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BACKGROUND AND PURPOSE: Sensitivity, positive predictive value (PPV), and negative predictive value (NPV) of conventional MR imaging in predicting glioma grade are not high. Relative cerebral blood volume (rCBV) measurements derived from perfusion MR imaging and metabolite ratios from proton MR spectroscopy are useful in predicting glioma grade. We evaluated the sensitivity, specificity, PPV, and NPV of perfusion MR imaging and MR spectroscopy compared with conventional MR imaging in grading primary gliomas.

METHODS: One hundred sixty patients with a primary cerebral glioma underwent conventional MR imaging, dynamic contrast-enhanced T2*-weighted perfusion MR imaging, and proton MR spectroscopy. Gliomas were graded as low or high based on conventional MR imaging findings. The rCBV measurements were obtained from regions of maximum perfusion. Metabolite ratios (choline [Cho]/creatine [Cr], Cho/N-acetylaspartate [NAA], and NAA/Cr) were measured at a TE of 144 ms. Tumor grade determined with the three methods was then compared with that from histopathologic grading. Logistic regression and receiver operating characteristic analyses were performed to determine optimum thresholds for tumor grading. Sensitivity, specificity, PPV, and NPV for identifying high-grade gliomas were also calculated.

RESULTS: Sensitivity, specificity, PPV, and NPV for determining a high-grade glioma with conventional MR imaging were 72.5%, 65.0%, 86.1%, and 44.1%, respectively. Statistical analysis demonstrated a threshold value of 1.75 for rCBV to provide sensitivity, specificity, PPV, and NPV of 95.0%, 57.5%, 87.0%, and 79.3%, respectively. Threshold values of 1.08 and 1.56 for Cho/Cr and 0.75 and 1.60 for Cho/NAA provided the minimum C2 and C1 errors, respectively, for determining a high-grade glioma. The combination of rCBV, Cho/Cr, and Cho/NAA resulted in sensitivity, specificity, PPV, and NPV of 93.3%, 60.0%, 87.5%, and 75.0%, respectively. Significant differences were noted in the rCBV and Cho/Cr, Cho/NAA, and NAA/Cr ratios between low- and high-grade gliomas (\(P < .0001, .0121, .001, \) and \(.0038\), respectively).

CONCLUSION: The rCBV measurements and metabolite ratios both individually and in combination can increase the sensitivity and PPV when compared with conventional MR imaging alone in determining glioma grade. The rCBV measurements had the most superior diagnostic performance (either with or without metabolite ratios) in predicting glioma grade. Threshold values can be used in a clinical setting to evaluate tumors preoperatively for histologic grade and provide a means for guiding treatment and predicting postoperative patient outcome.

Prospective grading of primary cerebral gliomas is an endeavor with many difficulties but with significant clinical benefit. Despite recent advances in technology, chemotherapeutic agents, radiation therapy options, and surgical techniques, few inroads have been made into improving the survival of patients with malignant gliomas. The mean survival rate remains dismal, with fewer than 10% of patients with glioblas-
toma multiforme alive after 2 years. Two major limitations are associated with histopathologic grading of gliomas: inherent sampling error associated with stereotactic biopsy and inability to evaluate residual tumor tissue after cytoreductive surgery. Malignant gliomas are known to infiltrate the parenchyma following vascular channels of the white matter tracts (1). This may not be readily appreciated if there is no signal intensity abnormality or enhancement on conventional MR images. Hence, histopathologic grading of gliomas has disadvantages and intrinsic error. The advantage of state-of-the-art MR imaging techniques in evaluating cerebral gliomas is the ability to sample not only the entire lesion, but also the adjacent brain tissue for physiologic and metabolite alterations.

Conventional MR imaging with gadolinium-based contrast agents is an established and useful tool in the characterization of cerebral tumors (2–6). Current 1.5-T clinical MR systems provide excellent anatomic or morphologic imaging of gliomas. However, despite optimization of sequences and protocols, the classification and grading of gliomas with conventional MR imaging is sometimes unreliable, with the sensitivity for glioma grading ranging from 55.1% to 83.3% (7–11). Kondziolka et al (10) demonstrated a 50% false-positive rate in evaluating supratentorial gliomas.

Conventional MR imaging provides important information regarding contrast material enhancement, perienhancement edema, distant tumor foci, hemorrhage, necrosis, mass effect, and so on, which are all helpful in characterizing tumor aggressiveness and hence tumor grade. Dean et al (8) determined that mass effect and necrosis were the two most important predictors of tumor grade. However, often a high-grade glioma may be mistaken for a low-grade glioma when it demonstrates minimal edema, no contrast material enhancement, no necrosis, and no mass effect (Fig 1A and B). Conversely, low-grade gliomas can sometimes demonstrate peritumoral edema, contrast material enhancement, central necrosis, and mass effect and be mistaken for a high-grade glioma (Fig 2A and B). Conventional MR imaging readily provides evidence of contrast material enhancement, signifying blood-brain barrier breakdown, which is often associated with higher tumor grade. However, contrast material enhancement alone is not always accurate in predicting tumor grade. Ginsberg et al (12) demonstrated that lack of enhancement of supratentorial gliomas does not equate with low-grade gliomas. In another study, all low-grade tumors showed contrast material enhancement, but almost one-fifth of glioblastoma multiforme tumors did not (11). The peritumoral hyperintensity on conventional T2-weighted MR images...
is nonspecific, representing tumor infiltration, vasogenic edema, or both. Moreover, conventional MR imaging does not provide reliable information on tumor physiology such as microvasculature, angiogenesis, metabolism, micronecrosis, or cellularity, all of which are also important in determining tumor grade (13–15).

Advanced MR imaging techniques such as perfusion MR imaging and proton MR spectroscopy have found increasing utility in studying brain tumors. Relative cerebral blood volume (rCBV) maps and measurements have been shown to correlate reliably with tumor grade and histologic findings of increased tumor vascularity (11, 16–25). Recent reports regarding MR spectroscopy support its use as a powerful tool in tumor grading as well. Specifically, elevation in choline (Cho) with depression of N-acetylaspartate (NAA) is a reliable indicator of tumor. There is extensive literature demonstrating the metabolite ratios of Cho/creatine (Cr), NAA/Cr, and myo-inositol/Cr and the presence of lipids and lactate to be useful in grading tumors and predicting tumor malignancy (7, 26–56). There is certainly compelling evidence that MR spectroscopy provides important supplemental information to that of conventional MR imaging. The recent finding of a direct correlation between Cho and Ki-67 levels or cellular proliferative activity provides objective confirmation of the potential of MR spectroscopy in predicting tumor grade (57, 58).

To date, there have been some efforts to combine perfusion MR imaging and MR spectroscopic techniques in characterizing gliomas (59, 60). Investigators have also combined these two techniques to evaluate postoperative patients (61) or pediatric patients
with brain tumors (62). Importantly, despite the numerous reports and widespread interest in the characterization of gliomas with these advanced MR imaging techniques, the current clinical role of rCBV and metabolite ratios requires further determination (63). There are surprisingly few reports in the literature describing the false-positive and false-negative rates for glioma grading by using rCBV, metabolite ratios, or both, compared with that for conventional MR imaging. Defining the role of these advanced MR imaging techniques in clinical practice, in terms of the sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV), and determining whether perfusion MR imaging, MR spectroscopy, or the combination of the two techniques is more superior need further investigation.

We evaluated our experience in applying both of these advanced techniques in the study of primary cerebral gliomas and compared the sensitivity, specificity, PPV, and NPV of perfusion MR imaging and MR spectroscopy in glioma grading with those of conventional MR imaging. Our purpose was to provide objective data on the clinical utility of perfusion MR imaging and MR spectroscopy in glioma grading and also to proffer some quantitative guidelines for distinguishing low-grade from high-grade gliomas.

Methods

Patients and Histopathologic Analysis

Approval for this study was obtained from the Institutional Board of Research Associates, and informed consent was obtained from all patients. Retrospective analysis of our database of 728 patients who underwent conventional, perfusion, and spectroscopic MR imaging yielded 160 patients who underwent preoperative MR examinations and had histopathologic results for comparison. Conventional MR imaging examinations were evaluated as all preoperative. One hundred seventeen patients received their perfusion and spectroscopic imaging before surgery and 43 patients after stereotactic biopsy or partial resection, but the postoperative images demonstrated residual contrast material enhancement. The MR examinations were acquired from November 1999 to July 2002. The patients’ ages ranged from 4 to 82 years, with an mean of 43 years. There were 108 male and 52 female patients.

Histopathologic evaluation was performed by an experienced neuropathologist (D.Z.) and was based on a modified Ringertz’s three-tier classification of gliomas (64): grade 1, low-grade glioma; grade 2, anaplastic glioma; and grade 3, glioblastoma multiforme. The imaging classification was divided into two tiers. Anaplastic gliomas and glioblastoma multiforme were considered high-grade gliomas, and this group comprised 120 patients. The low-grade gliomas group comprised 40 patients.

Conventional MR Imaging

Imaging was performed with a 1.5-T unit (Vision or Symphony; Siemens AG, Erlangen, Germany). A localizing sagittal T1-weighted image was obtained followed by nonenhanced axial T1-weighted (600/14 [TR/TE]), axial fluid-fluid attenuation inversion-recovery (FLAIR, 9000/110/2500 [TR/TE/TI]), and T2-weighted (3400/119) images. Contrast material–enhanced axial T1-weighted imaging was performed after the acquisition of the perfusion MR imaging data. Two experienced board-certified neuroradiologists (S.Y., E.K.), blinded to the perfusion and MR spectroscopic results, reviewed the conventional MR images and graded each tumor according to the two-tier imaging grading system: low- versus high-grade gliomas. A consensus was reached on the conventional grading of each lesion based on eight criteria: contrast material enhancement, border definition, mass effect, signal intensity heterogeneity, hemorrhage, necrosis, degree of edema, and involvement of the corpus callosum or crossing the midline (8, 9).

Dynamic Contrast-Enhanced Perfusion MR Imaging

Dynamic contrast–enhanced T2*-weighted gradient-echo echo-planar images were acquired during the first pass of a standard dose (0.1 mmol/kg) bolus of gadopentetate dimeglumine (Magnevist; Berlex Laboratories, Wayne, NJ). Seven to 10 sections were selected for perfusion MR imaging through the tumor based on T2-weighted and FLAIR images. The methods for acquiring perfusion data from a set of dynamic contrast-enhanced echo-planar images and the precise algorithm for calculating rCBV have been previously described (11). Data processing was performed by using a Unix workstation with analytic programs developed in-house by using C and IDL programming languages. After construction of an rCBV color map to target regions of maximal abnormality, four region-of-interest measurements were obtained, and the maximum rCBV was recorded. This method has been demonstrated to provide the most optimal interobserver and intraobserver reproducibility (66). The standardized region of interest, which measured approximately 2–3 mm², was used in most measurements. The rCBV measurements were obtained by a neuroradiologist (G.L.) experienced with perfusion data acquisition at our institution, once again blinded to the conventional and MR spectroscopic findings.

Proton MR Spectroscopic Imaging

Multivoxel 2D proton chemical shift imaging (CSI) or spectroscopic imaging was performed after administration of gadopentetate dimeglumine. This is the standard protocol at our institution, as the literature seems to demonstrate that gadolinium has a negligible effect on metabolite ratios and peak areas (66–68). The volume of interest (VOI) was confirmed by obtaining half-Fourier acquisition single-shot turbo spin-echo images (5/6/500/1 [TR/TE/TI/NEX]). Ten sections with 5-mm section thickness were obtained in 1 minute 15 seconds in the axial, coronal, and sagittal planes for all patients. A volume selective 2D CSI sequence with 1500/144 was used for MR spectroscopic imaging. The hybrid multivoxel CSI technique uses a point-resolved spectroscopy (PRESS) double spin-echo sequence for preselection of a VOI that was usually defined to include the abnormality as well as normal-appearing brain tissue when possible. To prevent strong contribution to the spectra from subcutaneous fat signals, the VOI was completely enclosed within the brain and positioned at the center of the phase-encoded field of view, which was large enough to prevent wraparound artifact. A typical VOI consisted of an 8 × 8-cm region placed within a 16 × 16-cm-field of view on a 1.5–2-cm transverse section. A 16 × 16 phase-encoding matrix was used to obtain the 8 × 8 array of spectra in the VOI, with an in-plane resolution of 1 × 1 cm and a voxel size of 1 × 1 × 1.5 cm³ or 1 × 1 × 2 cm³, depending on the size of the lesion. The metabolite peaks were assigned as follows: Cho, 3.22 ppm; Cr, 3.02 ppm; NAA, 2.02 ppm; mobile lipids, 0.5–1.5 ppm. Lactate was identified at 1.33 ppm by its characteristic doublet that is caused by J modulation and inverted at TE of 144 ms. Metabolite ratios were obtained by a neuroradiologist (M.L.) experienced with spectroscopy and again blinded to the perfusion MR imaging and conventional MR imaging data. Maximal Cho/Cr and Cho/NAA ratios and minimum NAA/Cr ratios were obtained from spectral maps. To ensure quality control and acceptable quality of spectroscopic data, normal values for NAA/Cr and Cho/Cr were obtained in normal appearing white matter in the contralateral hemisphere.
Statistical Analysis

Sensitivity, specificity, PPV, and NPV were calculated for correct identification of high-grade gliomas. Hence, tumors classified as high grade and found at histologic examination to be high grade were considered true-positive findings; low-grade gliomas that were histologically confirmed as low grade were considered true-negative findings. For rCBV and metabolite ratios, receiver operating characteristic (ROC) curve analyses were used to evaluate the performance of simple diagnostic tests that declared a glioma to be high grade if and only if the relevant measure (eg, rCBV) for that patient was greater than or equal to some value $K$. This permitted a determination of the sensitivity, specificity, PPV, NPV, and total error associated with each measure as a function of the threshold $K$ used to identify high-grade gliomas. To determine potentially useful threshold values for rCBV, Cho/Cr, and Cho/NAA in differentiating low- from high-grade gliomas, threshold values were found that 1) minimized the observed number of tumor grade misclassifications ($C_2$ error = fraction of misclassified tumors) and 2) maximized the average of the observed sensitivity and specificity ($C_1$ error). Hence, $C_1 = 1 - (\text{sensitivity} + \text{specificity})/2$. The Mann-Whitney test was used to compare histologically verified low- and high-grade gliomas in terms of rCBV, Cho/Cr, Cho/NAA, and NAA/Cr. A $P$ value less than .05 was considered to indicate a statistically significant difference.

Results

The sensitivity, specificity, PPV, and NPV for determination of a high-grade glioma with conventional MR imaging were 72.5%, 65.0%, 86.1%, and 44.1%, respectively. For a minimum $C_2$ error, a threshold value of 1.75 for rCBV provided sensitivity, specificity, PPV, and NPV of 95.0%, 57.5%, 87.0%, and 79.3%, respectively. For a minimum $C_1$ error, a threshold value of 2.97 for rCBV provided sensitivity, specificity, PPV, and NPV of 72.5%, 87.5%, 94.6%, and 51.5%, respectively (Table 1).

A threshold value of 1.08 for Cho/Cr provided the minimum $C_2$ error and 97.5%, 12.5%, 77.0%, and 62.5% for the sensitivity, specificity, PPV, and NPV for determination of a high-grade glioma. A threshold value of 1.56 provided the same sensitivity and specificity as those of conventional MR imaging. A threshold value of 1.6 for Cho/NAA provided minimum $C_1$ error and 97.5%, 42.5%, 81.2%, and 39.6% for the sensitivity, specificity, PPV, and NPV for determination of a high-grade glioma. Threshold values that provided the same sensitivity and specificity as those of conventional MR imaging were 1.61 and 1.88, respectively (Table 2).

The combination of rCBV, Cho/Cr, and Cho/NAA resulted in sensitivity, specificity, PPV, and NPV of 93.3%, 60.0%, 87.5%, and 75%, respectively, for minimum $C_2$ error, and 70.8%, 92.5%, 96.6%, and 51.4%, respectively, for minimum $C_1$ error. For the same sensitivity as that of conventional MR imaging (72.5%), the specificity and PPV were higher, and for the same specificity as that of conventional MR imaging (65.0%), the sensitivity, PPV, and NPV were higher than those of conventional MR imaging alone when using the combination of rCBV, Cho/Cr, and Cho/NAA (Table 4, Fig 3).

The mean, standard deviation, and ranges for rCBV, Cho/Cr, Cho/NAA, and NAA/Cr are shown in Table 5. A statistically significant difference was noted in the rCBV, Cho/Cr, Cho/NAA, and NAA/Cr between low- and high-grade gliomas ($P < .0001,$

### Table 1: Threshold values for rCBV for differentiation between low- and high-grade gliomas

<table>
<thead>
<tr>
<th>Description</th>
<th>rCBV</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
<th>$C_2$ Error</th>
<th>$C_1$ Error</th>
</tr>
</thead>
<tbody>
<tr>
<td>Minimum $C_2$ Error*</td>
<td>1.75</td>
<td>95.0</td>
<td>57.5</td>
<td>87.0</td>
<td>79.3</td>
<td>14.4</td>
<td>23.8</td>
</tr>
<tr>
<td>Minimum $C_1$ Error†</td>
<td>2.97</td>
<td>72.5</td>
<td>87.5</td>
<td>94.6</td>
<td>51.5</td>
<td>23.8</td>
<td>20.0</td>
</tr>
<tr>
<td>Same sensitivity as cMRI</td>
<td>2.97</td>
<td>72.5</td>
<td>87.5</td>
<td>94.6</td>
<td>51.5</td>
<td>23.8</td>
<td>20.0</td>
</tr>
<tr>
<td>Same specificity as cMRI</td>
<td>2.18</td>
<td>87.5</td>
<td>65.0</td>
<td>88.2</td>
<td>63.4</td>
<td>18.1</td>
<td>23.0</td>
</tr>
</tbody>
</table>

* $C_2$ = the percentage of observed data points misclassified.
† $C_1 = 1 - (\text{sensitivity} + \text{specificity})/2$. This maximizes the average of sensitivity and specificity.

### Table 2: Threshold values for Cho/Cr ratio for differentiation between low- and high-grade gliomas

<table>
<thead>
<tr>
<th>Description</th>
<th>Cho/Cr</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
<th>$C_2$ Error</th>
<th>$C_1$ Error</th>
</tr>
</thead>
<tbody>
<tr>
<td>Minimum $C_2$ Error*</td>
<td>1.08</td>
<td>97.5</td>
<td>12.5</td>
<td>77.0</td>
<td>62.5</td>
<td>23.8</td>
<td>45.0</td>
</tr>
<tr>
<td>Minimum $C_1$ Error†</td>
<td>1.56</td>
<td>75.8</td>
<td>47.5</td>
<td>81.2</td>
<td>39.6</td>
<td>31.3</td>
<td>38.3</td>
</tr>
<tr>
<td>Same sensitivity as cMRI</td>
<td>1.61</td>
<td>72.5</td>
<td>50.0</td>
<td>81.3</td>
<td>37.7</td>
<td>33.1</td>
<td>38.8</td>
</tr>
<tr>
<td>Same specificity as cMRI</td>
<td>1.88</td>
<td>55.0</td>
<td>65.0</td>
<td>83.5</td>
<td>33.3</td>
<td>41.9</td>
<td>38.8</td>
</tr>
</tbody>
</table>

* $C_2$ = the percentage of observed data points misclassified.
† $C_1 = 1 - (\text{sensitivity} + \text{specificity})/2$. This maximizes the average of sensitivity and specificity.
Within the high-grade glioma group, there were 73 anaplastic gliomas (anaplastic astrocytomas, n = 26; anaplastic oligodendrogliomas, n = 7; anaplastic mixed oligoastrocytomas, n = 40) and 47 glioblastoma multiforme. No statistically significant difference was noted in the rCBV, Cho/Cr, Cho/NAA, and Cho/Cr ratios between the anaplastic gliomas and glioblastoma multiforme groups (P = .549, .302, .363, and .915, respectively).

**Discussion**

Current methods of grading gliomas have inherent limitations. The current reference standard of histopathologic grading can be inaccurate when biopsy samples are not taken from the most malignant tumor region or when the tumor is not completely resected. This is a particular problem with glioma because of the infiltrative proliferation of the tumor. Although histopathologic grading is often performed on the enhancing portion of the tumor, vascular networks in the peritumoral region serve as a path for tumoral infiltration along perivascular spaces. The region of highest vascularity and malignancy may then be within the so-called peritumoral or perienhancing region (22).

Radiologic grading of tumors with conventional MR imaging is not always accurate, with sensitivity in identifying high-grade gliomas ranging from 55.1% to 83.3% in other studies (7, 8, 11) and 72.5% in this study. Yet, accurate tumor grading has important implications for treatment planning: Patients with an erroneous diagnosis of high-grade glioma will undergo unnecessary adjuvant therapy; patients with an erroneous diagnosis of low-grade glioma will be treated conservatively, with concomitant morbidity and mortality.

Results of several previous studies suggest that rCBV measurements may improve grading. Sugahara et al (18) correlated maximal rCBV values histologically and angiographically in 30 patients, with mean values of 7.52, 5.84, and 1.26 for glioblastomas, anaplastic astrocytomas, and low-grade gliomas, respectively. Aronen et al (16) found mean maximal rCBV values of 3.64 and 1.11 in high- and low-grade glio-
mas, respectively (n = 19). Knopp et al (11) had similar mean maximal rCBV values of 5.07 and 1.44 in high- and low-grade gliomas, respectively (n = 29). These values are comparable to our findings, with mean maximal rCBV values of 5.18 and 2.14 for high- and low-grade gliomas, respectively (Table 5, n = 160). Lev and Rosen (23) used an rCBV threshold value of 1.5 in discriminating among 32 consecutive patients with glioma. Thirteen (100%) of 13 astrocytomas were correctly categorized as high-grade gliomas. Three of these did not enhance after administration of contrast material. Of the nine low-grade astrocytomas, seven were correctly classified. The sensitivity and specificity with use of an rCBV of 1.5 as a threshold value were 100% and 69%, respectively. This compares with the results from this study of 95.0% and 57.5% sensitivity and specificity, respectively, by using 1.75 as the threshold value. More recently, Shin et al (24) demonstrated mean rCBV ratios of 4.91 in high-grade gliomas and 2.00 in low-grade gliomas, in 17 patients; these ratios were similar to the ratios in this study of 5.18 and 2.14 in high- and low-grade gliomas, respectively. The threshold or cutoff value of 2.93 with use of ROC curve analysis is comparable to our value of 2.97 minimizing for C1 error (Table 1).

Besides vascular proliferation, cellularity, mitotic activity, nuclear pleomorphism, and necrosis are important criteria in histopathologic grading of gliomas. Ki-67 labeling is used in histologic examination as a marker for cellular proliferation. A higher rate of Ki-67–positive cells corresponds to greater malignancy in gliomas. Metabolite ratios, in particular Cho levels, have correlated with Ki-67 levels in gliomas (58). MR spectroscopic measurements of Cho/Cr and Cho/NAA ratios should therefore be helpful in the grading of gliomas.

However, to date, there have been few systematic attempts to compare the sensitivity, specificity, PPV, and NPV of perfusion MR imaging and MR spectroscopy with those of conventional MR imaging in glioma grading. To make a comparison, we determined threshold values from logistic regression and ROC analyses. However, it is not always clear what criteria should be used in determining an “optimum” threshold. For example, one could choose to minimize C1 error, which minimizes the average of the false-positive and false-negative error rates. This would be appropriate if the consequence of misclassifying low-grade gliomas is the same as that of misclassifying high-grade gliomas and the two are equally likely to be presented to you for classification. Alternatively, one could choose to minimize the C2 error, the total number of misclassified tumors observed in the data. This adjusts for a difference in the relative frequency of low- and high-grade gliomas in the patient population. In particular, if high-grade gliomas are much more prevalent, then a high misclassification rate of high-grade gliomas would result in a high total number of misclassified gliomas. In the real world, high-grade gliomas are more common than low-grade gliomas. Hence, choosing the threshold to minimize C2 error will tend to yield high sensitivity and relatively low specificity. These are the reasons for presenting two sets of results with threshold values for rCBV and metabolite ratios that minimize both C1 and C2 error values. For application in clinical practice, however, a threshold that minimizes C2 error is most appropriate (Tables 1–3).

The sensitivity of rCBV in glioma grading in our study was 95.0% (minimizing for C2 error), indicating a high true-positive rate and low false-negative rate. Hence, if the rCBV is above 1.75, there is a high probability that the tumor will be a high-grade glioma. Conversely, when rCBV is below 1.75, the tumor is unlikely to be high-grade glioma. However, the relatively low specificity means that false-positive rates are relatively high and true-negative rates are correspondingly low. In other words, some low-grade gliomas will be falsely identified as high-grade gliomas. However, this is the lesser of two evils. First, low-grade gliomas are relatively less common than high-grade gliomas, so fewer errors will be made in absolute terms. Second, a low-grade glioma misidentified as a high-grade glioma will be treated aggressively with some increase in morbidity. However, a high-grade glioma misidentified as a low-grade glioma will be treated conservatively, resulting in potential rapid death.

The high NPV (79.3%) is likewise a significant finding, as gliomas with low rCBV (<1.75) are unlikely to have high-grade components. Hence, it is an excellent tool for excluding the presence of a high-grade glioma (23), an issue that often confronts the neuroradiologist and neuropathologist. The low total error when using rCBV alone suggests that rCBV in itself is an accurate predictor of tumor grade, with a 14.4% chance of error when using 1.75 as the arbitrary threshold value. When one chooses threshold values that demonstrate the same sensitivity as that of conventional MR imaging (72.5%), the specificity, PPV, and NPV are superior to those of conventional MR imaging. Similarly, when one chooses threshold values that demonstrate the same specificity as that of conventional MR imaging (65.0%), the sensitivity, PPV, and NPV are superior to those of conventional MR imaging (Table 1).

The sensitivities (minimized for C2 error) of Cho/Cr and Cho/NAA of 97.5 and 96.7%, respectively, in this study confirm that metabolite ratios can be useful in determining tumor grade. However, the low specificities (12.5% for Cho/Cr and 10.0% for Cho/NAA) are due to the high levels for Cho that we, and others, have observed in some low-grade gliomas. Again, however, high sensitivity in identifying high-grade gliomas is more important than high specificity because of the relatively fewer cases of low-grade glioma and the more serious consequences of false-negative findings.

Despite the low specificities, a significant difference was noted in Cho/Cr, Cho/NAA, and NAA/Cr ratios (P = .012, .001, and .004, respectively) for differentiating between low- and high-grade gliomas (Table 5). A review of the literature, taking into
account differences in MR spectroscopic technique such as the choice of TE and method for determination of metabolite ratios, demonstrates that the mean maximal values obtained for Cho/Cr and Cho/NAA and mean minimum values for NAA/Cr (Table 5) in our study are comparable to previously published data in differentiating between low- and high-grade gliomas (7, 31, 45, 49, 59). Other data in the literature present metabolite levels and direct metabolite concentrations in a slightly different manner than presented in the current study, but they clearly also show differences in Cho levels between low- and high-grade gliomas (35, 54). In the current study, we had a number of reasons for using Cho/Cr and Cho/NAA ratios for assessing tumor grade. First, histopathologically, the linear correlation of Cho with Ki-67 labeling index or cellular proliferative activity suggests that Cho may be a strong predictor of tumor grade (57, 58). Second, although it may be possible to provide better discrimination combining other metabolites such as alanine, lipids, lactate, myo-inositol, glutamine, and glutamate, it is often not possible to perform MR spectroscopy at multiple TEs in a clinical setting to provide metabolites such as myo-inositol, glutamine, and glutamate detectable best at short TE. Third, to analyze multiple metabolites, linear discriminant analysis (50) or automated spectral analysis software (69) is usually necessary but not commercially available as yet.

One of the challenges in spectroscopy, even with current automated techniques, is obtaining reliable and reproducible intrapatient and interpatient data. To ensure quality control in the data, measurements of normal Cho/Cr and NAA/Cr levels were also obtained in the contralateral unaffected white matter as part of the MR spectroscopy multivoxel, 2D CSI measurement. Despite possible regional variations in metabolite ratios within the brain, it was reassuring to find that normal values for Cho/Cr and Cho/NAA obtained in the low- and high-grade glioma groups were almost identical. Normal Cho/Cr and NAA/Cr levels were 0.88 and 1.72 for low-grade gliomas and 0.87 and 1.73 for high-grade gliomas (Table 5). This not only provides a means for ensuring that the MR spectroscopic data are reliable, but also allows for further comparison between the abnormal data and normative data. In this study, the comparison was made only between low- and high-grade gliomas; however, a comparison between metabolite ratios within the tumor and normal voxels in the same patient is shown in Table 5.

The role of necrosis in glioma grading is important (70). The presence of necrosis is one important distinction between anaplastic astrocytomas and glioblastoma multiforme. In our series, lipid and lactate were found in 5.0% of low-grade gliomas and in 16.6% of high-grade gliomas. There is certainly a difference in finding lipids and lactate between the two groups, and although formal quantification and analyses were not performed in this study, lipids and lactate do correlate with necrosis in high-grade glioma and may also be useful in differentiating glioma grades (27, 45, 71). The frequency of elevated lipid and lactate in the high-grade glioma group was relatively low compared with that of previous studies (45, 71). This may be explained by the population bias in the sample of 73 anaplastic gliomas versus 47 glioblastomas. Furthermore, as this study did not focus on lipids and lactate, formal quantification of these metabolites was not performed, and hence small amounts of lipids and lactate that may be obscured by baseline noise may not be detected.

In a clinical setting, where decisions such as extent of tumoral resection and addition and dose of postoperative chemotherapy, radiation therapy, and interval of follow-up must be made, threshold values can be used as important supplementary information in the noninvasive, neuroradiologic grading of gliomas. A lesion that may have nonspecific conventional MR imaging findings (lack of contrast material enhancement, no mass effect, no necrosis, and no edema), but that demonstrates an rCBV value of 7.72 and Cho/NAA ratio of 2.60, for example, has high sensitivity and specificity for being a high-grade glioma (Fig 1). A comparative lesion (Fig 2) with conventional MR imaging findings suggestive of a high-grade glioma (contrast material enhancement, necrosis, mass effect, and edema) but that demonstrates an rCBV of 1.70 and a Cho/NAA ratio of 0.90 is more in keeping with a low-grade glioma. Hence, using these threshold values in cases that may have nonspecific conventional MR imaging findings increases our confidence in glioma grading.

Of further clinical interest is the differentiation between anaplastic gliomas and glioblastoma multiforme in the high-grade glioma group. As with our findings, previously published data in a group of 26 high-grade gliomas (11) also indicate no significant difference in rCBV between anaplastic astrocytomas and glioblastoma multiforme. Discriminating between these two groups has been demonstrated with in vitro (72) and in vivo (71, 73) MR spectroscopy. A limitation in our study is the perfusion and spectroscopic imaging of a number of patients (n = 43) after surgery. Although it is possible that surgery and postoperative changes may affect tumor vascularity and cellularity, we believe that because these patients only had either partial resection or biopsy, the final statistical analysis would not have been affected greatly by this. We examined the preoperative patients separately, and the results and conclusion did not differ significantly. Another potential limitation of our study is the semiquantification of metabolite ratios by using an internal reference method within the same voxel. The effects of tumor heterogeneity, regional differences in absolute and relative metabolite concentrations, and various pathologies on metabolite T2 relaxation times is difficult to assess. Hence, using Cr as a reference metabolite may increase variability and inaccuracy. Methods to normalize Cho to contralateral normal Cho or contralateral normal Cr as well as methods to provide absolute quantification of metabolites in gliomas have been described (59, 74). Normalizing our results to contralateral normal Cho did
not seem to improve statistical significance substantially; however, it would seem that absolute metabolite quantification methods should improve the sensitivity and possibly the specificity of MR spectroscopy (74). Finally, the lower sensitivity and specificity of Cho compared with rCBV suggests that although Cho levels may correlate directly with tumor cellularity, tumor cellularity may not correlate directly with tumor grade. Tumor grade seems to be more reliably correlated with necrosis, nuclear atypia, mitoses, and vascular hyperplasia (13); hence, rCBV may provide stronger correlation with tumor grade than does Cho.

This study demonstrates that rCBV and metabolite measurements can improve preoperative tumor grading. MR spectroscopy and perfusion MR imaging are useful adjuncts to conventional MR imaging in planning postoperative chemotherapy, antiangiogenic therapy, and radiation therapy (75–77). Since these techniques avoid some of the problems of sampling error associated with histopathologic examination, it is conceivable that such methods may provide a more accurate overall assessment of tumors. Demonstrating that rCBV and MR spectroscopy are sensitive techniques in this study is an important step in this regard. However, the question of whether one can improve the long-term outcome for patients with gliomas remains. Long-term outcome studies are required to determine whether rCBV, Cho levels, or pathology is the best predictor of patient outcome.

**Conclusion**

Preoperative grading of gliomas based on conventional MR imaging is often unreliable. Independently and in combination, rCBV measurements and Cho/Cr and Cho/NAA ratios can significantly improve the sensitivity and predictive values of preoperative glioma grading. The rCBV measurements are the best parameter for glioma grading. This is extremely important for determining the most optimal therapy regimen and the regularity and aggressiveness of postoperative follow-up and treatment. At our institution, perfusion MR imaging and MR spectroscopy not only have become a routine part of glioma imaging, but also have become an important adjunctive tool in providing information in gliomas and regions of the brain that may have eluded histopathologic assessment. Perfusion MR imaging and MR spectroscopy can potentially overcome the limitation of sampling error with histopathologic grading of tumor by the ability to sample the entire lesion noninvasively in vivo. Ongoing data collection from longitudinal studies is crucial to determine if rCBV and metabolite ratios can, in the long run, be comparable to histopathologic examination in predicting tumor behavior and patient prognosis.

**References**
