

Apolipoprotein E ϵ 4 Does Not Modulate Amyloid- β Associated Neurodegeneration in Preclinical Alzheimer Disease

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ABSTRACT

BACKGROUND AND PURPOSE: Among cognitively healthy older individuals, the relationship among the 2 hallmark proteins of AD ($A\beta$ and τ *APOE* ϵ 4) and neurodegeneration is not well-understood. Here, we investigated the relationship between $A\beta$, p- τ , and *APOE* ϵ 4 on longitudinal brain atrophy in preclinical AD.

MATERIALS AND METHODS: We examined 107 cognitively healthy older adults who underwent longitudinal MR imaging and baseline lumbar puncture. Within the same linear mixed-effects model, we concurrently investigated main and interactive effects between the *APOE* ϵ 4 genotype and CSF $A\beta_{1-42}$, CSF p- τ and CSF $A\beta_{1-42}$, and the *APOE* ϵ 4 genotype and CSF p- τ on entorhinal cortex atrophy rate. We also examined the relationship of *APOE* ϵ 4, CSF p- τ , and CSF $A\beta_{1-42}$ on the atrophy rate of other AD-vulnerable neuroanatomic regions.

RESULTS: The full model with main and interactive effects demonstrated a significant interaction only between CSF p- τ and CSF $A\beta_{1-42}$ on entorhinal cortex atrophy rate, indicating elevated atrophy with time in individuals with increased CSF p- τ and decreased CSF $A\beta_{1-42}$. The *APOE* ϵ 4 genotype was significantly and specifically associated with CSF $A\beta_{1-42}$. However, the interaction between the *APOE* ϵ 4 genotype and either CSF $A\beta_{1-42}$ or CSF p- τ on entorhinal cortex atrophy rate was not significant. We found similar results in other AD-vulnerable regions.

CONCLUSIONS: On the basis of our findings and building on prior experimental evidence, we propose a model of the pathogenic cascade underlying preclinical AD in which *APOE* ϵ 4 primarily influences the pathology of Alzheimer disease via $A\beta$ -related mechanisms, and in turn, $A\beta$ -associated neurodegeneration occurs only in the presence of p- τ .

ABBREVIATIONS: $A\beta$ = amyloid- β ; AD = Alzheimer disease; *APOE* ϵ 4 = ϵ 4 allele of *apolipoprotein E*; HC = healthy controls; p- τ = phospho- τ_{181p} ; SE = standard error of the mean

Converging biochemical, molecular, and genetic evidence indicates that $A\beta$ plays a central role in the neurodegenerative process underlying AD.¹ The presence of $A\beta$ initiates loss of den-

dritic spines and synapses² and contributes to the dysfunction of neuronal networks.³ Reports based on mouse models suggest that multiple factors influence $A\beta$ -associated toxicity. The ϵ 4 allele of

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Data used in preparation of this article were obtained from the ADNI data base (<http://adni.loni.ucla.edu>). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in the analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.ucla.edu/wpcontent/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf.

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Table 1: Demographic, clinical, and imaging data for all older HC in this study, as assessed by P- τ and A β status

	P- τ -/A β - (n = 46)	P- τ -/A β + (n = 20)	P- τ +/A β - (n = 19)	P- τ +/A β + (n = 21)	P Value
Age (yr) (mean) (SE)	74.3 (0.6)	74.9 (1.1)	78.0 (1.4)	78.2 (1.0)	.02 ^a
Female (%)	24	31	29	38	.59 ^b
Education (yr) (mean) (SE)	15.5 (0.4)	14.8 (0.8)	15.5 (0.4)	16.7 (0.6)	.34 ^a
Baseline MMSE (mean) (SE)	29.1 (0.1)	29.1 (0.2)	28.8 (0.3)	29.3 (0.2)	.46 ^a
Entorhinal cortex APC (mean) (SE)	-0.6 (0.15)	-0.6 (0.18)	-0.6 (0.18)	-1.2 (0.25)	.005 ^c
AD-vulnerable ROI APC (mean) (SE)	-0.6 (0.08)	-0.5 (0.11)	-0.7 (0.14)	-1.1 (0.14)	.002 ^c

Note:—MMSE indicates Mini-Mental State Examination; APC = annualized percentage change.

^a Derived from analysis of variance.

^b Derived from a χ^2 test.

^c Derived from linear mixed-effects models (please see text for details).

Table 2: Demographic, clinical, and imaging data for all older HC in this study, as assessed by APOE ϵ 4 and A β status

	ϵ 4-/A β - (n = 61)	ϵ 4-/A β + (n = 21)	ϵ 4+/A β - (n = 5)	ϵ 4+/A β + (n = 20)	P Value
Age (yr) (mean) (SE)	75.7 (0.7)	76.2 (0.9)	71.7 (2.5)	77.1 (1.3)	.56 ^a
Female (%)	54	54	20	35	.23 ^b
Education (yr) (mean) (SE)	15.6 (0.3)	15.9 (0.6)	15 (1.1)	15.6 (0.8)	.98 ^a
Baseline MMSE (mean) (SE)	29.1 (0.1)	29.4 (0.2)	28.6 (0.9)	29 (0.2)	.73 ^a
Entorhinal cortex APC (mean) (SE)	-0.57 (0.13)	-0.67 (0.17)	-0.43 (0.30)	-1.17 (0.28)	.35 ^c
AD-vulnerable ROI APC (mean) (SE)	-0.6 (0.07)	-0.78 (0.14)	-0.65 (0.23)	-1.0 (0.16)	.28 ^c

Note:—MMSE indicates Mini-Mental State Examination; APC = annualized percentage change.

^a Derived from analysis of variance.

^b Derived from a χ^2 test.

^c Derived from linear mixed-effects models (please see text for details).

APOE ϵ 4, the most important genetic risk factor for late-onset AD,⁴ accelerates the onset of A β deposition into plaques⁵ and decreases the transport of A β across the blood-brain barrier.⁶ Reductions in τ , another hallmark protein of AD pathology, protect against A β -induced neuronal dysfunction,⁷ while the presence of τ potentiates A β -associated synaptotoxicity.⁸

In humans, evidence from genetic-at-risk cohorts and neuropathologic findings in clinically healthy older individuals suggest that the pathobiologic process underlying AD begins years before the onset of cognitive deficits or dementia symptoms.⁹ Biomarker studies in cognitively asymptomatic older adults have demonstrated significant relationships between structural MR imaging measures of brain atrophy and CSF A β levels,¹⁰⁻¹² enabling identification of clinically healthy individuals who may be in a presymptomatic or preclinical stage of AD.¹³

Recent evidence from our laboratory indicates that in clinically healthy older individuals and those with mild cognitive impairment, A β -associated volume loss occurs only in the presence of p- τ .¹⁴ However, it is unknown whether APOE ϵ 4 and CSF p- τ concurrently modulate the effect of CSF A β on longitudinal brain atrophy in preclinical AD. In this study, we investigated whether concurrent interactions between decreased CSF A β ₁₋₄₂ and APOE ϵ 4 and between decreased CSF A β ₁₋₄₂ and increased CSF p- τ are associated with increased brain atrophy in cognitively healthy older individuals.

MATERIALS AND METHODS

Selection of participants and analysis methods for MR imaging and CSF biomarkers are briefly summarized here, with details provided in the On-line Appendix.

We evaluated participants who were clinically diagnosed at baseline as cognitively and clinically healthy controls (global Clinical Dementia Rating = 0) from the Alzheimer Disease Neuroim-

aging Initiative. A total of 115 cognitively healthy older individuals had undergone longitudinal MR imaging, CSF lumbar puncture, and APOE ϵ 4 genotyping. Of these individuals, we restricted our analyses to those participants ($n = 107$) with quality-assured baseline and at least 1 follow-up MR imaging (6 months to 3.5 years; 10% with 6-month follow-up, 15% with 12-month follow-up, 34% with 23-month follow-up, and 41% with 36-month follow-up) available as of December 2011. We classified all participants on the basis of the presence (“carriers”) and absence (“noncarriers”) of at least 1 APOE ϵ 4 allele (Tables 1 and 2). Using recently proposed CSF cutoffs,¹⁵ we also classified all participants on the basis of high (>23 pg/mL, “positive”) and low (<23 pg/mL, “negative”) p- τ levels, and on low (<192 pg/mL, “positive”) and high (>192 pg/mL, “negative”) A β ₁₋₄₂ levels (Tables 1 and 2).

We examined 417 T1-weighted MR images. We performed quantitative surface-based analysis of all MR images by using an automated region-of-interest labeling technique¹⁶ and primarily focused on entorhinal cortex, a medial temporal lobe region that is selectively affected in the earliest stages of AD.¹⁷⁻²⁰ To additionally investigate neuroanatomic regions that are involved in the later stages of the disease process^{17,18} and to minimize multiple comparisons, we averaged longitudinal volume change in the temporal pole, parahippocampal gyrus, inferior temporal gyrus, banks of the superior temporal sulcus, inferior parietal lobule, amygdala, and hippocampus to create an “AD-vulnerable” region of interest (Fig 1). Using an automated method developed in our laboratory,²¹ we assessed longitudinal subregional change in gray matter volume (atrophy) on serial MR images.

We asked whether p- τ and APOE ϵ 4 independently influence A β -associated neurodegeneration. To investigate this question, we examined the main and interactive effects of CSF A β ₁₋₄₂ and

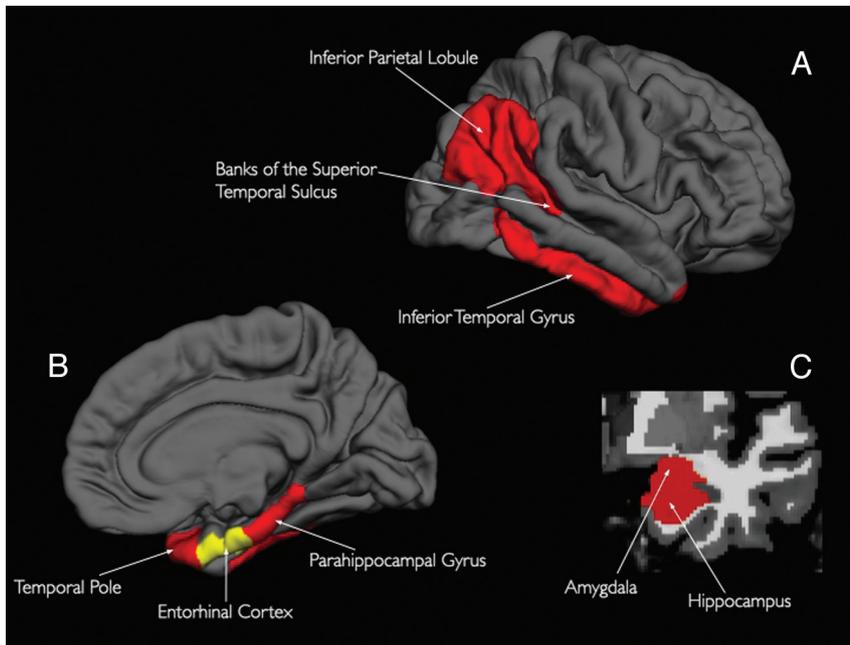


FIG 1. 3D representations of the neuroanatomic regions examined in the current study (only 1 hemisphere is shown). All of the neocortical regions are visible in the lateral (A) and medial (B) views of the gray matter surface, and the 2 non-neocortical regions (ie, the hippocampus and amygdala, C) are visible in the coronal view of a T1-weighted MR image. Regions illustrated in red constitute the AD-vulnerable region of interest (for further details please see text).

APOE $\epsilon 4$, and CSF $A\beta_{1-42}$ and CSF $p-\tau$ on entorhinal cortex atrophy rate in a mixed-effects model, covarying for the effects of age and sex, specifically

$$\begin{aligned} \Delta v = & \beta_0 \times \Delta t + \beta_1 APOE \epsilon 4_status \times \Delta t + \\ & \beta_2 CSF_A\beta_{1-42_status} \times \Delta t + \beta_3 CSF_p-\tau_status \times \Delta t + \\ & \beta_4 [APOE \epsilon 4_status \times CSF_A\beta_{1-42_status} \times \Delta t] + \\ & \beta_5 [CSF_p-\tau_status \times CSF_A\beta_{1-42_status} \times \Delta t] + \\ & covariates \times \Delta t + \epsilon. \end{aligned}$$

Here, Δv is entorhinal cortex atrophy (millimeters) and Δt is the change in time from baseline MR imaging (in years). Using the same linear mixed-effects framework, we also investigated the main and interactive effects of CSF $A\beta_{1-42}$ and *APOE* $\epsilon 4$, and CSF $A\beta_{1-42}$ and CSF $p-\tau$ on the atrophy rate in the AD-vulnerable region of interest.

RESULTS

Results from the full model with both interactive terms showed that the interaction between CSF $A\beta_{1-42}$ and CSF $p-\tau$ status on entorhinal cortex atrophy rate was significant ($\beta_5 = -0.39$, SE = 0.14, $P = .005$), indicating elevated atrophy with time in individuals with positive CSF $p-\tau$ and positive CSF $A\beta_{1-42}$ status (Fig 2A) as previously reported.¹⁴ In contrast, the interaction between CSF $A\beta_{1-42}$ and *APOE* $\epsilon 4$ on entorhinal cortex atrophy rate was not significant ($\beta_4 = -0.17$, SE = 0.18, $P = .35$). With both interaction terms in the model, the main effects of *APOE* $\epsilon 4$, CSF $A\beta_{1-42}$ status, and CSF $p-\tau$ status were not significant. Follow-up analyses

demonstrated that positive CSF $A\beta_{1-42}$ status was associated with an elevated entorhinal cortex atrophy rate only among CSF $p-\tau$ -positive individuals (β -coefficient = -0.32 , SE = 0.11, $P = .008$). There was no association between positive CSF $A\beta_{1-42}$ status and entorhinal cortex atrophy rate among CSF $p-\tau$ -negative individuals (β -coefficient = 0.10, SE = 0.08, $P = .23$) (Fig 2A). There was no association between positive CSF $A\beta_{1-42}$ status and entorhinal cortex atrophy rate either among *APOE* $\epsilon 4$ carriers (β -coefficient = -0.11 , SE = 0.19, $P = .58$) or noncarriers (β -coefficient = -0.02 , SE = 0.08, $P = .76$) (Fig 2B).

Similar results were obtained when examining the association of CSF protein and *APOE* $\epsilon 4$ status on the atrophy rate in the AD-vulnerable region of interest: The interaction of CSF $A\beta_{1-42}$ and CSF $p-\tau$ status on the AD-vulnerable region-of-interest atrophy rate was significant (β -coefficient = -0.34 , SE = 0.11, $P = .002$), but the interaction of CSF $A\beta_{1-42}$ and *APOE* $\epsilon 4$ was not (β -coefficient = -0.15 ,

SE = 0.14, $P = .28$). None of the main effects of *APOE* $\epsilon 4$, CSF $A\beta_{1-42}$ status, and CSF $p-\tau$ were significant with both interaction terms in the model. Follow-up analyses demonstrated that positive CSF $A\beta_{1-42}$ status was associated with an elevated AD-vulnerable region-of-interest atrophy rate among CSF $p-\tau$ -positive individuals (β -coefficient = -0.30 , SE = 0.09, $P = .001$) but not among CSF $p-\tau$ -negative individuals (β -coefficient = 0.03, SE = 0.07, $P = .61$). There was no association between positive CSF $A\beta_{1-42}$ status and atrophy rate in the AD-vulnerable region of interest either in *APOE* $\epsilon 4$ carriers (β -coefficient = -0.19 , SE = 0.13, $P = .09$) or noncarriers (β -coefficient = -0.06 , SE = 0.07, $P = .38$).

We also examined the possibility that *APOE* $\epsilon 4$ modulates AD-associated neurodegeneration via $p-\tau$ -related mechanisms. Using the same linear mixed-effects model framework described above, we concurrently examined the main and interactive effects of *APOE* $\epsilon 4$ and CSF $p-\tau$, CSF $A\beta_{1-42}$ and *APOE* $\epsilon 4$, and CSF $A\beta_{1-42}$ and CSF $p-\tau$ on the atrophy rate of entorhinal cortex and the AD-vulnerable region of interest. We did not find a significant interaction between *APOE* $\epsilon 4$ and CSF $p-\tau$ either on the atrophy rate of entorhinal cortex (β -coefficient = -0.04 , SE = 0.18, $P = .78$) or the AD-vulnerable region of interest (β -coefficient = 0.19, SE = 0.15, $P = .18$). Most important, even within this triple interaction model, the only significant effect was the interaction between CSF $A\beta_{1-42}$ and CSF $p-\tau$ on the atrophy rate of entorhinal cortex (β -coefficient = -0.38 , SE = 0.15, $P = .01$) and the AD-vulnerable region of interest (β -coefficient = -0.41 , SE = 0.12, $P = .001$).

Finally, although our results did not demonstrate a significant interaction between *APOE* $\epsilon 4$ and CSF $A\beta_{1-42}$ on longitudinal brain atrophy among HC, we examined whether the presence of

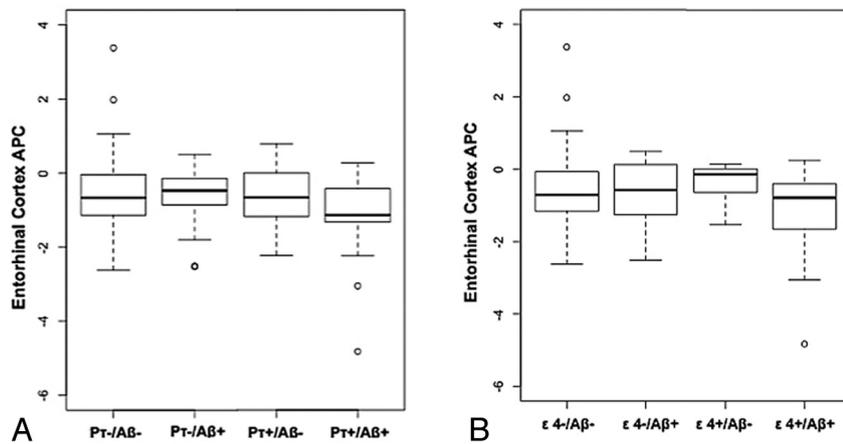


FIG 2. Box-and-whisker plots for all healthy control participants illustrating entorhinal cortex atrophy rate, measured as annualized percentage change (APC) based on CSF p- τ and CSF A β status (A) and $\epsilon 4$ genotype and CSF A β status (B). For each plot, thick black lines show the median value. Regions above and below the black line show the upper and lower quartiles, respectively. The dashed lines extend to the minimum and maximum values with outliers shown as open circles. As illustrated in A, the p τ +/A β + HC demonstrate the largest cortical atrophy rate (ie, more negative percentage change). In comparison as noted in B, the $\epsilon 4$ +/A β + HC show equivalent rates of atrophy compared with the other groups.

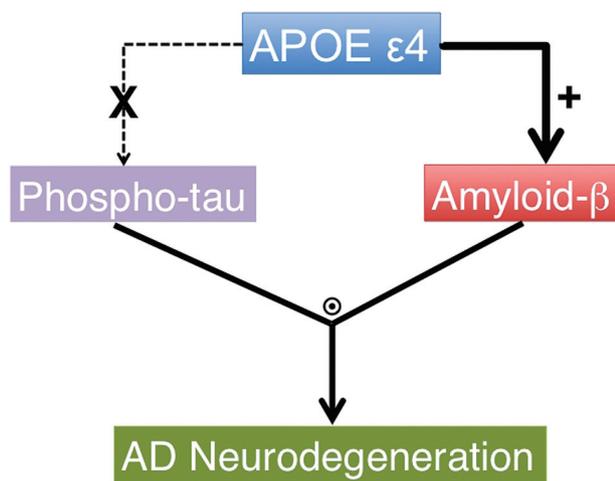


FIG 3. A conceptual model of AD-associated neurodegeneration in the preclinical phase of the disease process based on data from our mixed-effects models (please see text for details). The thickness of the arrows illustrates the magnitude of effect. The circle with a dot inside illustrates an interactive effect, the plus sign illustrates a positive effect, and X illustrates no significant effect.

APOE $\epsilon 4$ is associated with decreased CSF A β_{1-42} and increased CSF p- τ by using a generalized linear model, covarying for age and sex, specifically

$$\text{Logit}([\text{CSF_A}\beta_{1-42}\text{_status or CSF_p-}\tau\text{_status}]) = \beta_0 + \beta_1 \text{APOE } \epsilon 4\text{_status} + \beta_2 \text{Age} + \beta_3 \text{Sex.}$$

We found a significant relationship between *APOE* $\epsilon 4$ status and positive CSF A β_{1-42} status (β -coefficient = 0.40, SE = 0.07, $P = 4.82 \times 10^{-7}$), indicating increased A β deposition in $\epsilon 4$ carriers. In contrast, there was no relationship between *APOE* $\epsilon 4$ carriers and positive CSF p- τ status (β -coefficient = 0.05, SE = 0.09, $P = .55$).

DISCUSSION

In this study, we show that in cognitively healthy older individuals, though the presence of the $\epsilon 4$ allele is specifically associated with A β deposition, *APOE* $\epsilon 4$ does not affect A β -associated volume loss. In contrast, we found that p- τ modulates A β -associated neurodegeneration in clinically healthy individuals, as previously reported.¹⁴ These findings, in conjunction with recent experimental observations,^{22,23} support a conceptual model of the pathogenic cascade underlying preclinical AD (Fig 3), in which *APOE* $\epsilon 4$ primarily influences Alzheimer pathology via A β -related mechanisms; and in turn, A β -associated neurodegeneration occurs only in the presence of p- τ . This model provides a representation of the disease process that can be assessed with currently validated biomarkers, not a comprehensive framework of all pathologic

processes occurring in the earliest stages of AD. As such, it can be expanded to include future findings such as mechanistic details regarding the effect of genetic susceptibility loci on AD-associated neurodegeneration.

These findings provide important insights into the preclinical stage of AD. Although several studies in cognitively asymptomatic older individuals have demonstrated a significant relationship among *APOE* $\epsilon 4$ genotype, A β deposition, and neurodegeneration,^{10-12,24-26} there has been limited evaluation of the role of p- τ in modulating these relationships. Our findings indicate that in clinically healthy older individuals, A β deposition by itself, either in $\epsilon 4$ carriers or noncarriers, is not associated with volume loss; the presence of p- τ represents a critical link among the *APOE* $\epsilon 4$ genotype, A β deposition, and neurodegeneration. Consistent with prior reports,^{27,28} our results illustrate that the $\epsilon 4$ allele primarily affects AD in an indirect fashion via A β . In contrast, these findings do not support a role for *APOE* $\epsilon 4$ either in affecting intracranial p- τ levels or modulating AD pathology via p- τ -related mechanisms.

From a quantitative neuroimaging perspective, our results demonstrate the feasibility of using automated MR imaging-based measures of longitudinal brain atrophy as an *in vivo* biomarker even at the preclinical stage of the disease process. Building on prior neuroimaging studies in cognitively healthy older adults,^{10-12,24-26} these findings indicate that volume loss can be detected in older individuals testing positive for both A β and p- τ . Furthermore, the pattern of atrophy detected in this study is consistent with previous neuropathologic studies demonstrating neuronal loss within entorhinal cortex in the earliest stages of AD.^{19,20} Taken together, these findings suggest that the regionally specific volume loss occurring in a subset of cognitively healthy older adults is neuropathologically consistent with early AD.

This study has limitations. One concern is that CSF biomarkers provide an indirect assessment of amyloid and neurofibrillary pathology and may not fully reflect the pathologic processes underlying Alzheimer disease. Another limitation is that we primar-

ily focused on the *APOE* $\epsilon 4$ genotype and CSF biomarkers of the 2 pathologic hallmarks of AD. Additional genetic and cellular markers may also interact with $A\beta$ to predict neurodegeneration in cognitively healthy elders. Finally, the individuals examined here may represent a group of highly selected, generally healthy older adults who are motivated to participate in research studies. These findings therefore need to be further validated on an independent community-based cohort of older individuals who would be more representative of the general older population.

Clinically, these results indicate that a biomarker profile evaluating both $A\beta$ and $p\text{-}\tau$ may better identify those older individuals who are at an elevated risk of progressing to eventual dementia than either biomarker by itself. Consistent with prior clinical observations from our laboratory,²⁹ our current findings suggest that early intervention trials should take into account both the $p\text{-}\tau$ and $A\beta$ status of participants because older individuals with increased CSF $p\text{-}\tau$ and decreased CSF $A\beta_{1-42}$ levels are likely to have significantly elevated rates of volume loss compared with individuals with normal CSF $p\text{-}\tau$ and decreased CSF $A\beta_{1-42}$ levels. Finally, in addition to the current emphasis on $A\beta$, our findings identify the need for developing novel therapies that target *APOE*- and τ -related processes. It is likely that a complex interplay between multiple genetic and molecular entities determines AD pathogenesis.^{30,31} As such, targeting “upstream” events such as neuronal lipids and cholesterol transporters that interact with *APOE* in $\epsilon 4$ carriers with normal AD biomarker levels as well as “downstream” events such as τ phosphorylation and aggregation in older individuals with both decreased CSF $A\beta_{1-42}$ and increased CSF $p\text{-}\tau$ levels may represent additionally beneficial treatment strategies.

CONCLUSIONS

We show that in cognitively healthy older individuals, $p\text{-}\tau$ modulates the effect of $A\beta$ on neurodegeneration. In contrast, although the presence of the $\epsilon 4$ allele is specifically associated with $A\beta$ deposition, *APOE* $\epsilon 4$ does not influence $A\beta$ -associated volume loss. These findings provide important insights into the pathogenic cascade underlying preclinical AD and illustrate the importance of examining both $A\beta$ and $p\text{-}\tau$ in secondary prevention trials.

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Institutes of Health,* Alzheimer’s Association,* Baxter,* Pfizer,* Lilly,* Ole A. Andreassen—*RELATED: Grant:* Research Council of Norway,* *Comments:* travel expenses for and support for sabbatical at University of California at San Diego, *UNRELATED: Other:* pharmaceutical companies, *Comments:* I have received honoraria for lectures from pharmaceutical companies involved in medication for psychiatric disorders (Lundbeck, Lilly, Janssen, GlaxoSmithKline, MSD). Some of these companies may have medication for Alzheimer disease, but I am not aware of it. Bradley T. Hyman—*RELATED:* NIH grant to the Alzheimer Disease Research Center.* Reisa A. Sperling—*RELATED: Grant:* Regents of the University of California, San Diego,* *Comments:* 4-ADNI-PIB/ADC-029 (“PET Imaging of Brain Amyloid”), *UNRELATED: Consultancy:* Bayer,* Bristol-Myers-Squibb,* Eisai,* Janssen,* Kyowa Hakkō Kirin,* Pfizer,* Roche,* Satori,* Avid, *Comments:* Avid is unpaid, *Grants/Grants Pending:* NIH.* Anders Dale—*RELATED:* NIH/National Institute on Aging,* *Comments:* grant No. R01AG031224, *UNRELATED: Grants/Grants Pending:* NIH,* *Patents (planned, pending, or issued):* University of California at San Diego,* Massachusetts General Hospital/Harvard Medical School,* *Comments:* I am an inventor of multiple pending and issued patents on MRI acquisition and analysis methods, filed through University of California at San Diego or Massachusetts General Hospital/Harvard Medical School, *Stock/Stock Options:* Cortechs Labs Inc, *Comments:* I am a founder and equity holder of Cortechs Labs, Inc and also serve on its Scientific Advisory Board. The terms of this arrangement have been reviewed and approved by the University of California, San Diego, in accordance with its conflict-of-interest policies, *Other:* GE Healthcare,* *Comments:* I am also the Principal Investigator of a research agreement between GE Healthcare and University of California at San Diego. *Money paid to the institution.

REFERENCES

- Selkoe DJ. Resolving controversies on the path to Alzheimer’s therapeutics. *Nat Med* 2011;17:1060–65
- Spires-Jones TL, Meyer-Luehmann M, Osetek JD, et al. Impaired spine stability underlies plaque-related spine loss in an Alzheimer’s disease mouse model. *Am J Pathol* 2007;171:1304–11
- Kuchibhotla KV, Goldman ST, Lattarulo CR, et al. Abeta plaques lead to aberrant regulation of calcium homeostasis in vivo resulting in structural and functional disruption of neuronal networks. *Neuron* 2008;59:214–25
- Farrer LA, Cupples LA, Haines JL, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease: a meta-analysis—APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* 1997;278:1349–56
- Holtzman DM, Bales KR, Tenkova T, et al. Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer’s disease. *Proc Natl Acad Sci U S A* 2000; 97:2892–97
- Deane R, Sagare A, Hamm K, et al. apoE isoform-specific disruption of amyloid beta peptide clearance from mouse brain. *J Clin Invest* 2008;118:4002–13
- Roberson ED, Scarce-Levie K, Palop JJ, et al. Reducing endogenous tau ameliorates amyloid beta-induced deficits in an Alzheimer’s disease mouse model. *Science* 2007;316:750–54
- Ittner LM, Ke YD, Delerue F, et al. Dendritic function of tau mediates amyloid-beta toxicity in Alzheimer’s disease mouse models. *Cell* 2010;142:387–97
- Morris JC. Early-stage and preclinical Alzheimer disease. *Alzheimer Dis Assoc Disord* 2005;19:163–65
- Storandt M, Mintun MA, Head D, et al. Cognitive decline and brain volume loss as signatures of cerebral amyloid-beta peptide deposition identified with Pittsburgh compound B: cognitive decline associated with Abeta deposition. *Arch Neurol* 2009;66:1476–81
- Ewers M, Insel P, Jagust WJ, et al. CSF biomarker and PIB-PET-derived beta-amyloid signature predicts metabolic, gray matter, and cognitive changes in nondemented subjects. *Cereb Cortex* 2012; 22:1993–2004
- Chételat G, Villemagne VL, Villain N, et al. Accelerated cortical atrophy in cognitively normal elderly with high β -amyloid deposition. *Neurology* 2012;78:477–84
- Sperling RA, Aisen PS, Beckett LA, et al. Toward defining the preclinical stages of Alzheimer’s disease: recommendations from the National Institute on Aging-Alzheimer’s Association workgroups

- on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dementia* 2011;7:280–92
14. Desikan RS, McEvoy LK, Thompson W, et al. **Amyloid- β associated volume loss occurs only in the presence of phospho-tau.** *Ann Neurol* 2011;70:657–61
 15. Shaw LM, Vanderstichele H, Knapik-Czajka M, et al. **Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects.** *Ann Neurol* 2009;65:403–13
 16. Desikan RS, Ségonne F, Fischl B, et al. **An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest.** *Neuroimage* 2006;31:968–80
 17. Braak H, Braak E. **Neuropathological staging of Alzheimer-related changes.** *Acta Neuropathol* 1991;82:239–59
 18. Arnold SE, Hyman BT, Flory J, et al. **The topographical and neuro-anatomical distribution of neurofibrillary tangles and neuritic plaques in the cerebral cortex of patients with Alzheimer's disease.** *Cereb Cortex* 1991;1:103–16
 19. Gómez-Isla T, Price JL, McKeel DW Jr, et al. **Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer's disease.** *J Neurosci* 1996;16:4491–500
 20. Kordower JH, Chu Y, Stebbins GT, et al. **Loss and atrophy of layer II entorhinal cortex neurons in elderly people with mild cognitive impairment.** *Ann Neurol* 2001;49:202–13
 21. Holland D, Dale AM. **Nonlinear registration of longitudinal images and measurement of change in regions of interest.** *Med Image Anal* 2011;15:489–97
 22. Ittner LM, Gotz J. **Amyloid- β and tau: a toxic pas de deux in Alzheimer's disease.** *Nat Rev Neurosci* 2011;12:65–72
 23. Kim J, Basak JM, Holtzman DM. **The role of apolipoprotein E in Alzheimer's disease.** *Neuron* 2009;64:632–44
 24. Schott JM, Bartlett JW, Fox NC, et al. **Increased brain atrophy rates in cognitively normal older adults with low cerebrospinal fluid A β 1–42.** *Ann Neurol* 2010;68:825–34
 25. Becker JA, Hedden T, Carmasin J, et al. **Amyloid- β associated cortical thinning in clinically normal elderly.** *Ann Neurol* 2011;69:1032–42
 26. Kantarci K, Lowe V, Przybelski SA, et al. **APOE modifies the association between A β load and cognition in cognitively normal older adults.** *Neurology* 2012;78:232–40
 27. Sunderland T, Mirza N, Putnam KT, et al. **Cerebrospinal fluid beta-amyloid1–42 and tau in control subjects at risk for Alzheimer's disease: the effect of APOE epsilon4 allele.** *Biol Psychiatry* 2004;56:670–76
 28. Morris JC, Roe CM, Xiong C, et al. **APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging.** *Ann Neurol* 2010;67:122–31
 29. Desikan RS, McEvoy LK, Thompson WK, et al. **Amyloid- β -associated clinical decline occurs only in the presence of elevated p-tau.** *Arch Neurol* 2012;69:709–13
 30. Holtzman DM, Morris JC, Goate AM. **Alzheimer's disease: the challenge of the second century.** *Sci Transl Med* 2011;3:77sr1
 31. Hyman BT. **Amyloid-dependent and amyloid-independent stages of Alzheimer disease.** *Arch Neurol* 2011;68:1062–64