
B L Horowitz, M V Chari, R James and R N Bryan

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MR of Intracranial Epidermoid Tumors: Correlation of In Vivo Imaging with In Vitro $^{13}$C Spectroscopy

We analyzed the MR findings of five patients with benign intracranial epithelial tumors, commonly called epidermoids. The neoplasms were categorized into two groups on the basis of T1-weighted MR signal intensity (relative to brain): high-signal-intensity masses (short $T_1$) and low-signal-intensity masses (long $T_1$). Surgical specimens were obtained and analyzed by means of $^{13}$C MR spectroscopy. Epidermoids with short $T_1$ values (white epidermoids) had a high lipid content comprising mixed triglycerides containing unsaturated fatty acid residues. Epidermoids with long $T_1$ values (black epidermoids) exhibited a much reduced lipid content with no triglycerides or fatty acids. There was evidence of trace amounts of cholesterol in the black epidermoids.

Our data indicate that epidermoids are a heterogeneous group of neoplasms that behave differently with $T_1$-weighted MR imaging and $^{13}$C MR spectroscopy. The combination of MR imaging and spectroscopy holds the potential of further elucidating the nature of epidermoids as well as of other forms of neoplasms.


Epidermoids (epidermoidoma, congenital or primary cholesteatoma, pearly tumor) are slow-growing, extraaxial lesions that are thought to arise from epithelial inclusions formed at the time of closure of the neural tube between the third and fifth week of fetal life [1–4]. Intracranially, epidermoids are most common in the cerebellopontine angle and parasellar areas. They may also be seen within the calvarium or skull base (intraosseous) and, rarely, are found intraaxially, within the cerebral hemispheres or ventricular system [5, 6]. Histopathologically, epidermoids are known to have a component of stratified squamous epithelium with variable degrees of keratinization. The squamous epithelium rests on an outer layer of connective tissue, which may be moderately vascular [7]. Epidermoids may be solid or cystic [6, 8] and are thought to contain high concentrations of both lipid and cholesterol [6, 9–12]. The outer surface of the neoplasm frequently exhibits an irregular, nodular texture with frondlike projections and a shiny mother-of-pearl appearance. Calcification may be present [4, 9, 13].

Classical imaging methods—such as plain films, polytomography, and, more recently, CT—may demonstrate sharply margined lytic bone lesions or bone erosion with scalloped, sclerotic borders. On CT, the classical appearance of an epidermoid is that of a low-density mass with Hounsfield units approximating that of water or even fat. There may be associated calcification, and, rarely, contrast enhancement. Reports are present in the literature, however, that document occasional epidermoids exhibiting hyperdensity prior to contrast administration [14, 15] as well as cases of such tumors with contrast enhancement [16].

Several authors [9, 17–19] have documented the MR imaging characteristics of epidermoids. Most commonly, these lesions have been noted to exhibit prolonged $T_1$ and $T_2$ values with $T_1$-weighted images demonstrating a CSF-intensity mass. However, additional cases have been noted in which these lesions have demonstrated shortened $T_1$ and prolonged $T_2$ values with high signal intensity on $T_1$-
weighted images [20, 21]. This suggests the presence of mobile protons in a lipid-type substance.

The purpose of this study was to analyze a group of intracranial epidermoids with MR and to correlate the imaging characteristics with the results of in vitro $^{13}$C MR spectroscopy on the excised operative specimens. It was hoped that such a correlative study would lead to a better understanding of the nature and origins of these neoplasms and particularly of their lipid components. Furthermore, such an approach in the future might allow both MR imaging and in vivo spectroscopy to more accurately define and histologically characterize various forms of neoplasms.

**Subjects and Methods**

Five patients with proved benign intracranial, extraaxial epidermoids were studied with clinical MR imaging. The excised surgical specimens were subjected to in vitro $^{13}$C MR spectroscopy. Additionally, all five patients had routine CT examinations during the course of their preoperative evaluation.

**MR imaging**

All patients were imaged at 0.5 T on a Siemens Magnetom unit. T1-weighted images were obtained with routine spin-echo pulse sequences with short TEs (16-35) and short TRs (300–500). Spin-density- and T2-weighted images were also obtained, but were less characteristic. T1 values for both types of tumors were determined on an IBM PC-10 instrument.

**$^{13}$C MR Spectroscopy**

All $^{13}$C MR spectra were run on a wide-bore FT-NMR spectrometer (Bruker AM 400, Bruker A.G., Karlsruhe, W. Germany) operating at 100.61 MHz with proton decoupling. Spectra were accumulated in 16K data points with a spectral width of 22 kHz and a recycle time of 2 sec. Equivalent amounts (1.0 g) of fresh, postsurgical tumor specimens were used for spectral analysis. The tissues themselves were investigated in 10-mm NMR tubes, without sample spinning. A concentric capillary containing D$_2$O served as the external lock in the case of the tissue determinations. The tissue FIDs (free induction decay) were processed with a line-broadening factor of 10 Hz and automatic baseline correction. In addition, 1.0 g of each neoplasm was subjected to extraction with 10 ml of a chloroform-methanol mixture (5:1), filtered and evaporated. The dried residue in each case...
was dissolved in 0.4 ml CHCl₃ for the spectral determinations in 5-
mm NMR tubes. The spectrum of the white epidermoids was refer-
cenced by assigning the chemical shift value of 14.1 ppm to the
highest frequency signal due to the terminal methyl groups of the
fatty acid residues within the triglycerides. The spectrum of the black
tumors was referenced by assigning the chemical shift value of 130
ppm to the broad olefinic resonance.

Results

White epidermoids (found in three patients) were charac-
terized surgically as being cystic with dense, adherent cap-
sules and on CT as having lucent, nonenhancing mass effect
with negative (fatty) Hounsfield numbers (Fig. 1A). On MR
imaging (Fig. 1B), they had bright signal due to short T₁
values (≈ 500). These epidermoids had a very high lipid
content and the 13C MR spectra exhibited an excellent signal/
noise ratio even after only 500 scans (Figs. 1C and 1D). The
predominant lipid component seems to be mixed triglycerides
on the basis of the intense resonances at 61.9 ppm and 70.0
ppm (peak intensity ratio 2:1) for the C-1, C-3, and C-2
carbons of the glycerol moiety, respectively, as well as the
relatively strong carbonyl resonance at 171.8 ppm. The chem-
ical shift of the carbonyl resonance also precludes the pres-
ence of free fatty acids in this tumor. The region at 115–135
ppm for olefinic carbon resonances showed several strong
signals (Figs. 1C and 1D), the chemical shifts of which indicate
the presence of polyunsaturated fatty acid residues in the
mixed triglyceride [22] present within the tumor. There was
no evidence in the 13C MR spectra of the white tumor and its
CHCl₃-MeOH extract for the presence of cholesterol or its
derivatives.

The black epidermoids (found in two patients) were found at
surgery to be the classical type Pearly tumor with a solid
consistency and with nodular and frondlike glistening excre-
cences. CT in these patients demonstrated a water-density
mass indistinguishable from the density of CSF (Fig. 2A).
They were hypointense on T₁-weighted images (Fig. 2B) with
long T₁ values (~ 1100). These epidermoids exhibited a much
lower lipid content than the white epidermoids, and an ac-

Fig. 2.—A, Axial CT scan of black epidermoid shows a relatively well-defined water-density
mass in perimesencephalic cistern, which com-
presses and displaces the brainstem.
B, Axial T₁-weighted MR image of black epi-
dermoid shows a hypointense, well-defined extraxial mass indenting and compressing the
brainstem. The hypointense epidermoid has rel-
atively long T₁ values of approximately 1100
msec.
C and D, 13C MR spectra of black epidermoid
(C) and chloroform-methanol extract of black epidermoid (D). The lack of signal at 70.0 ppm
for the C-2 of glycerol indicates an absence of
triglycerides. Strong resonances are seen in the
40–60 ppm region of chloroform-methanol ex-
tract as a result of small amounts of cholesterol.
ceptable signal/noise ratio in the $^{13}$C MR spectrum was achieved only after an overnight run (26,000 scans) (Figs. 2C and 2D). The lack of signal at 70.0 ppm for the C-2 of the glycerol moiety in triglycerides indicated an absence of this lipid in the black tumor. The region from 115 to 135 ppm for olefinic carbons showed only broad, ill-defined resonances at 120 ppm, 130 ppm, and 140 ppm, respectively. However, the region from 40 to 60 ppm exhibited strong resonances, which were absent in the spectrum of the white epidermoid. All the resonances for cholesterol were exhibited in the $^{13}$C MR spectrum of the CHCl$_3$-MeOH extract of the black epidermoid only.

**Discussion**

Benign intracranial epidermoids are a heterogeneous group of neoplasms that not only present a varied appearance on CT studies but also behave quite differently on MR imaging when T1-weighted partial saturation techniques are used. The underlying basis for these differences becomes apparent when $^{13}$C MR spectroscopy is used for the analysis of tumor samples. One type of epidermoid tumor is associated with negative Hounsfield numbers on CT and is white (short T1) with T1-weighted MR imaging. This group is cystic and is associated with a high lipid content with mixed triglycerides containing polyunsaturated fatty acids and no cholesterol. The second group of epidermoids consists of the classical pearly tumor with a water-density mass on CT and is black (long T1) on T1-weighted MR imaging. These tumors are solid and have no triglyceride. The presence of cholesterol was not obvious in the $^{13}$C MR spectrum of the tissue, though the CHCl$_3$-MeOH extract clearly showed all the signals corresponding to this steroid. While it is true that the differences noted between the black and white epidermoids on the T1-weighted MR scans could theoretically be due to a difference in proton concentration, such a difference would be reflected in the spin-density-weighted MR scans. No such differences were observed, indicating that the proton concentrations in the two tumors are similar. Thus, we feel confident in ascribing the difference to a lipid-dependent decrease in T1.

This study suggests an approach to the characterization of neoplasms based on both MR imaging and spectroscopy. The application of these methods holds the potential of both demonstrating the anatomy of the various forms of neoplasms as well as providing a biochemical analysis of the individual lesions.

**REFERENCES**

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