neck neoplasms with ¹H-MR spectroscopy was the Cho/Cr ratio.^{6–13,17} However, some studies found that the Cho/Cr ratio could only be measured in half or less of extracranial head and neck neoplasms.^{8–11} In our series, the peaks of Cr metabolites could only be identified in approximately 25% of masticator space lesions. This might be due to the fact that Cr peaks were at 3.0 ppm and could not be separated from the background noise. The difficulty in detecting Cr signals made the criterion of Cho/Cr ratio less useful in the assessment of masticator space lesions. The Cho/noise ratio has previously been applied to evaluate malignant tumors elsewhere in the body.^{20,21} Furthermore, we found that it was easier to measure the Cho/noise ratio than the Cho/Cr ratio in the masticator space lesions. The RMS noise can be automatically measured by the MR imaging scanner with the same algorithm and method during each acquisition of the MR spectrum.

The drawbacks with *in vivo* single-voxel ¹H-MR spectroscopy in masticator space lesions should be pointed out. First, shimming difficulty existed in this study due to considerable magnetic susceptibility differences in the extracranial head and neck.^{6,11} Unavoidable inclusion of adjacent bony and air-containing structures in the large VOI reduced the likelihood of obtaining an adequate shim, which resulted in decreased spectral resolution.⁶ Second, there was a 23.5% (4/17) false-negative rate for malignant tumors in the current study. This indicated that there was overlap of the MR spectroscopic findings between chronic infections and malignant tumors with the current standards. Third, the VOI used in the current study was >4 cm³, which limited its use for small lesions. Using a VOI <4 cm³ in the masticator space may result in failure of *in vivo* single-voxel ¹H-MR spectroscopy. Therefore, other physiologic techniques, such as MR diffusion-weighted imaging and dynamic enhanced MR imaging, might have potential in the assessment of small masticator space lesions. Finally, because the current study was preliminary and the sample size for both lesions was relatively small, further studies with larger sample sizes are required to validate these findings.

Conclusion

The current study demonstrated that there were 3 different types of ¹H-MR spectroscopic manifestations in both masticator space lesions. The differences of Cho signals and Cho/noise ratios among these 3 manifestations might be potentially useful in the differentiation of chronic infections from malignant tumors in the masticator space.

References

- Mafee MF, Barany M, Gotsis ED, et al. Potential use of *in vivo* proton spectroscopy for head and neck lesions. *Radiol Clin North Am* 1989;27:243–54
- Dawson DE, Sekhar V, Pearson G, et al. Nuclear magnetic resonance spectroscopy of plasma for the detection of head and neck cancer. *Am J Otolaryngol* 1989;10:244–49
- Scher RL, Ropka ME, Neal DA, et al. NMR spectroscopy evaluation of plasma “oncolipids” in head and neck cancer. *Otolaryngol Head Neck Surg* 1990;102:34–40
- Chow JM, Gotsis ED, Jarc M, et al. Proton nuclear magnetic resonance spectroscopy of plasma lipoproteins in head and neck cancer patients. *Am J Otolaryngol* 1990;11:332–38
- Shah GV, Gandhi D, Mukherji SK. Magnetic resonance spectroscopy of head and neck neoplasms. *Top Magn Reson Imaging* 2004;15:87–94
- Mukherji SK, Schiro S, Castillo M, et al. Proton MR spectroscopy of squamous cell carcinoma of the extracranial head and neck: *in vitro* and *in vivo* studies. *AJNR Am J Neuroradiol* 1997;18:1057–72
- Star-Lack JM, Adalsteinsson E, Adam MF, et al. *In vivo* ¹H-MR spectroscopy of human head and neck lymph node metastasis and comparison with oxygen tension measurements. *AJNR Am J Neuroradiol* 2000;21:183–93
- King AD, Yeung DKW, Ahuja AT, et al. *In vivo* proton MR spectroscopy of primary and nodal nasopharyngeal carcinoma. *AJNR Am J Neuroradiol* 2004;25:484–90
- King AD, Yeung DKW, Ahuja AT, et al. Salivary gland tumors at *in vivo* proton MR spectroscopy. *Radiology* 2005;237:563–69
- King AD, Yeung DKW, Ahuja AT, et al. *In vivo* ¹H MR spectroscopy of thyroid carcinoma. *Eur J Radiol* 2004;54:112–17
- King AD, Yeung DKW, Ahuja AT, et al. Human cervical lymphadenopathy: evaluation with *in vivo* ¹H-MRS at 1.5 T. *Clinical Radiol* 2005;60:592–98
- Bisdas S, Baghi M, Huebner F, et al. *In vivo* proton MR spectroscopy of primary tumours, nodal and recurrent disease of the extracranial head and neck. *Eur Radiol* 2007;17:251–57
- Bezabeh T, Odulum O, Nason R, et al. Prediction of treatment response in head and neck cancer by magnetic resonance spectroscopy. *AJNR Am J Neuroradiol* 2005;26:2108–13
- Som PM, Curtin HD. Parapharyngeal and masticator space lesions. In: Som PM, Curtin HD, eds. *Head and Neck Imaging*. St Louis: Mosby; 2003:1996
- Vuyser SD, Hermans R, Sciot R, et al. Extraorbital inflammatory pseudotumor of the head and neck: CT and MR findings in three patients. *AJNR Am J Neuroradiol* 1999;20:1133–39
- Rulz-Cabello J, Cohen JS. Phospholipid metabolites as indicators of cancer cell function. *NMR Biomed* 1992;5:226–33
- Maheshwari SR, Mukherji SK, Neelon B, et al. The choline/creatinine ratio in five benign neoplasms: comparison with squamous cell carcinoma by use of *in vitro* MR spectroscopy. *AJNR Am J Neuroradiol* 2000;21:1930–35
- Schwarz AJ, Maisey NR, Collins DJ, et al. Early *in vivo* detection of metabolic response: a pilot study of ¹H MR spectroscopy in extracranial lymphoma and germ cell tumors. *Br J Radiol* 2002;75:956–66
- Wang CK, Li CW, Hsieh TJ, et al. Characterization of bone and soft-tissue tumors with *in vivo* ¹H MR spectroscopy: initial results. *Radiology* 2004;232:599–605
- Bartella L, Morris EA, Dershaw DD, et al. Proton MR spectroscopy with choline peak as malignant marker improves positive predictive value for breast cancer diagnosis: preliminary results. *Radiology* 2006;239:686–92. Epub 2006 Apr 7
- Lee J, Yamaguchi T, Abe A, et al. Clinical evaluation of choline measurement by proton MR spectroscopy in patients with malignant tumors. *Radiat Med* 2004;22:148–54