Heterogeneity of Cortical Lesion Susceptibility Mapping in Multiple Sclerosis

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ABSTRACT

BACKGROUND AND PURPOSE: Quantitative susceptibility mapping has been used to characterize iron and myelin content in the deep gray matter of patients with multiple sclerosis. Our aim was to characterize the susceptibility mapping of cortical lesions in patients with MS and compare it with neuropathologic observations.

MATERIALS AND METHODS: The pattern of microglial activation was studied in postmortem brain tissues from 16 patients with secondary-progressive MS and 5 age-matched controls. Thirty-six patients with MS underwent 3T MR imaging, including 3D double inversion recovery and 3D-echo-planar SWI.

RESULTS: Neuropathologic analysis revealed the presence of an intense band of microglia activation close to the pial membrane in subpial cortical lesions or to the WM border of leukocortical cortical lesions. The quantitative susceptibility mapping analysis revealed 131 cortical lesions classified as hyperintense; 33, as isointense; and 84, as hypointense. Quantitative susceptibility mapping hyperintensity edge found in the proximity of the pial surface or at the white matter/gray matter interface in some of the quantitative susceptibility mapping–hypointense cortical lesions accurately mirrors the microglia activation observed in the neuropathology analysis.

CONCLUSIONS: Cortical lesion susceptibility maps are highly heterogeneous, even at individual levels. Quantitative susceptibility mapping hyperintensity edge found in proximity to the pial surface might be due to the subpial gradient of microglial activation.

ABBREVIATIONS: CL = cortical lesion; DIR = double inversion recovery; EDSS = Expanded Disability Status Scale; MHC = major histocompatibility complex; MOG = myelin oligodendrocyte glycoprotein; NAGM = normal-appearing gray matter; nQSM = QSM value in the NAGM; nQSMcontra = median value of susceptibility from the reference tissue mask in the contralateral hemisphere; nQSMedge = median value of susceptibility from the reference tissue mask surrounding the lesion; QSM = quantitative susceptibility mapping; RRMS = relapsing-remitting multiple sclerosis; SPMS = secondary-progressive multiple sclerosis

In recent years, it has become increasingly clear that cortical and deep gray matter are not spared in multiple sclerosis.1,2 Several neuropathologic studies have consistently demonstrated that cortical gray matter lesions (CLs) are frequent in MS and that their accumulation strongly correlates with long-term disability measures.3 These observations have been confirmed and extended by imaging studies showing that CLs correlate with both physical and cognitive disability.4,5 Unfortunately, despite these data, little is known about the pathogenetic mechanisms underlying CL development.

Neuropathologic studies have highlighted the lack of substantial focal immune infiltrates, complement deposition, and blood-brain barrier damage in MS CLs6,7 and have suggested that meningeal inflammation and activated microglia may have a key role in GM damage.3,8 In particular, most of the examined CLs in postmortem brain samples exhibit a chronic inflammatory phenotype and rims of activated microglia close to the pial surface or the lesion edge.3,8,9

In a previous MR imaging/histopathologic combined study,10 hypointense rings on T2*, representing activated microglia or...
Gene expression in CLs, with a predominant iron loss hypothesis. Similar rings have been reported in white matter lesions by using T2* phase imaging. These results, in line with those by Kooi et al., showed that some patients with MS had rims of activated microglia at the border of the CLs, whereas others did not. More recently, a study on ultra-high-field MR imaging on postmortem specimens of 2 patients with MS did not find rings of activated iron-laden microglia within CLs, and all CLs appeared darker in R2* images and brighter in magnitude images.

Magnetic susceptibility is a fundamental physical tissue property, which is known to reflect clinically relevant tissue characteristics, such as tissue iron content. During the last decades, phase imaging, SWI, and T2* imaging have been used to qualitatively assess magnetic susceptibility variations in cerebral tissue, including deep and cortical gray matter. Increased paramagnetic susceptibility-weighted filtered phase values were observed in the putamen in patients with clinically isolated syndrome compared with healthy controls. This finding suggests that susceptibility is sensitive to MS even in the early phase of the disease. In this context, quantitative susceptibility mapping (QSM), which overcomes several nonlocal restrictions of susceptibility-weighted and phase imaging, could shed some light on the pathologic process taking place in the cortical GM of patients with MS. A recent study has investigated MS, using QSM in the basal ganglia, showing that iron accumulation correlated with disease progression even in a patient with clinically isolated syndrome. Moreover, another group showed how WM MS lesions could be investigated longitudinally with QSM; the investigation could provide insight into the pathogenesis of those lesions. However, limited quantitative susceptibility data of cortical GM and, especially, of CLs are available. Indeed, only a recent study at 7T showed, in a restricted cohort of patients with MS, heterogeneity in CLs, with a predominant iron loss hypothesis.

In the present study, the characteristics of CLs were first determined with an analysis of microglial/macrophage activity in the postmortem MS brain, followed by the analysis of similar lesions in patients. 3D echo-planar imaging, which uses phase data to quantify local tissue susceptibility, was combined with a 3D double inversion recovery (DIR) at 3T to characterize the in vivo susceptibility of CLs in patients with MS.

MATERIALS AND METHODS
Neuropathologic Analysis
This study was performed on postmortem brain tissues from 16 patients with secondary-progressive multiple sclerosis (SPMS) (mean age at death, 44.4 ± 6.2 years, Table 1; disease duration, 23.3 ± 8.55 years; time to wheelchair [from onset to Expanded Disability Status Scale (EDSS) 7], 41.21 ± 7.83 years; relapse rate in the first 2 years of the disease, 2.6 ± 1.3) selected for the presence of widespread cortical demyelination associated with meningeal inflammation and by more rapid and severe disease outcome associated with intense inflammatory activity among a larger group of 48 patients with SPMS previously extensively characterized for the presence and levels of inflammatory features and the extent of gray and white matter demyelination. Postmortem brain tissues from 8 controls, with no neurologic diseases, were also examined. In 3 snap-frozen tissue blocks for each SPMS and control case, the presence and extent of demyelination and characterization of CL activity were determined by immunostaining with monoclonal antibodies for myelin oligodendrocyte glycoprotein (MOG) and major histocompatibility complex (MHC class II). CLs were classified, as previously described into leukocortical type I, intracortical type II, or subpial type III.

In Vivo Study Population
Thirty-six patients with MS having at least 1 MR imaging-visible CL were studied (Table 2). Twenty-one had relapsing-remitting MS (RRMS), and 15 had secondary-progressive MS. At study entry, most patients were under immunomodulatory therapy: Twelve were treated with interferon 1a/b1b or glatiramer acetate; 6, with fingolimod; 4, with natalizumab; 3, with azathioprine; and 3, with dimethyl fumarate; and 8 were untreated.

Each patient was assessed with the Expanded Disability Status Scale and underwent 3T MR imaging as described below. The study was approved by the University of Verona ethics committee, and informed consent was obtained from all patients.

MR Imaging Acquisition and Analysis
All patients were scanned with an Achieva 3T MR imaging scanner (Philips Healthcare, Best, the Netherlands). Isotropic 3D DIR (1 × 1 × 1 mm, 10 minutes 49 seconds), 3D T1-MPRAGE (1 × 1 × 1 mm, 5 minutes 50 seconds), and 3D EPI-SWI (0.55 × 0.55 × 0.55 mm, 5 minutes 51 seconds) images were acquired. Quantification of susceptibility maps was performed by using the recently introduced total generalized variation framework.

After coregistration with 3D EPI, 3D DIR images were visually

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<table>
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<th>Disease</th>
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<td>M/68</td>
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<td>M/51</td>
<td>22</td>
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<td>C48</td>
<td>M/68</td>
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</tr>
<tr>
<td>C54</td>
<td>M/66</td>
<td>16</td>
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The appearance of CLs (identified with the DIR) on the QSM map was then evaluated. Each lesion was manually segmented on the QSM. T1 was segmented to obtain a normal-appearing gray matter (NAGM) map. A threshold of \( P > .05 \) was used to ensure including mainly GM. The NAGM mask was then moved to the QSM space. The obtained GM mask was then used to segment a portion of the NAGM surrounding the lesion. Each CL mask was dilated (a circle of 7 pixels was used as the kernel) and was used to reduce the whole GM mask to the surrounding NAGM tissue. The surrounding mask was then used to segment a portion of the NAGM surrounding the lesion. Each CL was identified following the recent recommendations for CL scoring in patients with MS. All DIR images were assessed by consensus of experienced observers who were blinded to patient identity.

The number of hyperintense CLs was higher in RRMS (mean, 6.3; range, 0–14) compared with SPMS (mean, 2.2; range, 0–5), while the number of hypointense CLs was higher in patients with SPMS (mean, 0.8 ± 1.1; range, 0–4 in RRMS; mean, 3.5 ± 3.5; range, 0–11 in SPMS). Indeed, in RRMS, 61.4% of CLs were hyperintense and 23.4% were hypointense, whereas in SPMS, 40.7% of CLs were hyperintense and 48.5% were hypointense.

### RESULTS

#### Neuropathologic Analysis of Microglia Activity in CLs

To evaluate the microglia/macrophage activation in the different types of CLs, we performed immunostaining for MHC class II and CD68 on tissue blocks from 16 SPMS cases. The activity of all 127 CLs identified in the examined SPMS cases was analyzed: the largest proportion (45.0%) was chronic active, identified as cortical areas with MHC-II+ cells mainly localized at the lesion edge (Fig 1A), in agreement with previous studies. This type of CL was more abundant compared with both chronic inactive (25.9%), with very low MHC-II+ cell density through the entire lesion, and active (29.1%) CLs, characterized by numerous MHC-II+ lesions in the lesion core and borders (Fig 1A).

Examination of the inflammatory activity of large CLs (Fig 1B) showed that MHC class II immunostaining was mainly restricted to activated microglia with ramified morphology, with a higher density in the most external cortical layers close to the pial membrane in subpial type III CLs (Fig 1C) or close to the WM portion in type I CLs (Fig 1D), either in actively demyelinating or chronic active lesions. In chronic inactive lesions, a lower density of MHC-II+ cells was present, scattered in all demyelinated areas (Fig 1E-1F). Furthermore, for each of the examined patients with MS, all the different types of CL activity were found. Large CLs (Fig 1B) sometimes contained, simultaneously, rims of activated microglia either close to the pial surface (Fig 1C) or toward the WM lesion border (Fig 1D).

#### Imaging Data

Two hundred fifty-five CLs were identified in the living population. On the basis of the DIR sequence, 126 CLs were pure intracortical and 129 CLs were leukocortical; on the basis of QSM, 131 were hyperintense, 33 were isointense, and 84 were hypointense (Fig 2). Seven CLs were discarded from the analysis because discordant results of the test between nQSMsurr and nQSMcontra were found. Among the 36 patients with MS, 32 showed at least 1 QSM-hyperintense CL, 16 showed at least 1 QSM-isointense CL, and 23 showed at least 1 QSM-hypointense CL. Twenty-six patients (72.2%) showed at least 2 QSM subtypes of CLs, and 11 patients showed all QSM subtypes of CLs at the same time. Table 2 shows the differences between intracortical and mixed GM/WM lesions.

The number of hyperintense CLs was higher in RRMS (mean, 4.1 ± 1.4; range, 0–14) compared with SPMS (mean, 2.2 ± 1.3; range, 0–5), while the number of hypointense CLs was higher in patients with SPMS (mean, 0.8 ± 1.1; range, 0–4 in RRMS; mean, 3.5 ± 3.5; range, 0–11 in SPMS). Indeed, in RRMS, 61.4% of CLs were hyperintense and 23.4% were hypointense, whereas in SPMS, 40.7% of CLs were hyperintense and 48.5% were hypointense.

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**Table 2: Demographic, clinical, and QSM-related characteristics of the patient population**

<table>
<thead>
<tr>
<th></th>
<th>RRMS</th>
<th>SPMS</th>
<th>Whole Group</th>
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</thead>
<tbody>
<tr>
<td>No.</td>
<td>21</td>
<td>15</td>
<td>36</td>
</tr>
<tr>
<td>Age (yr)</td>
<td>36.2 ± 5.8</td>
<td>49.5 ± 9.2</td>
<td>40.5 ± 8.0</td>
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<tr>
<td>Disease duration (yr)</td>
<td>9.7 ± 6.3</td>
<td>16.9 ± 7.0</td>
<td>12.7 ± 7.5</td>
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<tr>
<td>Sex (F/M)</td>
<td>16:5</td>
<td>9:6</td>
<td>25:11</td>
</tr>
<tr>
<td>EDSS (mean) (range)</td>
<td>2.0 (1.0–5.5)</td>
<td>5.0 (4.0–7.0)</td>
<td>3.0 (1.0–7.0)</td>
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<tr>
<td>No. of intracortical lesions (mean)</td>
<td>4.5 ± 3.6</td>
<td>2.1 ± 2.6</td>
<td>3.5 ± 3.4</td>
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<tr>
<td>QSM-hyperintense</td>
<td>2.8 ± 2.3</td>
<td>0.9 ± 1.2</td>
<td>2.0 ± 2.1</td>
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<tr>
<td>QSM-isointense</td>
<td>0.6 ± 0.7</td>
<td>0.1 ± 0.4</td>
<td>0.4 ± 0.6</td>
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<tr>
<td>QSM-hypointense</td>
<td>1.1 ± 1.5</td>
<td>0.9 ± 1.8</td>
<td>1.0 ± 1.6</td>
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<tr>
<td>No. of leukocortical lesions (mean)</td>
<td>2.6 ± 3.1</td>
<td>5.0 ± 3.5</td>
<td>3.6 ± 3.5</td>
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<tr>
<td>QSM-hyperintense</td>
<td>1.5 ± 1.8</td>
<td>1.9 ± 1.5</td>
<td>1.7 ± 1.7</td>
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<tr>
<td>QSM-isointense</td>
<td>0.5 ± 0.9</td>
<td>0.6 ± 1.3</td>
<td>0.5 ± 1.1</td>
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<tr>
<td>QSM-hypointense</td>
<td>0.5 ± 0.9</td>
<td>2.4 ± 2.4</td>
<td>1.3 ± 1.9</td>
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<tr>
<td>No. of total lesions (mean)</td>
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<td>7.1 ± 5.0</td>
<td>7.1 ± 5.1</td>
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<tr>
<td>QSM-hyperintense</td>
<td>4.2 ± 3.4</td>
<td>2.8 ± 2.3</td>
<td>3.6 ± 3.0</td>
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<tr>
<td>QSM-isointense</td>
<td>1.0 ± 1.2</td>
<td>0.7 ± 1.3</td>
<td>0.9 ± 1.3</td>
</tr>
<tr>
<td>QSM-hypointense</td>
<td>1.6 ± 1.8</td>
<td>3.3 ± 3.7</td>
<td>2.3 ± 2.8</td>
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</table>
A moderate inverse correlation was observed between hyperintense intracortical CLs and disease duration ($r = -0.48$), while no correlation was observed between disease duration and hypointense or isointense CLs.

The total number of CLs correlated with EDSS ($r = 0.51$). In patients with RRMS, the correlation between both total QSM-hyperintense CLs and intracortical hyperintense CLs with EDSS was marked ($r = 0.63$ and 0.71, $P < .05$), while no correlation was observed between EDSS and QSM-hypointense or isointense CLs. Moreover, no correlation was found among hyperintense, hypointense, or isointense CLs and EDSS in patients with SPMS.

**Neuropathology/Imaging Comparison**

The QSM hyperintensity edge found in proximity to the pial surface in some examined QSM-hyperintense CLs mirrored the intense microglia activation found close by the pial surface in most subpial CLs observed in postmortem SPMS cases. Indeed, the intense band of microglia activation frequently found at the depth of cerebral sulci of active CLs (Fig 3A, B, -E, blue arrowheads), with a characteristic gradient of increased microglia density in the external cortical layers (I-III), decreasing toward the most inner cortical layers (Fig 3E), resembled the hyperintense QSM signal seen in Fig 3J (blue arrowheads). Similar intense MHC-II immunostaining was also observed along the surface of the cerebral sulci (Fig 3A, -C yellow arrowheads), especially in regions of active demyelinating subpial GM, comparable with the QSM hyperintensity edge found in proximity to the pial surface within cerebral sulci (Fig 3J, yellow arrowheads). Also, the QSM hyperintensity edge found at the WM/GM border seems to resemble the microglia/macrophage activation detected at the WM/GM interface in leukocortical lesions (Fig 3H). Most interesting, a marked difference in MHC-II+ density and morphology has been observed in WM and GM (Fig 3H) portions of type I CLs, suggesting functional differences in the resident microglia cells in these areas and a different ratio of microglia/monocyte-derived macrophages in the presence of a higher myelin density.

**DISCUSSION**

In this preliminary study, we combined neuropathologic analysis of GM lesion activity and a new imaging approach with 3D DIR and QSM, with the aim of better characterizing CLs in patients with RRMS and SPMS. Our results suggest that combined 3D DIR and QSM may reveal the area of intense microglia/macrophage activity and density in CLs as also detected by neuropathologic assessment. Furthermore, our data show that CLs are highly heterogeneous as revealed by their susceptibility maps. This heterogeneity was present across, and in some cases within, the lesions. These data are partially in contrast to a recent MR imaging study performed at 7T showing negative relative susceptibility values in 25 of the 27 CLs.22 However, the low number of patients and CLs analyzed might account for this discordance. Moreover, it is likely that the
susceptibility of CLs changes with time in relation to their activity as previously shown for WM lesions. Therefore, the low activity of the patients with MS and of the number of CLs detected at the time of the MR imaging might also help explain this contradiction.31-33

The high variability of CLs in inflammatory activity in patients with MS has been already described by several neuropathologic studies mainly based on SPMS patients with a relatively long disease duration3,8,12,34 or on material from biopsies of patients with early MS with tumefactive lesions.9 Despite these neuropathologic observations, there are no available data in vivo that characterize the inflammatory activity of CLs in the early phase of the disease. In a previous MR imaging study, we observed that a small subgroup of CLs was significantly hyperperfused; this finding suggested the presence of an active inflammatory process within cortical GM.35

Although the number of patients with SPMS was quite low for drawing definitive conclusions, QSM-hyperintense CLs appeared to be more frequent in the RRMS group, while QSM-hypointense lesions were more frequent in SPMS. This result, along with the moderate inverse correlation between the number of QSM-hyperintense CLs and disease duration, might suggest that QSM-hyperintense lesions characterize the more inflammatory and acute phases of the disease, while QSM-hypointense CLs characterize the chronic (less inflammatory) disease stage. Nevertheless, the presence of each type of QSM-visible CL in both RRMS and SPMS suggests that the heterogeneity of GM lesion types persists during the entire disease course. Of course, we are aware that the DIR sequence detects only a reduced portion of CLs36 and that the use of other sequences such as phase-sensitive inversion recovery might help improve their detection.37 However, recent comparative histologic/MR imaging studies have demonstrated that the “tip of the iceberg” detected by MR imaging and its “bulk” differ only in size and that the number of detectable CLs correlates with their overall number and with the overall percentage of cortical demyelination.36 The ability of QSM in detecting CLs itself has not been tested; however, we do not advise acquiring only the SWI sequence. Using the DIR or even the phase-sensitive inversion recovery sequence is recommended as a guide for detecting CLs.

Although the factors contributing to the susceptibility in the cortex are not fully established,38 initial studies of healthy brains both in vivo and postmortem have suggested that myelin and both heme and nonheme iron have dominant effects in conditioning the susceptibility map.39,40 In contrast to R2*, which increases proportionally to the concentration of both iron41 and myelin,42 the 2 substances have opposing effects on the magnetic

FIG 2. Illustration of examples of cortical lesion appearance on the quantitative susceptibility map obtained with the total generalized variation algorithm from the 3D EPI susceptibility-weighted scan. Each panel denoted by a letter is divided in 3 subpanels: 1) QSM with contoured CL (red line); 2) QSM with a superimposed CL (red), the NAGM reference tissue used in the CL classification (yellow), and NAGM obtained from the segmentation of the 3D T1 MPRAGE (cyan); 3) QSM with superimposed the 3D double inversion recovery sequence, where the CL detection and segmentation was performed. A–D, Hypointense lesions. E–H, Hyperintense lesions. The classification of CLs was performed with a 2-sided t test between QSM estimates in the lesion (with subtracted the median value of the reference QSM value in the NAGM) and a zero mean Gaussian distribution. When the t test was significant and the mean of nQSM was greater (lesser) than zero, the lesion was classified as hyperintense (hypointense).
FIG 3. Combined neuropathologic and MR imaging characterization of lesion activity in CLs. A–H, Immunohistochemistry staining of MOG (A–C and G) and MHC class II (D–F and H) in subpial (A–F) and leukocortical (G and H) CLs in postmortem MS brains. I–K, MR images of heterogeneous CLs in patients with MS in vivo; I, 3D double inversion recovery. J, Quantitative susceptibility mapping calculated with the total generalized variation algorithm. K, DIR superimposed on the QSM map. MOG and MHC-II immunostaining on serial sections show ongoing subpial demyelination (blue arrowheads in A, higher magnification in B) and intense pick of MHC-II+ glia activation (D, higher magnification in E) at the depth of a cerebral sulcus, resembling the QSM hyperintense signal (blue arrowheads in J) in similar regions. Concurrent complete subpial demyelination (yellow arrowheads in A, higher magnification in C) and intense peak of MHC-II+ glia activation (F) were also detected along the pial surface of the same cerebral sulcus (yellow arrowheads in A), respectively, resembling similar QSM hyperintense signal (yellow arrowheads in J). In type I leukocortical lesions, shown by MOG immunostaining (G), high occurrence and density of MHC-II+ activated microglia and macrophages (H) were observed at the GM/WM interface (green arrowheads in G and H). Higher magnification of the GM/WM interface (green arrowheads in H) reveals higher density of MHC-II+ cells mainly in proximity to inflammatory infiltrates (asterisks in G) within the WM border of the lesions, possibly corresponding to the frequent QSM hyperintense signal shown by green arrowheads in J. Original magnifications ×100 (A, D, and G), ×200 (B, C, E, F, and H).
susceptibility maps, because the proteins and lipids associated with myelin render its susceptibility diamagnetic.

Our neuropathologic data, in line with previous studies, confirmed that a significant proportion of type III and type I chronic active CLs are associated with a gradient of increased microglia activation in the most external cortical layer, close to the pial surface (in subpial type III CLs; Fig 3, yellow arrows) or to the WM interface (in leukocortical type I CLs; Fig 3, blue arrows). This finding was also observed in a recent neuropathologic study showing that in a subset of patients with MS with CLs, part of the CLs were characterized by a rim of activated microglia at their border. The examined postmortem SPMS cohort included MS cases that, at the time of death, had differences in both disease duration and clinical disability compared with the in vivo SPMS cases. These differences may explain the increased activity of the CLs detected in the postmortem SPMS cases.

Although our pathologic and MR imaging analyses are derived from 2 different cohorts of patients and therefore a direct comparison is only theoretic, on the basis of the similarity of the neuropathologic and MR images, we hypothesize that the QSM-hypointensity could be due to activated microglia/macrophages that phagocytose nonheme iron–rich cellular debris. As suggested by previous pathologic and imaging studies, the presence of a hyperintense rim in part of the detected CLs could indicate the presence of iron accumulation in microglia/macrophages at the subpial edge (in type III CLs) or at the WM/GM interface. If this is the case, the sensitivity of QSM in detecting activated microglia might explain some of the subpial hyperintensities observed with this sequence and not with the DIR sequence, which is usually not sensitive enough for subpial lesions (Fig 2). A recent study on a limited number of patients showed the capability of QSM to better discriminate intracortical and leukocortical lesions, thus suggesting that QSM could be useful in predicting and detecting early modifications of normal-appearing tissues. Nevertheless, further studies that combine postmortem MR imaging and neuropathologic analysis on the same cohort are currently in progress.

The QSM hyperintense signal frequently observed in proximity to, or within, the WM portion of the detected leukocortical lesions might resemble the similar increased density of MHC-II+ microglia found in the WM portion of the type I CLs. This resemblance might indicate the expanding inflammatory rim involved in the neuropathogenesis of type I CLs.

At least 2 alternative hypotheses might explain the presence of a hyperintense QSM signal: The hyperintense QSM signal could be generated by the increased iron release from intracellular deposits to extracellular spaces or by the iron leakage following blood-brain barrier damage in the acute phases of the disease. However, it is not usual within the cortical GM to have a BBB breakdown. The second hypothesis suggests that oligodendrocytes constitute an important source of iron, and changes in iron signal could be associated with a loss of oligodendrocytes and reduction of nonheme iron within oligodendrocytes and myelin in MS plaques and periplaque areas. Therefore, the detected QSM-hypointensity could be due to a decrease of nonheme tissue iron, which characterizes those inactive GM lesions without activated microglia and is more frequent in patients with a long disease duration.

Several observations have suggested that an abnormal deposition of iron might also play a significant role in the pathophysiology of GM damage in patients with MS as also is shown for many age-related degenerative disorders. While confirming the well-known relationship between CLs and disability, our data have extended these results, showing a significant correlation between QSM-hyperintense CLs (more than the hypointense CLs) and EDSS within the RRMS group. The reason for the lack of correlation in the SPMS group may be likely found in the lower number of patients included in the study and in the lower number of QSM-hyperintense CLs; this possibility is in line with the neuropathologic observation of the higher frequency of chronic active lesions in the examined postmortem SPMS brains. Nevertheless, the correlation with disability has not been found with hypointense or isointense CLs because of the possible detrimental role of chronic activated microglia. Further longitudinal studies on larger sample sizes may clarify this interesting clinical point.

CONCLUSIONS
Our study revealed that CLs in MS are heterogeneous during the entire course of the disease, not only among patients but also within the same patient and, sometimes, within the same lesion. These results corroborate the hypothesis of substantial clinical and immunopathologic heterogeneous patterns of MS inflammation/demyelination during the disease course. Moreover, the highest frequency of QSM-hyperintense CLs in RRMS and their correlation with EDSS in this group of patients seem to suggest a key role of activated microglia/macrophages in the early and acute phases of the disease. The combined use of QSM and DIR could be a useful tool to monitor the disease evolution and to identify those patients with higher rates of inflammatory cortical demyelination and associated neurodegeneration.

REFERENCES
7. van Horssen J, Brink BP, de Vries HE, et al. The blood-brain barrier

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