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# Midbrain-Hindbrain Involvement in Septo-Optic Dysplasia

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## ABSTRACT

**BACKGROUND AND PURPOSE:** Midbrain-hindbrain involvement in septo-optic dysplasia has not been well described, despite reported mutations of genes regulating brain stem patterning. We aimed to describe midbrain-hindbrain involvement in patients with septo-optic dysplasia and to identify possible clinical-neuroimaging correlations.

**MATERIALS AND METHODS:** Using MR imaging, we categorized 38 patients (21 males) based on the presence (group A, 21 patients) or absence (group B, 17 patients) of visible brain stem anomalies. We measured height and anteroposterior diameter of midbrain, pons, and medulla, anteroposterior midbrain/pons diameter (M/P ratio), vermian height, and tegmento-vermian angle, and compared the results with 114 healthy age-matched controls. Furthermore, patients were subdivided based on the type of midline anomalies. The associations between clinical and neuroradiological features were investigated. Post hoc tests were corrected according to Bonferroni adjustment ( $p_B$ ).

**RESULTS:** Patients with brain stem abnormalities had smaller anteroposterior pons diameter than controls ( $p_B < .0001$ ) and group B ( $p_B = .012$ ), higher M/P ratio than controls ( $p_B < .0001$ ) and group B ( $p_B < .0001$ ), and smaller anteroposterior medulla diameter ( $p_B = .001$ ), pontine height ( $p_B = .00072$ ), and vermian height ( $p_B = .0009$ ) than controls. Six of 21 patients in group A had thickened quadrigeminal plate, aqueductal stenosis, and hydrocephalus; 3 also had agenesis of the epithalamus. One patient had a short midbrain with long pons and large superior vermis. There was a statistically significant association between brain stem abnormalities and callosal dysgenesis ( $P = .011$ ) and developmental delay ( $P = .035$ ), respectively.

**CONCLUSION:** Midbrain-hindbrain abnormalities are a significant, albeit underrecognized, component of the septo-optic dysplasia spectrum, and are significantly associated with developmental delay in affected patients.

**ABBREVIATIONS:** AP = anteroposterior; MH = midbrain-hindbrain; SOD = septo-optic dysplasia

Septo-optic dysplasia (SOD) is a highly heterogeneous, usually sporadic condition with a multifactorial etiology. Currently, genetic abnormalities are identified only in 1% of patients, whereas the etiology remains unclear in most.<sup>1</sup> However, an increasing number of early developmental transcription factors and associated pathway genes have been implicated in the etiology of SOD, ie, *HESX1*, *SOX2*, *SOX3*, *OTX2*, *PROKR2*, *FGF1*, and

*FGF8*.<sup>2-4</sup> These genes are expressed in regions that determine the formation of forebrain and related midline structures, such as the hypothalamus and pituitary gland, and mutations in these genes are therefore associated with marked phenotypic heterogeneity. Classically, the diagnosis of SOD is made when 2 or more features of the classic triad of optic nerve hypoplasia, pituitary hormone abnormalities, and midline brain defects are present.<sup>1</sup> However, only one-third of patients present with all cardinal features of SOD.<sup>3</sup> Visual deficits due to optic nerve hypoplasia and ocular malformations are usually the first presenting sign, whereas endocrine dysfunction may become apparent later on.<sup>1</sup> The severity of pituitary-hypothalamic dysfunction is highly variable, ranging from isolated deficit of pituitary hormones to panhypopituitarism.<sup>3</sup> Neurologic deficits are common, and range from global retardation to focal deficits such as epilepsy or hemiparesis.<sup>5</sup>

Interestingly, *OTX2* and *FGF8* are also pivotal genes regulating brain stem patterning during early embryologic development.<sup>6-9</sup> In recent years, there has been a growing interest in the descrip-

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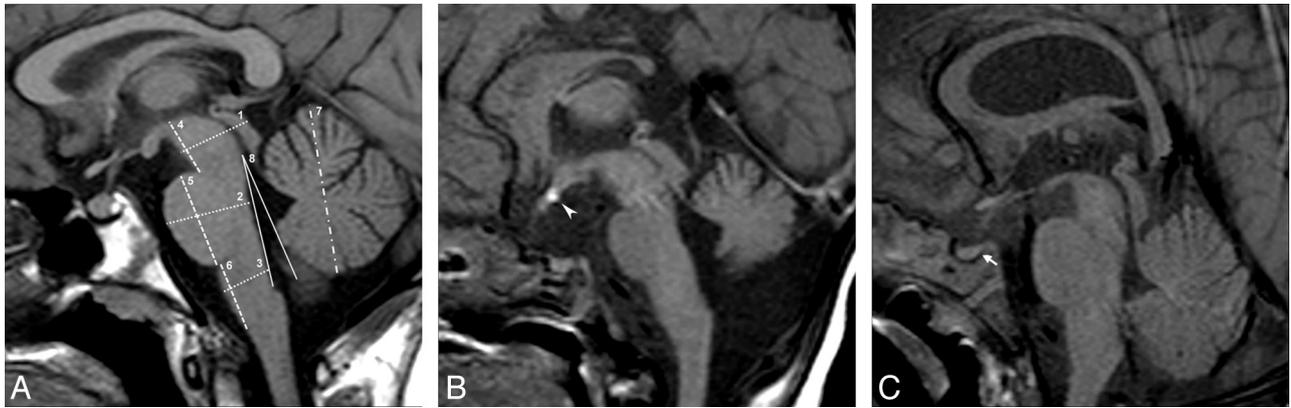
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**FIG 1.** A, Midsagittal T1-weighted image in a healthy child demonstrates how the measurements were performed: dotted lines indicate the AP diameter of the midbrain (1), pons (2), and medulla (3), dashed lines indicate the height of the midbrain (4), pons (5), and medulla (6), dash-dot line indicates the height of the vermis (7), and solid lines indicate the tectum-vermian angle (8). B, Midsagittal T1-weighted image in a patient with SOD with visible brain stem abnormalities (group A) reveals hypoplasia of the pons and cerebellar vermis. Note the hypoplasia of the pituitary gland with ectopic posterior lobe (arrowhead) and associated small corpus callosum. C, Midsagittal T1-weighted image in a patient with SOD without visible brain stem abnormalities (group B) reveals grossly normal brain stem and vermis. Note the absence of the normal posterior pituitary bright spot (arrow).

tion of congenital brain stem abnormalities. However, midbrain-hindbrain (MH) involvement in SOD has not been described so far. Here, we aimed to describe midbrain abnormalities and to identify possible correlations with clinical features in a relatively large group of patients with SOD.

## METHODS

### Subjects

This was a single-center retrospective case-control study. We reviewed the MR studies of 38 children who were diagnosed with SOD between 2006 and 2012 (21 males, mean age  $61.8 \pm 60.7$  months, and age range from 2 days to 18.3 years at the time of scan). The study received institutional review board approval. Exclusion criteria were uncertain SOD diagnosis and poor-quality or incomplete images. Complete neurologic evaluation was available in 32 of 38 patients. Griffiths Mental Development and Binet-Simon scales were used to define the developmental delay. Karyotype was performed in all patients. One patient harbored a mosaicism of trisomy chromosome 8 in skin fibroblast karyotypes, whereas the remainder presented normal karyotype. Comparative genomic hybridization by microarray was performed in 12 out of 38 patients, revealing *de novo* chromosomal aberrations in 2 cases: 1 duplication at 16p13.3p13.13 and 1 deletion at 14q22.1q23.1.

We selected 114 age- and sex-matched healthy children as a control group, ie, 3 controls for every patient with SOD. Controls underwent brain MR imaging for transient febrile convulsion or headaches. All were diagnosed as neurologically and developmentally normal by pediatric neurologists and had normal MR imaging findings.

### Imaging Analysis

All MR imaging examinations were performed on a 1.5 T magnet (Intera Achieva 2.6; Philips, Best, the Netherlands). Children under 6 years of age or uncooperative patients were sedated during examinations. All MR imaging studies included 3-mm-thick sagittal spin-echo T1-weighted images, axial and coronal FSE T2-weighted images, axial FLAIR images, and axial DWI. Additional sequences performed in patients with SOD included axial and

sagittal 0.6-mm-thick 3D driven equilibrium sequences, sagittal FSE T2-weighted images, and 3D turbo-field-echo T1-weighted images.

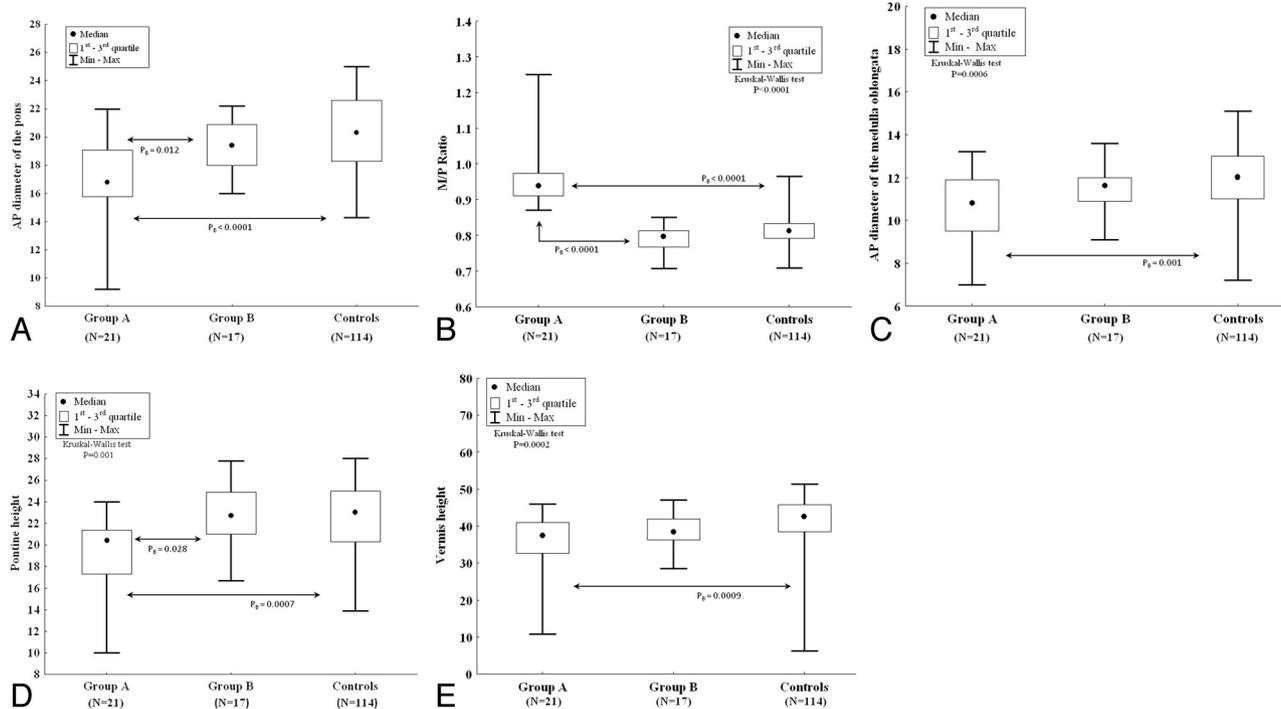
Patients with SOD were subjectively categorized based on the presence (group A, 21 patients) or absence (group B, 17 patients) of visible brain stem anomalies. Systematic visual analysis included midbrain dorsoventral and rostrocaudal size, tectal thickness, and collicular morphology, as well as dorsoventral and rostrocaudal size of the pons and medulla oblongata, and vermian size. We then measured the height and anteroposterior (AP) diameter of midbrain, pons, and medulla, the vermian height, and the tectum-vermian angle (Fig 1A). All measurements were performed by one pediatric neuroradiologist (M.S.) on midline sagittal T1-weighted images. We calculated the ratio of AP midbrain diameter to AP pons diameter (M/P ratio). We then compared the measurements and the M/P ratios of patients with SOD versus the controls.

Patients with SOD were also subdivided based on the presence and type of midline anomalies into 3 subtypes: group I (septal agenesis, 23 patients; complete in 13 cases and partial in 10 cases); group II (abnormal corpus callosum, 7 patients); and group III (no midline brain defects, 8 patients). Then, we investigated the association between brain stem and midline abnormalities.

Images were also analyzed for abnormalities involving the eyes, optic nerves, pituitary gland, olfactory bulbs, hypothalamus and hippocampi, as well as for cortical malformations including polymicrogyria, periventricular nodular heterotopia, and schizencephaly. Optic nerve hypoplasia was evaluated on coronal T2-weighted images at the level of the intraorbital segment, and was defined as an optic nerve caliber below 2 mm. Finally, we studied the association between brain stem abnormalities and the presence or absence of pituitary gland abnormalities, hippocampal malrotation, cortical malformations, developmental delay (irrespective of the degree of cognitive impairment), and epilepsy.

### Statistics

Descriptive statistics were firstly performed: categorical variables were reported in terms of absolute frequencies and percentages;



**FIG 2.** Boxplots of MH measurements in patients with SOD (group A and B) and healthy controls. AP diameter of the pons (A) and medulla (B), midbrain to pons ratio (C), height of the pons (D), and vermis (E) are shown.

quantitative variables were reported in terms of median values with 1st–3rd quartiles. Differences between groups were evaluated by means of the nonparametric ANOVA (Kruskal-Wallis test) and post hoc tests were corrected according to Bonferroni adjustment ( $p_B$ ). The association between categorical variables was evaluated by  $\chi^2$  test, or by the Fisher exact test in case of expected frequencies  $< 5$ . All tests were 2-sided and a  $P$  value less than .05 was considered as statistically significant. Data were analyzed by means of the statistical software Statistica 9 (StatSoft Corporation, Tulsa, Oklahoma) and STATA 7 (StataCorp, College Station, Texas).

## RESULTS

Clinical and neuroradiologic characteristics of patients with SOD are described in On-line Table 1. The AP pontine diameter in patients with SOD with brain stem abnormalities (group A, Fig. 1B) was significantly smaller (median: 16.8 mm; 1st–3rd quartiles: 15.8–19.1) than in the control group (median: 19.4 mm; 1st–3rd quartiles: 17.8–22.2,  $p_B < .0001$ ) and in patients with SOD with normal-appearing brain stem (group B, Fig. 1C) (median: 19.4 mm; 1st–3rd quartiles: 18.0–20.9,  $p_B = .012$ ) (Fig. 2A). No significant statistical difference was found in the AP diameter of midbrain between patients with SOD (both group A and B) and healthy controls. As a consequence, the M/P ratio was higher in patients with SOD with visible brain stem abnormalities (group A, median: 0.94; 1st–3rd quartiles: 0.91–0.97) compared with controls (median: 0.81; 1st–3rd quartiles: 0.79–0.84,  $p_B < .0001$ ) and to group B (median: 0.80; 1st–3rd quartiles: 0.77–0.81,  $p_B < .0001$ ) (Fig. 2B).

The AP diameter of the medulla in patients with SOD with brain stem abnormalities (group A) was significantly smaller (median: 10.8 mm; 1st–3rd quartiles: 9.5–11.9) (Fig. 1B) than in the

control group (median 11.8 mm; 1st–3rd quartiles: 10.6–13,  $p_B = .001$ ) (Fig. 2C).

The height of the pons (median: 20.4 mm; 1st–3rd quartiles: 17.3–21.4) and of the vermis (median: 37.4 mm; 1st–3rd quartiles: 32.7–41.0) was significantly shorter in patients with SOD with visible brain stem abnormalities (group A) than in controls (median: 22.0 mm; 1st–3rd quartiles: 19.3–25.0,  $p_B = .0007$  and median: 41.4 mm; 1st–3rd quartiles: 36.3–45.0,  $p_B = .0009$ , respectively) (Fig. 2D, -E).

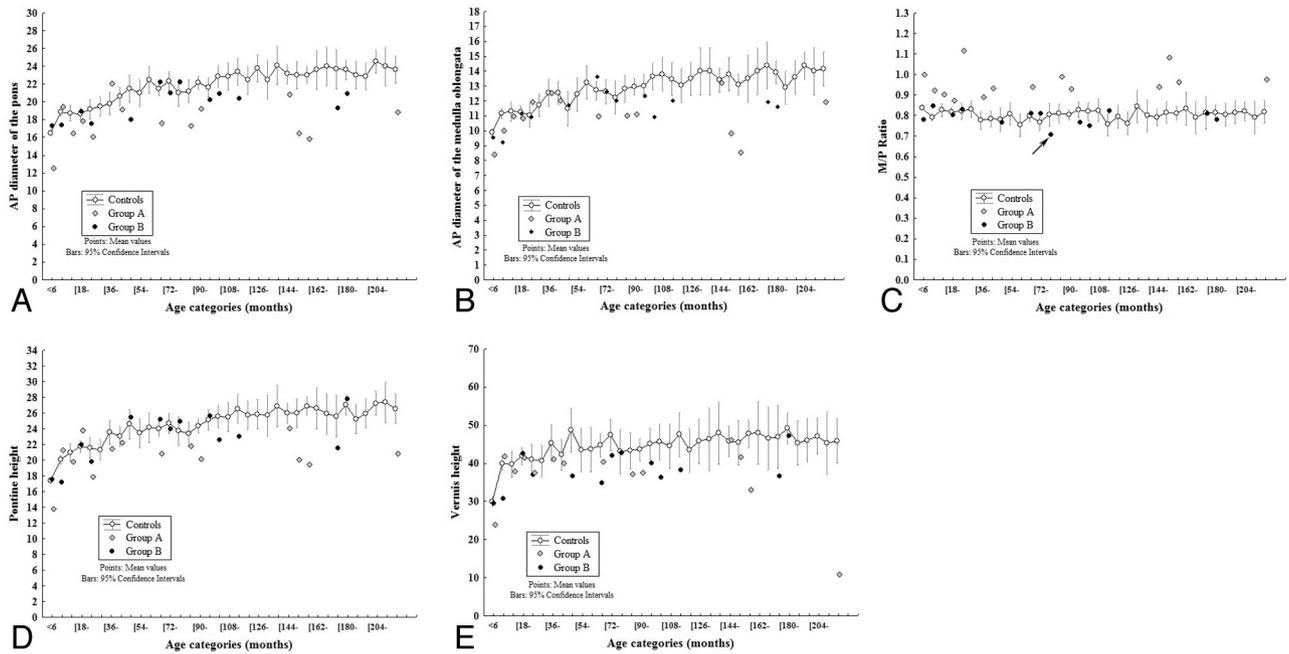
In patients of group A, the AP diameter of the pons and medulla and the pontine and vermian height were lower compared with the values observed in control subjects of the same age category (Fig. 3); in patients of group B, pontine and vermis height values tended to be lower compared with the values of control subjects of the same age category but these differences did not reach statistical significance.

No significant statistical difference was found regarding the tectum-vermian angle and height of midbrain and medulla between patients with SOD (both group A and B) and controls. There were no patients with isolated vermian anomalies.

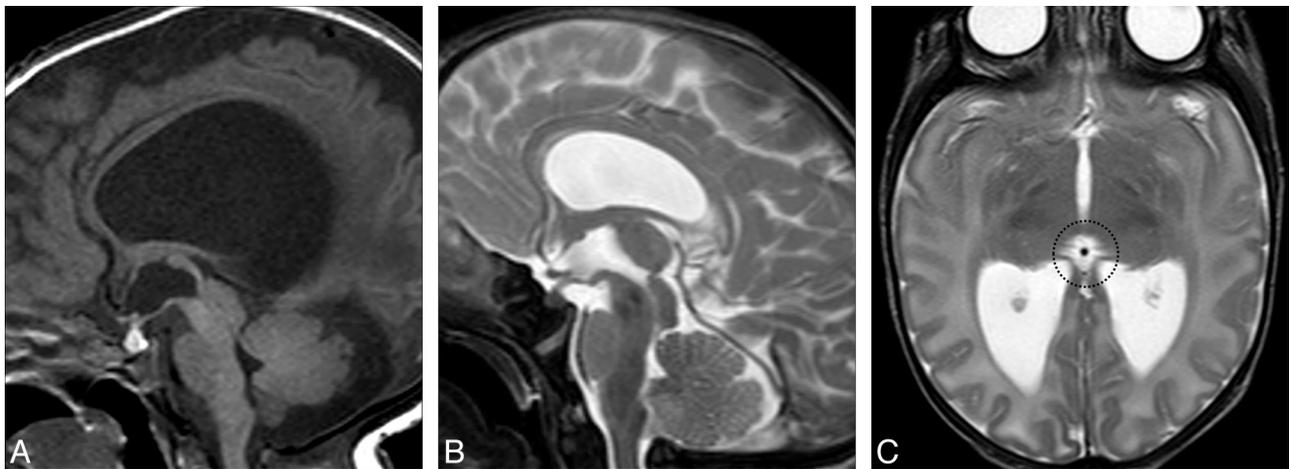
Six of 21 patients (28.6%) with brain stem abnormalities (group A) showed thickening of the quadrigeminal plate with aqueductal stenosis and triventricular hydrocephalus (Fig. 4A); 3 also had agenesis of the epithalamus (Fig. 4B, -C).

Callosal dysgenesis was present in 7 patients with SOD (group II) with complete agenesis in 3 cases and partial agenesis in 4 cases. Brain stem abnormalities were more frequent in patients with callosal dysgenesis (7/21, 33.3%) compared with those without visible brain stem abnormalities (0/17, 0%) ( $P = .011$ , On-line Table 2).

Finally, in 1 patient harboring 14q22.1-q23.1 deletion the



**FIG 3.** Scatterplots of MH measurements for age categories in patients with SOD and healthy controls. AP diameter of the pons (A) and medulla (B), midbrain to pons ratio (C), height of the pons (D), and vermian height (E) are shown. In healthy controls, the AP diameters and height of brain stem structures and vermian height showed an exponential growth curve. The growth spurt was steep until 16 months of age. Thereafter, it became less steep, reaching the adult level at about 8 years of age. On the other hand, the M/P ratios remained stable in controls over time, with a mean value of 0.82. In patients of group A, the AP diameter of the pons and medulla and the pontine and vermian height were lower compared with the values observed in control subjects of the same age category; in patients of group B, pontine and vermian height values tended to be lower compared with the values of control subjects of the same age category but these differences did not reach statistical significance. C, Note the lower M/P ratio in the patient harboring the 14q22.1-q23.1 deletion (arrow) compared with the corresponding values in age-matched controls.



**FIG 4.** A, Midline sagittal T1-weighted image demonstrates hypoplasia of the pons and small vermian, and enlarged quadrigeminal plate with secondary aqueductal stenosis and triventricular hydrocephalus. Midline sagittal (B) and axial (C) T2-weighted images in another patient reveal agenesis of the epithalamus including agenesis of the pineal gland (dotted circle), stria medullaris, and posterior commissure.

midbrain was smaller (height 11 mm and AP diameter 13.8 mm versus 13.5–16.2 mm and 16.6–17.2 mm minimal-maximal values of age-matched controls, respectively) compared with the pons, which appeared slightly elongated (pons height 25.4 mm versus 23.2–25.3 mm minimal-maximal values of age-matched controls) (Fig 5). The M/P ratio in this patient was lower (0.76) compared with the corresponding minimal-maximal values in age-matched controls (0.78–0.81) (Fig 3C).

Twenty-six of 38 patients presented with pituitary hypoplasia (68.4%) with additional ectopic posterior pituitary lobe in 7 cases and absent posterior bright spot in 2 cases. Hypothalamic hamar-

tomias were additionally present in 2 cases, whereas hypothalamic dysgenesis was evident in 3 other patients. Twenty-seven of 38 patients presented with optic nerve hypoplasia (71%) with microphthalmia in 3 cases and colobomas in 4 cases. Septal agenesis and optic nerve hypoplasia without any pituitary abnormality were present in 10 patients. Seventeen patients presented with hippocampal malrotation (44.7%) whereas 14 patients presented with a cortical malformation (36.8%). In particular, unilateral or bilateral polymicrogyria was present in 8 patients whereas 2 patients presented with an abnormal cortical gyration pattern. Periventricular nodular heterotopias were evident in 6 patients. Only 1 patient



**FIG 5.** Midsagittal T1-weighted image of the patient harboring the 14q22.1-q23.1 deletion shows a small and short midbrain with elongated pons and relatively larger superior portion of the cerebellar vermis. Note the small anterior pituitary gland with ectopic posterior lobe along the pituitary stalk (*arrowhead*).

had a unilateral schizencephaly. The association between these neuroradiologic abnormalities and brain stem malformation are summarized in On-line Table 3. We found a statistically significant association between pituitary and brain stem anomalies ( $P = .002$ ,  $\chi^2$ ). Seventeen of 32 patients (53.1%) were developmentally delayed; 5 of 32 patients (15.6%) had epilepsy. We found a statistically significant association between brain stem abnormalities and developmental delay ( $P = .035$ ;  $\chi^2$ ).

## DISCUSSION

The wide spectrum of imaging findings in SOD includes variable combinations of abnormalities of midline brain structures, the pituitary gland, optic nerves and eyes, olfactory bulbs, and other brain structures.<sup>10</sup> Midline brain defects classically consist of complete or partial absence of the septum pellucidum with fused midline fornices (60% of cases) and/or corpus callosum abnormalities, such as agenesis, dysplasia, or hypoplasia.<sup>1,10,11</sup> Pituitary gland malformations include anterior pituitary hypoplasia, ectopic posterior lobe, and/or thin or interrupted pituitary stalk.<sup>3</sup> Optic nerve hypoplasia is frequently associated with ocular anomalies such as coloboma, anophthalmia, and microphthalmia.<sup>12</sup> The olfactory bulbs may be absent or hypoplastic.<sup>13</sup> Other common associations include schizencephaly (so-called SOD-plus), gray matter heterotopia, polymicrogyria, and hippocampal malformations.<sup>14-16</sup>

MH abnormalities have not been described in patients with SOD to the best of our knowledge. Qualitative assessment of MH structures in the present study revealed abnormalities in 55% of patients, especially in the corpus callosum agenesis subgroup and in patients with pituitary gland abnormalities. Careful measurement and comparison of the height and AP diameter of midbrain, pons, medulla, and vermian height in patients with SOD versus healthy controls confirmed the visual analysis. Hypoplasia of the

pons was the major finding, leading to a significantly higher midbrain to pons ratio compared with controls. Moreover, hypoplasia of the vermis and medulla was also associated, resulting in a globally small hindbrain. The term “hypoplasia” refers to a condition of incomplete or arrested development in which an organ or part of it remains below the normal size. Generalized or segmental brain stem hypoplasia may be an isolated malformative feature or part of a more complex MH malformation, such as pontocerebellar hypoplasia,<sup>17</sup> horizontal gaze palsy with progressive scoliosis,<sup>18</sup> pontine tegmental cap dysplasia,<sup>19</sup> and anteroposterior or dorsoventral MH patterning defects.<sup>20,21</sup> More frequently, brain stem hypoplasia is identified in patients with cerebral malformations such as congenital muscular dystrophies,<sup>22</sup> lissencephalies,<sup>23</sup> or cerebral commissural anomalies.<sup>20</sup> As previously described in other MH malformations,<sup>24</sup> we found that cognitive impairment or developmental delay was more frequent in patients with SOD with small hindbrain, worsening the prognosis and impairing rehabilitative strategies.

Recently, mutations in *OTX2* and *FGF8* genes have been described in patients with SOD.<sup>2-4</sup> Interestingly, *OTX2* and *FGF8* play an important role both in the development of the anterior forebrain<sup>25</sup> and in the differentiation along the anteroposterior axis of the neural tube, with formation of the primary and secondary brain vesicles.<sup>26</sup> The latter process determines the correct regionalization of the forebrain (which rapidly divides into telencephalon and diencephalon), mesencephalon (midbrain), and hindbrain (which further divides into the rostral metencephalon, ie, pons and cerebellum, and caudal myelencephalon, ie, medulla oblongata). This delicate embryologic process requires the orderly expression and interaction of several signaling molecules and the formation of patterning centers such as the isthmus organizer at the MH junction.<sup>27</sup> In particular, *Otx2* is expressed in the midbrain both in mice<sup>28</sup> and humans,<sup>29</sup> whereas *Gbx2* is expressed in the anterior hindbrain, with a shared border at the level of IsO.<sup>30</sup> In rodent fetuses, the formation and normal location of IsO depends on a delicate interplay between the expression of *Otx2*, *Gbx2*,<sup>6,7</sup> and *Fgf8*.<sup>30</sup> Animal models have shown that repression of *Otx2* expression induces *Gbx2* formation to establish the location of the MH junction rostrally, resulting in a short midbrain and long pons. On the other hand, overexpression of *Otx2* shifts the MH junction caudally, resulting in an elongated midbrain with a small pons and vermis.<sup>27</sup> Similarly, alterations of *Pax6* or *En1/Pax2* will change the location of the diencephalic-mesencephalic junction.<sup>27</sup>

Remarkably, 6 patients with SOD spectrum and hypoplasia of hindbrain structures also showed hypertrophy of the mesencephalic tectum with consequent aqueductal stenosis and triventricular hydrocephalus. Broccoli et al<sup>6</sup> assessed the effect of ectopic expression of *Otx2* on MH development in mutant mice, and noticed that the superior and inferior colliculi appeared enlarged and were shifted posteriorly in all analyzed mutants. Furthermore, 3 patients also showed evidence of agenesis of the epithalamus (ie, stria medullaris, habenular trigone, pineal gland, and posterior commissure). Larsen et al<sup>29</sup> recently demonstrated that *Otx2* is expressed in the pineal gland at later stages in human fetuses. Gene ablation experiments in mice revealed that *Otx2* is a key regulatory gene for the development of the pineal gland.<sup>31</sup>

Furthermore, agenesis of the pineal gland has been described in patients with mutations of *PAX6* gene and congenital aniridia.<sup>32,33</sup> Intriguingly, *PAX6* is another important gene involved in the AP patterning of the brain. In particular, in murine and chick models, the correct formation of the diencephalic-mesencephalic junction depends on the interaction of Pax6 from the diencephalon and En1/Pax2 from the rostral mesencephalon, with the contributing role of Fgf8 that induces forebrain cells to secrete Pax6.<sup>27</sup> Webb et al<sup>34</sup> recently studied the 24-hour melatonin profiles in 6 patients with SOD and sleep disorders. Interestingly, 2 children appeared to produce virtually no melatonin throughout the 24-hour period of measurement. These findings raise the suspicion of a possible correlation between pineal gland agenesis and abnormally reduced melanin production in patients with SOD, which awaits confirmation in prospective studies.

Finally, 1 patient in this study harboring a de novo deletion at 14q22.1–23.1 presented with a short, small midbrain with elongated pons and enlarged superior cerebellar vermis, suggesting an abnormality of MH anteroposterior patterning due to rostral misplacement of the IsO.<sup>20–21</sup> This patient also had right anophthalmia and left colobomatous microphthalmia, small corpus callosum, bilateral hippocampal malformation, and hypoplasia of the anterior pituitary lobe with an ectopic posterior lobe. Interestingly, the 14q22.1–23.1 deletion includes the *Otx2* gene, supporting the hypothesis that loss of *Otx2* expression may determine rostral misplacement of the IsO also in humans. *OTX2* heterozygous mutations or deletions in humans have been linked to severe ocular malformations associated with forebrain abnormalities such as hippocampal and callosal malformations, and combined or multiple pituitary hormone deficiencies.<sup>12</sup> However, no careful description of MH structures has been provided so far in these patients. We reviewed the available brain MR midline sagittal images including the posterior cranial fossa of previously reported patients with *Otx2* mutations<sup>35–38</sup> or chromosome 14 deletions including *Otx2*,<sup>39–43</sup> and we recognized a similar midbrain hypoplasia with a long pons and large superior vermis in 5 patients.<sup>35,36,40,41,43</sup> We therefore suggest that MH abnormalities may have been underestimated in patients with *Otx2* mutations or deletion; further studies on larger series are awaited to address this hypothesis.

This study has several limitations, mainly including its retrospective design, the relatively small sample size, and the incomplete genetic assessment. Furthermore, the functional role and clinical implications of MH abnormalities in patients with SOD remain uncertain, and require further prospective studies.

## CONCLUSION

MH abnormalities are a significant, albeit under-recognized, component of the SOD spectrum. We demonstrated that the pons, medulla, and vermis were significantly hypoplastic in more than 50% of patients with SOD. Additional features included midbrain tectum enlargement and agenesis of the epithalamus. Patients with these abnormalities were more likely to present with developmental delay. Interestingly, these anomalies are similar to those reported in mutant mice with caudal shift of the MH junction due to ectopic *Otx2* expression

or Fgf8 mutations. Conversely, a short midbrain with an elongated pons and large superior vermis was observed in one SOD patient with 14q22.1–23.1 deletion, supporting the hypothesis that deletion of the *Otx2* gene may determine a rostral shift of the MH also in humans.

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## REFERENCES

1. Webb EA, Dattani MT. **Septo-optic dysplasia.** *Eur J Hum Genet* 2010;18:393–97
2. McCabe MJ, Gaston-Massuet C, Tziaferi V, et al. **Novel FGF8 mutations associated with recessive holoprosencephaly, craniofacial defects, and hypothalamo-pituitary dysfunction.** *J Clin Endocrinol Metab* 2011;96:1709–18
3. McCabe MJ, Alatzoglou KS, Dattani MT. **Septo-optic dysplasia and other midline defects: the role of transcription factors: HESX1 and beyond.** *Best Pract Res Clin Endocrinol Metab* 2011;25:115–24
4. Raivio T, Avbelj M, McCabe MJ, et al. **Genetic overlap in Kallmann syndrome, combined pituitary hormone deficiency, and septo-optic dysplasia.** *J Clin Endocrinol Metab* 2012;97:694–99
5. Signorini SG, Decio A, Fedeli C, et al. **Septo-optic dysplasia in childhood: the neurological, cognitive and neuro-ophthalmological perspective.** *Dev Med Child Neurol* 2012;54:1018–24
6. Broccoli V, Boncinelli E, Wurst W. **The caudal limit of *Otx2* expression positions the isthmic organizer.** *Nature* 1999;401:164–68
7. Millet S, Campbell K, Epstein DJ, et al. **A role for *Gbx2* in repression of *Otx2* and positioning the mid/hindbrain organizer.** *Nature* 1999;401:161–64
8. Sato T, Joyner AL. **The duration of Fgf8 isthmic organizer expression is key to patterning different tectal-isthmo-cerebellum structures.** *Development* 2009;136:3617–26
9. Beby F, Lamonerie T. **The homeobox gene *Otx2* in development and disease.** *Exp Eye Res* 2013;111:9–16
10. Severino M. **Septo-optic dysplasia.** In Rumboldt Z, Castillo M, Huang B, Rossi A, eds. *Brain Imaging With MRI and CT: An Image Pattern Approach.* New York: Cambridge University Press; 2012: 146–47
11. Belhocine O, André C, Kalifa G, et al. **Does asymptomatic septal agenesis exist? A review of 34 cases.** *Pediatr Radiol* 2005;35:410–18
12. Ragge NK, Brown AG, Poloschek CM, et al. **Heterozygous mutations of *OTX2* cause severe ocular malformations.** *Am J Hum Genet* 2005;76:1008–22
13. Levine LM, Bhatti MT, Mancuso AA. **Septo-optic dysplasia with olfactory tract and bulb hypoplasia.** *Journal of AAPOS* 2001;5:398–99
14. Barkovich AJ, Fram EK, Norman D. **Septo-optic dysplasia: MR imaging.** *Radiology* 1989;171:189–92
15. Miller SP, Shevell MI, Patenaude Y, et al. **Septo-optic dysplasia plus: a spectrum of malformations of cortical development.** *Neurology* 2000;54:1701–03
16. Riedl S, Vosahlo J, Battelino T, et al. **Refining clinical phenotypes in septo-optic dysplasia based on MRI findings.** *Eur J Pediatr* 2008;167: 1269–76
17. Namavar Y, Barth PG, Poll-The BT, et al. **Classification, diagnosis and potential mechanisms in pontocerebellar hypoplasia.** *Orphanet J Rare Dis* 2011;12:6:50
18. Rossi A, Catala M, Biancheri R, et al. **MR imaging of brain-stem**

- hypoplasia in horizontal gaze palsy with progressive scoliosis. *AJNR Am J Neuroradiol* 2004;25:1046–48
19. Briguglio M, Pinelli L, Giordano L, et al. Pontine tegmental cap dysplasia: developmental and cognitive outcome in three adolescent patients. *Orphanet J Rare Dis* 2011;6:36
  20. Barkovich AJ, Millen KJ, Dobyns WB. A developmental classification of malformations of the brainstem. *Ann Neurol* 2007;62:625–39
  21. Barkovich AJ, Millen KJ, Dobyns WB. A developmental and genetic classification for midbrain-hindbrain malformations. *Brain* 2009;132:3199–230
  22. Clement E, Mercuri E, Godfrey C, et al. Brain involvement in muscular dystrophies with defective dystroglycan glycosylation. *Ann Neurol* 2008;64:573–82
  23. Jissendi-Tchofo P, Kara S, Barkovich AJ. Midbrain-hindbrain involvement in lissencephalies. *Neurology* 2009;72:410–18
  24. Soto-Ares G, Joyes B, Lemaître MP, et al. MRI in children with mental retardation. *Pediatr Radiol* 2003;33:334–45
  25. Matsuo I, Kuratani S, Kimura C, et al. Mouse *Otx2* functions in the formation and patterning of rostral head. *Genes Dev* 1995;9:2646–58
  26. Acampora D, Gulisano M, Broccoli V, et al. *Otx* genes in brain morphogenesis. *Prog Neurobiol* 2001;64:69–95
  27. Nakamura H, Watanabe Y. Isthmus organizer and regionalization of the mesencephalon and metencephalon. *Int J Dev Biol* 2005;49:231–35
  28. Simeone A, Acampora D, Gulisano M, et al. Nested expression domains of four homeobox genes in developing rostral brain. *Nature* 1992;358:687–90
  29. Larsen KB, Lutterodt MC, Møllgård K, et al. Expression of the homeobox genes *OTX2* and *OTX1* in the early developing human brain. *J Histochem Cytochem* 2010;58:669–78
  30. Liu A, Joyner AL. *EN* and *GBX2* play essential roles downstream of *FGF8* in patterning the mouse mid/hindbrain region. *Development* 2001;128:181–91
  31. Nishida A, Furukawa A, Koike C, et al. *Otx2* homeobox gene controls retinal photoreceptor cell fate and pineal gland development. *Nat Neurosci* 2003;6:1255–63
  32. Abouzeid H, Youssef MA, ElShakankiri N, et al. *PAX6* aniridia and interhemispheric brain anomalies. *Mol Vis* 2009;15:2074–83
  33. Bamiou DE, Free SL, Sisodiya SM, et al. Auditory interhemispheric transfer deficits, hearing difficulties, and brain magnetic resonance imaging abnormalities in children with congenital aniridia due to *PAX6* mutations. *Arch Pediatr Adolesc Med* 2007;161:463–69
  34. Webb EA, O'Reilly MA, Orgill J, et al. Rest-activity disturbances in children with septo-optic dysplasia characterized by actigraphy and 24-hour plasma melatonin profiles. *J Clin Endocrinol Metab* 2010;95:198–203
  35. Ashkenazi-Hoffnung L, Lebenthal Y, Wyatt AW, et al. A novel loss-of-function mutation in *OTX2* in a patient with anophthalmia and isolated growth hormone deficiency. *Hum Genet* 2010;127:721–29
  36. Tajima T, Ohtake A, Hoshino M, et al. *OTX2* loss of function mutation causes anophthalmia and combined pituitary hormone deficiency with a small anterior and ectopic posterior pituitary. *J Clin Endocrinol Metab* 2009;94:314–19
  37. Wyatt A, Bakrania P, Bunyan DJ, et al. Novel heterozygous *OTX2* mutations and whole gene deletions in anophthalmia, microphthalmia and coloboma. *Hum Mutat* 2008;29:278–83
  38. Schilter KF, Schneider A, Bardakjian T, et al. *OTX2* microphthalmia syndrome: four novel mutations and delineation of a phenotype. *Clin Genet* 2011;79:158–68
  39. Elliott J, Maltby EL, Reynolds B. A case of deletion 14(q22.1->q22.3) associated with anophthalmia and pituitary abnormalities. *J Med Genet* 1993;30:251–52
  40. Lemyre E, Lemieux N, Decarie JC, et al. Del(14)(q22.1q23.2) in a patient with anophthalmia and pituitary hypoplasia. *Am J Med Genet* 1998;77:162–65
  41. Nolen LD, Amor D, Haywood A, et al. Deletion at 14q22–23 indicates a contiguous gene syndrome comprising anophthalmia, pituitary hypoplasia, and ear anomalies. *Am J Med Genet A* 2006;140:1711–18
  42. Hayashi S, Okamoto N, Makita Y, et al. Heterozygous deletion at 14q22.1-q22.3 including the *BMP4* gene in a patient with psychomotor retardation, congenital corneal opacity and feet polysyndactyly. *Am J Med Genet A* 2008;146:2905–10
  43. Bakrania P, Efthymiou M, Klein JC, et al. Mutations in *BMP4* cause eye, brain, and digit developmental anomalies: overlap between the *BMP4* and hedgehog signaling pathways. *Am J Hum Genet* 2008;82:304–19