

The Effect of Pulse Sequence Parameters and Contrast Agent Dose on Percentage Signal Recovery in DSC-MRI: Implications for Clinical Applications

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ABSTRACT

BACKGROUND AND PURPOSE: Both technical and pathophysiologic factors affect PSR in DSC-MR imaging. We aimed to determine how TE, flip angle (α), and contrast dose impact PSR in high-grade gliomas.

MATERIALS AND METHODS: We retrospectively computed PSR maps for 22 patients with high-grade gliomas, comparing 3 DSC-MR imaging methods by using single-dose gadodiamide without preload administration: A ($n = 7$), $\alpha = 35^\circ$, TE = 54 ms; B ($n = 5$), $\alpha = 72^\circ$, TE = 30 ms; C ($n = 10$), $\alpha = 90^\circ$, TE = 30 ms. Methods A-C served as preload for subsequent dynamic imaging using method D (method C parameters but with double-dose contrast). We compared first- and second-injection tumor PSR for methods C and D (paired *t* test) and tumor PSR for both injections grouped by the first-injection acquisition method (3-group nonparametric 1-way ANOVA). We compared PSR in tumor and normal brain for each first- and second-injection method group (paired *t* test).

RESULTS: First-injection PSR in tumor and normal brain differed significantly for methods B ($P = .01$) and C ($P = .05$), but not A ($P = .71$). First-injection tumor PSR increased with T1 weighting with a significant main effect of method groupings ($P = .0012$), but there was no significant main effect for first-injection normal brain ($P = .93$), or second-injection tumor ($P = .95$) or normal brain ($P = .13$). In patients scanned with methods C and D, first-injection PSR significantly exceeded second-injection PSR for tumor ($P = .037$) and normal brain ($P < .001$).

CONCLUSIONS: PSR strongly depends on the T1 weighting of DSC-MR imaging, including pulse sequence (TE, α) and contrast agent (dose, preload) parameters, with implications for protocol design and the interpretation and comparison of PSR values across tumor types and imaging centers.

ABBREVIATIONS: DSC = dynamic susceptibility contrast; GBM = glioblastoma multiforme; PSR = percentage signal recovery; rCBV = relative cerebral blood volume

First-pass transient signal loss is the fundamental feature of DSC-MR imaging signal-time curves and the principal determinant of rCBV estimates. However, postbolus signal depression or elevation is frequently observed and represents failure of the signal-time curve, $S(t)$, and the corresponding relaxivity-time

curve, $\Delta R2^*(t)$, to return to the prebolus baseline as expected in idealized tracer kinetics. The variable return of the postbolus signal to the prebolus baseline has been termed “percentage signal recovery” and was proposed as a tool for differentiating nonenhancing gliomas and non-neoplastic lesions¹; progressive metastatic disease from radiation injury after stereotactic radiosurgery^{2,3}; recurrent GBM from radiation necrosis⁴; and common intra-axial brain neoplasms such as metastasis, high-grade glioma, and lymphoma.⁵⁻⁸

Although PSR has promise for elucidating tumor biology, its etiology is poorly characterized and likely the complex multifactorial interplay of technical (contrast dose, DSC-MR imaging pulse sequence) and pathophysiologic (vascular permeability, tumor cell volume fraction) factors. A spectrum of pulse-sequence parameters with low (long TE³), intermediate (low flip angle, intermediate TE^{2,4,5} and high flip angle, intermediate TE^{1,7}), and high (high flip angle, short TE⁶) T1 weighting have been published for quantifying PSR, with some conflicting inconsistent

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results. Although PSR studies have typically used single-dose contrast without preload, PSR may be evaluated with rCBV, for which higher contrast dose and preload administration for reducing T1 leakage contamination are beneficial^{9,10}; the dependence of PSR on contrast dose and preload is therefore important. For instance, elevated postbolus relaxivity is observed following second contrast agent injections in normal brain¹¹ and brain tumors having BBB disruption, especially following preload administration that diminishes T1 leakage effects.¹² An understanding of the dependence of postbolus signal on these nonphysiologic factors is important for designing protocols that accentuate physiology-dependent differences in PSR and for interpreting PSR results in the clinical setting.

In this study, we aimed to determine how TE, flip angle, and contrast agent dose in DSC-MR imaging impact PSR and to explain why there may be discrepancies in reported PSR values for similar tumors. We hypothesized that PSR depends, in part, on the T1 weighting of the pulse sequence and the total dose of administered contrast agent, including preload. Specifically, at low contrast agent doses, postbolus signal can be dominated by T1 shortening in the intravascular (normal brain) and/or extravascular extracellular space (tumor) when a T1-weighted pulse sequence is used, yielding relatively high PSR. However, with reduced T1 weighting, increased T2* weighting, and increased contrast agent dose, T2* effects can dominate, with substantially smaller PSR. The choice of pulse-sequence parameters (TE, flip angle) and contrast agent dose (administration of preload), which impact the accuracy of rCBV estimation,^{9,10} also has implications for PSR estimates, and understanding this relationship is important for properly selecting PSR imaging parameters, interpreting PSR results, and comparing values across tumor types and imaging centers.

MATERIALS AND METHODS

Patient Selection

We retrospectively analyzed DSC-MR imaging examinations for patients with a cerebral glioma who were previously recruited for perfusion-weighted imaging as a supplement to conventional contrast-enhanced brain MR imaging. Inclusion and exclusion criteria are described in Paulson and Schmainda.¹² All patients provided informed written consent under Health Insurance Portability and Accountability Act—compliant guidelines established by the institutional review board at the Medical College of Wisconsin. Tumors were classified and graded according to the World Health Organization 1993 Guidelines.

MR Imaging

All studies were performed at 1.5T (CVi; GE Healthcare, Milwaukee, Wisconsin) with commercial gradients and phased-array radio-frequency coils. To assess the dependence of PSR on pulse-sequence parameters, we compared 3 common DSC-MR imaging-acquisition methods with increasing degrees of T1 weighting, each by using single-dose (0.1 mmol/kg) gadodiamide (Omniscan; Nycomed Amersham, Princeton, New Jersey) without preload contrast administration: method A ($n = 7$), single echo with a low flip angle ($\alpha = 35^\circ$) and long TE (54 ms) yielding minimum T1 sensitivity¹³⁻¹⁵; method B ($n = 5$), second echo of a dual-echo spiral acquisition (first-echo TE = 3.3 ms) with an intermediate flip angle ($\alpha = 72^\circ$) and short TE (30 ms), yielding

Table 1: Summary of DSC-MRI pulse-sequence parameters compared in this study

MRI Method	No.	Preload (mmol/kg)	Bolus (mmol/kg)	α (°)	TE (ms)
A	7	0	0.1	35	54
B ^a	5	0	0.1	72	30
C	10	0	0.1	90	30
D	22	0.1	0.2	90	30

^a Second echo of a dual-echo acquisition (first-echo TE = 3.3 ms); other methods were single-echo acquisitions. All methods used gradient-echo EPI.

intermediate T1 sensitivity^{16,17}; and method C ($n = 10$), single echo with high flip angle ($\alpha = 90^\circ$) and short TE (30 ms), yielding the greatest T1 sensitivity.^{9,18,19} Method D, applied to all patients, was identical to method C but used double-dose contrast (0.2 mmol/kg) following the 0.1 mmol/kg preload that served as the bolus for dynamic imaging in methods A–C. Table 1 summarizes the pulse sequence parameters for each method. For single-echo acquisitions, a lipid-suppressed single-shot gradient-echo EPI pulse sequence acquired images for 1 minute before and 2 minutes after bolus injection of contrast agent at 3–5 mL/s (FOV = 24 cm², matrix = 64 × 64, TR = 1000 ms, section thickness = 5 mm, intersection gap = 1.5 mm, 12 sections). For the dual-echo acquisition, a custom lipid-suppressed single-shot dual-echo gradient-echo spiral-out sequence was used with the same general parameters as the single-echo acquisitions.²⁰ Following DSC-MR imaging, postcontrast T1-weighted spin-echo images were acquired (TR = 450 ms, TE = 11 ms, NEX = 2, matrix = 256 × 256).

PSR Computation

For each patient, we constructed voxelwise PSR maps from the time course data for both contrast agent injections by using a standard technique⁷:

$$\text{PSR} = (S_1 - S_{\min}) / (S_0 - S_{\min}) \times 100\%$$

where baseline (S_0) and postbolus signal intensity (S_1) equals the average signal intensity of time points 25–55 and 125–155, respectively, and S_{\min} is the minimum signal intensity within the first-pass bolus. We selected a region of interest (30–50 mm²) for enhancing tumor, excluding areas of necrosis and nontumor macrovessels; and a region of interest of identical size was specified for contralateral normal-appearing brain. The same ROIs for each patient were used for both contrast agent injections. Mean and standard error of the PSR within tumor and normal brain ROIs were computed for both injections for each patient by translating each region of interest onto the appropriate PSR map.

Statistical Methods

Statistical analysis was performed by using GraphPad Prism, Version 5.0d (GraphPad Software, LA Jolla, California). To investigate the dependence of PSR on contrast agent dose and preload, we compared mean tumor PSR from the first and second injections (differing in bolus dose and the presence of preload) in patients scanned with the otherwise identical methods C (first injection) and D (second injection) by using a 2-tailed paired t test. We repeated the analysis for mean PSR in normal brain. To investigate the dependence of PSR on pulse-sequence parameters, we compared first-injection mean tumor PSR in patients grouped by acquisition methods A–C by using a 3-group nonparametric

Table 2: Mean PSRs grouped by DSC-MRI acquisition method

MRI Method ^a (Inj1, Inj2)	Mean PSR (% ± SEM)								
	Tumor Characteristics					Mean PSR (% ± SEM)			
	No.	GIII	GIV	New	Rec	1st Injection		2nd Injection	
						Tumor	Normal	Tumor	Normal
A, D	7	1	6	1	6	91 ± 10	96 ± 5	81 ± 10	96 ± 8
B, D	5	1	4	0	5	217 ± 32	103 ± 14	76 ± 8	78 ± 4
C, D	10	5	5	7	3	372 ± 122	95 ± 2	79 ± 8	80 ± 2

Note:—SEM indicates standard error of the mean; Inj1, injection 1; Inj2, injection 2; GIII, grade III gliomas; GIV, grade IV gliomas; New, treatment-naïve or residual postoperative; Rec, recurrent (after conventional chemoradiation).

^a MRI methods refer to first-injection and second-injection acquisition techniques for each patient (Table 1).

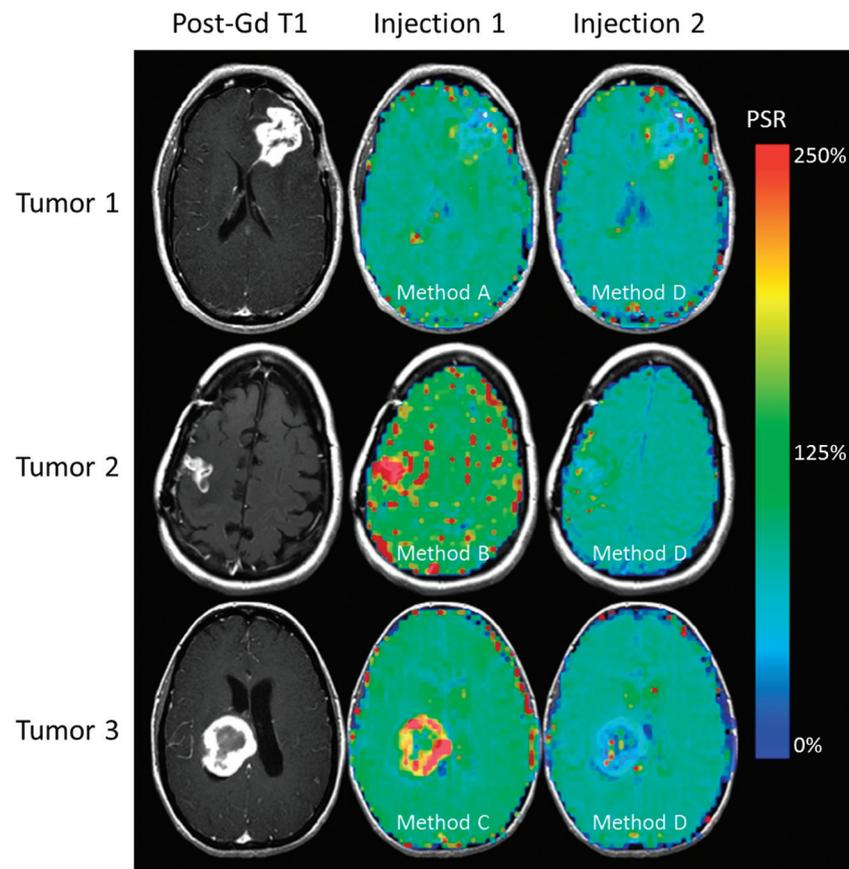


FIG 1. Three sample tumors (first column) with corresponding first-injection (middle column) and second-injection (third column) PSR maps. First-injection PSR progressively increases with increasing T1 weighting of methods A (first row), B (second row), and C (third row). Second-injection maps (all by using method D) have lower PSR with poor contrast between tumor and normal brain, despite flip angles and TEs identical to those in method C. Preload administration decreases T1 sensitivity and PSR contrast, regardless of flip angle and TE.

1-way ANOVA (Kruskal-Wallis test), and we performed follow-up group comparisons by using the Dunn multiple comparison test. We repeated this analysis for second-injection mean tumor PSR, and for first- and second-injection mean PSR in normal brain. We also compared mean PSR in tumor and normal brain for each first- and second-injection method group, by using a 2-tailed paired *t* test. For all tests, we assumed a statistically significant threshold of *P* = .05.

RESULTS

Study Cohort

Twenty-two patients (12 men, 10 women; 21–76 years of age; mean age, 49 years) with high-grade gliomas were enrolled. Grade III tu-

mors included new (*n* = 1) and recurrent (*n* = 2) anaplastic astrocytomas, recurrent anaplastic mixed gliomas (*n* = 2), new (*n* = 1) and recurrent (*n* = 1) anaplastic oligodendrogliomas, and malignant glioneuronal tumor (*n* = 1). Grade IV tumors included newly diagnosed (*n* = 5), recurrent (*n* = 7), and residual GBMs (*n* = 2). At the time of DSC-MR imaging, 15 patients were being treated with steroids. Table 2 summarizes the tumor characteristics grouped by the first-injection acquisition method, as well as intragroup mean and standard error in tumor and normal brain for both first (methods A–C) and second (all method D) injections. For the first injection (no preload), methods A–C have increasing T1 weighting (increasing flip angle, decreasing TE), with increasing PSR in tumors that substantially exceeds 100% for methods B–C and equivalent PSR ≈ 100% for all methods in normal brain. For the second injection (with preload), all 3 groups used method D, with essentially equivalent PSRs in tumor (76%–81%) and normal brain (78%–96%).

Figure 1 illustrates 3 cases, each with contrast-enhanced tumor (first column) and corresponding first-injection (middle column) and second-injection (third column) PSR maps. First-injection methods A (first row), B (second row), and C (third row) represent increasing T1 weighting, with progressively higher PSR values. With increasing flip angles and decreasing TEs, T1 sensitivity and PSR increase. The second-injection maps (all acquired with method D) demonstrate poor PSR contrast with substantially lower PSR. Although method D has flip angles and TEs identical to those in method C, which yields high first-injection PSR, preload contrast administration substantially diminishes T1 effects, reducing tumor PSR to levels seen with the first-injection method A.

In the presence of preload contrast administration, T1 sensitivity and PSR decrease, regardless of flip angle and TE.

Dependence of PSR on Contrast Agent Dose and Timing

Figure 2A illustrates sample first-injection signal-time curves for tumor and normal brain acquired with method C (heavily T1 weighted; α = 90°, and TE = 30 ms). Without preload, there is high postbolus signal intensity and PSR > 100% for tumor, with PSR ≈ 100% for normal brain. Figure 2B illustrates the corresponding second-injection signal-time curves for identical ROIs and acquisition parameters (method D). Preload diminishes the T1 effects in tumor from extravasated contrast, yielding reduced postbolus signal and PSR < 100%. In normal brain, post-

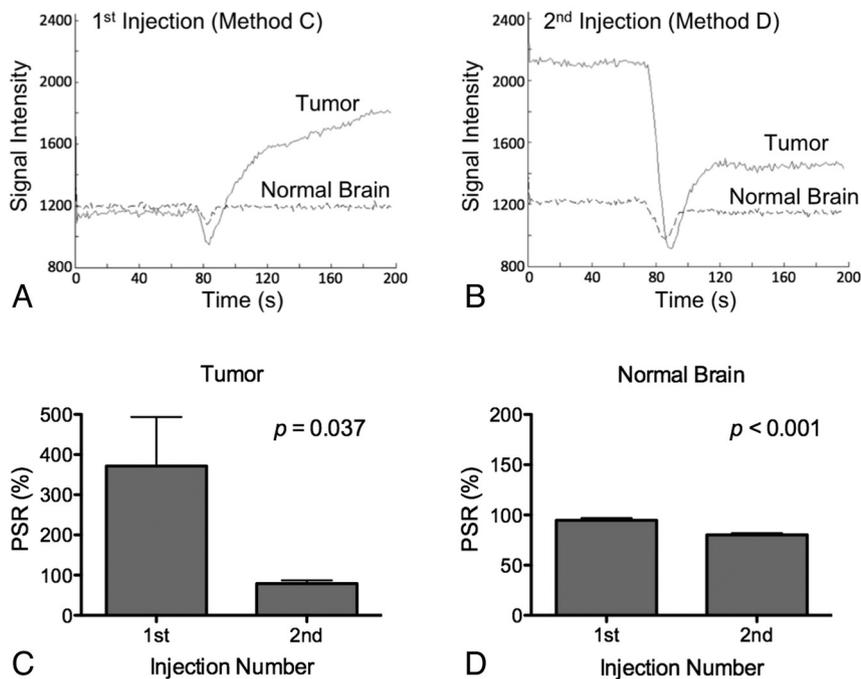


FIG 2. Sample first-injection (A) and second-injection (B) signal-time curves for tumor and normal brain by using methods C and D ($\alpha = 90^\circ$, TE = 30 ms). Without preload (A), tumor PSR is $>100\%$, whereas with preload (B), tumor PSR is $<100\%$. First-injection PSR (method C) is significantly larger than second-injection PSR (method D) for both tumor (C; $P = .037$) and normal brain (D; $P < .001$) by using the 2-tailed paired *t* test.

bolus signal does not quite reach baseline, with PSR $< 100\%$. Figure 2C, -D compares mean PSR on the first (method C) and second (method D) injections, both with $\alpha = 90^\circ$ and TE = 30 ms, in tumor and normal brain, respectively. First-injection PSR significantly exceeds second-injection PSR for both tumor ($P = .037$) and normal brain ($P < .001$) by using the 2-tailed paired *t* test. These results demonstrate that PSR can differ significantly, depending on whether preload contrast is administered.

Dependence of PSR on MR Imaging Acquisition Parameters

For the first injection, there was no significant difference between mean PSR in tumor and normal brain for method A ($P = .71$), but there was a significant difference for methods B ($P = .01$) and C ($P = .05$). For the second-injection method (method D), there was no significant difference in patients grouped by the first-injection method A ($P = .15$), B ($P = .71$), or C ($P = .85$).

Figure 3 compares mean PSR values in tumor and normal brain for first-injection methods A–C (Fig 3A, -C) and for second-injection method D (Fig 3B, -D) with patients grouped by the first-injection method. For the first injection in tumors, there was a significant main effect of method groupings on PSR by the Kruskal-Wallis test ($P = .0012$; Fig 3A), with mean PSR monotonically increasing with T1 weighting. Follow-up group comparisons (Dunn multiple comparison test) revealed that method A tumors had significantly lower PSR than method B and method C tumors; there was no significant difference in PSR between method B tumors and method C tumors. For normal brain, there was no significant main effect of first-injection acquisition-method groupings on PSR by the Kruskal-Wallis test ($P = .93$;

Fig 3C). For the second injection, all patients were scanned with method D, regardless of first-injection methodology. There was no significant main effect of the first-injection acquisition-method groupings on second-injection PSR by the Kruskal-Wallis test for tumor ($P = .95$; Fig 3B) or normal brain ($P = .13$; Fig 3D).

DISCUSSION

PSR compares postbolus DSC-MR imaging signal recovery with prebolus baseline and has the potential for differentiating neoplastic and non-neoplastic primary lesions¹; progressive primary⁴ or metastatic^{2,3} neoplasms from radiation injury following radiosurgery; and common intra-axial brain tumors including metastasis, high-grade glioma, and lymphoma.⁵⁻⁸ However, a formal study of the technical factors contributing to PSR has not been performed, to our knowledge. This analysis is important for understanding the degree to which PSR variations from technical factors can overshadow desired variations due to underlying pathophysiology.

The postbolus DSC-MR imaging signal has T1 and T2* contributions resulting from the complex interplay of biophysical factors, including vascular, cell, and interstitial volume fractions and vascular permeability, which govern the relative contrast agent concentrations in the vascular and extravascular extracellular space.²¹ T2* relaxivity arises from the superposition of the “positive” susceptibility effect due to intravascular compartmentalization of contrast agent within tumor microvasculature and the “inverse” susceptibility effect, due to exclusion from cells of extravasated contrast agent within the extravascular extracellular space, and depends on the proton diffusion rate within tissue. T1 relaxivity arises from dipole-dipole interactions between protons and contrast agent weighted by relative concentrations and volumes of distribution in the intravascular and extravascular extracellular compartments. Presumably, differing tumor-specific cell densities, vascular permeabilities, and vascular volume fractions in GBM, lymphoma, and metastases yield unique PSR signatures permitting their differentiation,⁷ with analogous physiologic differences accounting for discrepant PSR in tumor, radiation necrosis, and other non-neoplastic processes.

Pulse-sequence parameters also affect PSR by conferring different T1 and T2* weighting on DSC-MR imaging signal, and our results indicate that this effect can be profound. For enhancing tumors with extravasated contrast, increased T1 weighting with relatively low T2* weighting yields higher PSR estimates, which can substantially exceed 100% for high-flip-angle low-TE acquisitions and TR = 1000 ms (Fig 3A). In fact, our signal-time curves for some high-grade gliomas by using method C with high T1 weighting (Fig 2A) look nearly identical to those for lymphoma by Mangla et al (Fig 1B),⁷ exemplifying how PSR dependence on

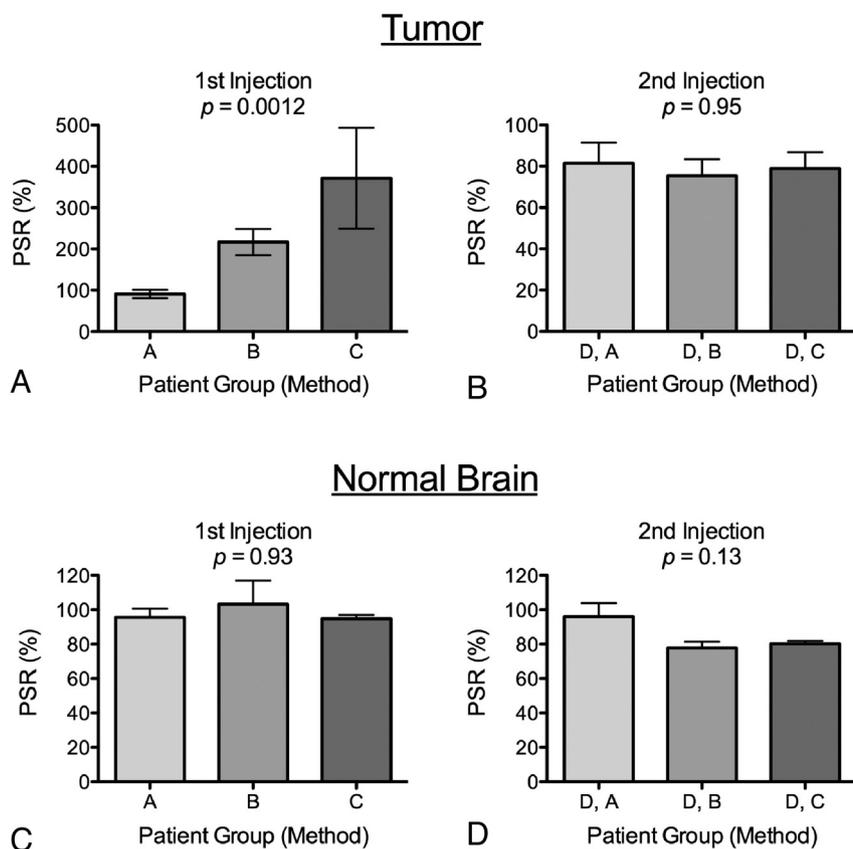


FIG 3. Comparison of PSR in tumor (A) and normal brain (C) for methods A-C and in tumor (B) and normal brain (D) for second-injection method D grouped by first-injection method. There is a significant main effect of method groupings on first-injection tumor PSR (A; $P = .0012$), with PSR monotonically increasing with T1 weighting and no significant main effect on first-injection normal brain PSR (C; $P = .93$). There is no significant main effect of first-injection method groupings on second-injection PSR for tumor (B; $P = .95$) or normal brain (D; $P = .13$).

pulse sequence can confound clinical interpretations of PSR. Pre-load administration eliminates PSR dependence on T1 weighting even in tumors (Fig 3B) by saturating the baseline extravascular extracellular compartment T1-weighted signal intensity, thereby diminishing T1-induced increased signal during second-injection DSC-MR imaging. It may also reduce the gradient of contrast efflux. This effect is desirable for rCBV estimation, in which pre-load and postprocessing algorithms reduce error in rCBV estimates by minimizing T1 contamination,^{9,10} but it is suboptimal for PSR mapping because T1 sensitivity plays a pivotal role in establishing unique physiology-dependent PSR signatures of different neoplastic and non-neoplastic enhancing lesions. For instance, there was no significant difference in PSR between tumor and normal brain for first-injection method A with low T1 weighting, and second-injection postpreload PSR for tumor and normal brain was not significantly different, even for the high T1 weighting of method D.

Optimal DSC-MR imaging pulse sequences for PSR discrimination that have substantial T1 weighting may, therefore, be suboptimal for accurate CBV imaging, in which contaminating T1 effects should be minimized with preload and a low flip angle. There are several approaches to a comprehensive DSC protocol. PSR imaging could be performed dynamically on the preload injection with subsequent dynamic CBV imaging on the second

injection. An alternative approach uses a dual-echo acquisition during single contrast agent injection, when parameters for the second echo are selected to provide T1 and T2* weighting for PSR derivation and the first echo is used with the second echo to eliminate T1 weighting for rCBV mapping.²⁰

Our measured PSR values match the literature values for comparable acquisition parameters. We found the mean first-injection tumor PSR with method A to be 91%, in good agreement with mean values of 81% observed by Cha et al⁵ and 80% by Barajas et al⁴, both obtained with identical flip angles and TEs for GBMs. Our slightly higher value may result from a mix of high-grade gliomas in our population, shorter TRs (1000 ms versus 1000–1250 ms), and more baseline points, which may lower the baseline estimate due to reduced non-steady-state contamination.

Our study does not investigate the impact of TR on PSR. We used TR = 1000 ms for all methods; this was our protocol for DSC examinations geared toward rCBV estimation, because the high temporal resolution of $\Delta R2^*(t)$ improves rCBV map SNR.²² Longer TR confers greater T2 weighting, and this effect can be substantial. For example, TE and α for our method C are identical to those used by Liao et al,⁶ but their mean PSR in GBM was 93% for TR =

1440 ms, compared with our value of 372%. In fact, the morphology of our S(t) curve and corresponding PSR value for these parameters are closer to those found by Liao et al for lymphoma than for GBM. Furthermore, our measured first-injection PSR for method A of 91% is similar to the mean GBM PSR of 78% found by Mangla et al⁷ and the mean nonenhancing glioma PSR of 88% found by Liu et al¹ for TE = 50 ms, with the substantially increased T2 weighting for their TR = 1500 ms apparently offsetting the much higher T1 weighting for their $\alpha = 80^\circ$.

Because pulse-sequence parameters can significantly affect the magnitude of measured PSR, care should be taken when interpreting and comparing PSR results from the literature. For instance, Huang et al³ found no difference in PSR for progressive metastases versus radiation injury by using a heavily T2-weighted acquisition (TR/TE = 2480/98 ms), whereas Barajas et al² found a significant difference by using intermediate T1 weighting (TR/TE/ $\alpha = 1250$ ms/54 ms/35°). Furthermore, mean glioma PSRs between 78% and 93% have been reported by using techniques ranging from high α , low TE⁶ to low α , intermediate TE,⁴ all with intermediate TRs between 1250 and 1500 ms. Although this range of PSR is smaller than ours obtained with a much smaller TR, there is overlap of the highly T1-weighted comparative GBM data⁶ with our first-injection normal brain PSRs and of lower T1-weighted comparative data with our second-injection normal

brain PSRs, further emphasizing the sensitivity of PSR estimates to DSC-MR imaging methodology.

This study has several limitations. Our sample population was heterogeneous and included grade III and IV tumors that are both treatment-naïve and recurrent, with variable steroid administration; this may introduce some variability. Vascular volume fraction for GBM exceeds that for grade III gliomas,⁹ with cell density and vascular permeability differences; chemoradiation and steroids can affect tumor physiology as well. However, tumor populations were mixed in our 3 method groups, and the aggregate populations are probably similar. Intergroup comparison (Fig 3) should, therefore, emphasize true pulse sequence differences rather than variable underlying tumor physiology. Furthermore, we demonstrated significant differences in PSR due to contrast agent preload (Fig 2) by using a paired nonparametric *t* test in which each tumor was its own control, and a heterogeneous tumor population would not affect this analysis. Our retrospective data did not permit us to isolate the effects of contrast agent dose versus preload on PSR. To do so would require a comparison, for instance, of single- and double-dose contrast agent for the first injection or single-dose administrations of 2 different contrast agents with different relaxivities.

CONCLUSIONS

PSR strongly depends on the T1 weighting of the DSC-MR imaging acquisition as determined by pulse sequence (TE, flip angle) and contrast agent (dose and preload) factors. Understanding this relationship is important for properly selecting PSR imaging parameters, interpreting PSR results, and comparing values across tumor types and imaging centers. Because accurate CBV imaging requires minimization of T1 effects, comprehensive DSC-MR imaging protocols that aim to measure rCBV and PSR for assessing tumor physiology must address these competing factors.

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