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In Vivo Experimental Intracranial Aneurysm Models: A Systematic Review

SUMMARY: Animal models are necessary to develop and test innovations in aneurysm therapy before clinical introduction. This review aims at identifying the most likely candidates for standardizing preclinical testing of aneurysm devices. We systematically searched electronic databases for publications on animal aneurysm models from 1961–2008 to assess the methodologic quality of the studies and collect data on the patency and angiographic and pathologic outcomes of treatments. There has been a steady increase in the annual number of publications with time. Species that were most frequently used were dogs, rabbits, and rodents, followed by swine. Most publications are single-laboratory studies with variables and poorly validated outcome measures, a small number of subjects, and limited standardization of techniques. The most appropriate models to test for recurrences after endovascular occlusion were the surgical bifurcation model in dogs, and the elastase-induced aneurysm model in rabbits. A standardized multicenter study is needed to improve the preclinical evaluation of endovascular devices in aneurysm therapy.

ABBREVIATIONS: Angio = angiography; ICA = intracranial aneurysms; IHC = immunohistochemistry; SEM = scanning electron microscope; w = width

Therapeutic innovations carry with them the promise of new treatments as well as the fear of new adverse events. Some form of preclinical screening test is necessary to minimize the risks for patients and to limit large-scale clinical investigations to the most promising devices. If initial development often involves in vitro experiments and bench testing, the last steps of device assessment before clinical introduction must take into account the complexity of the in vivo environment. Animal models are used for that purpose.

Naturally occurring intracranial aneurysms are rare in laboratory animals. Therefore, various models have been designed in many species, including mice, rats, rabbits, swine, sheep, dogs, and primates. Each model has advantages and limitations, and the choice will depend on the purpose. Experimental aneurysm models can be used for the following: 1) understanding the mechanisms involved in the initiation, progression, and rupture of aneurysms; 2) testing endovascular devices; and 3) training interventionists.

Using a variety of physiologic and pathologic manipulations, such as carotid ligation, renovascular hypertension, and lathyrism in rodents and primates, Hashimoto et al¹ developed 1 model that closely resembled the morphologic, histologic, and hemodynamic features of human intracranial aneurysms. One advantage of this model is the induction of cerebral aneurysms without direct manipulation of intracranial arteries.² These models could offer insight into the potential efficacy of pharmaceutical prevention of aneurysm formation and improved understanding of aneurysmal growth.

However, the small size of aneurysms in mice and rats render these models inappropriate for the testing of endovascular interventions.

Murine models are useful for looking into the biology of arterial aneurysms, far beyond what can be achieved in large animals, thanks to the availability of sophisticated molecular and cellular techniques, including a wide array of genetically modified mice strains. An effort to retain these advantages in testing human devices has led some teams to construct aneurysms on the lateral wall of the abdominal aorta of rats or mice by using microsurgical techniques,³ and others to simplify models by eliminating the need for an aneurysm altogether and studying the direct implantation of coils into normal arteries^{4,5} or stents and flow diverters to determine branch preservation. Even though simpler designs such as arterial occlusion models can provide important insight regarding fundamental vascular phenomena, such as recanalization after coil embolization,^{5,6} they cannot comprehensively anticipate the potential advantages and complications related to the use of devices in human aneurysms. Hence, the development and refinement of new endovascular therapies have mostly relied on larger aneurysm models such as those in rabbits, swine, and canines,^{7–11} induced by vessel ligations combined with enzymatic injury or surgically constructed from autologous venous pouches. These larger models will be the focus of this review.

Inspired by the pioneering work of German and Black (1954),¹² surgical constructions in rabbits, swine, and dogs varied in complexity, from lateral wall types by end-to-side suturing of a segment of vein to carotid arteries,^{13–15} to bifurcation aneurysms, anastomosing the right and left carotid arteries at the site of venous pouch implantation.^{8,16–19} (For a review, see Massoud et al 1994.²⁰) The elastase-induced aneurysm model, most frequently used in rabbits,^{21,22} has been modified in various ways.^{23,24} Our aim was to review in the literature the main features of these models and their strengths and weaknesses in an effort to identify those that could serve as the most appropriate final assessor of new aneurysm devices immediately before clinical use. We paid special attention to models assessing coil embolization.

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Materials and Methods

Search Strategy and Inclusion Criteria

We performed a computerized search strategy of Medline/PubMed (1961–2008) and EMBASE (1980–2008) for reports on animal models of aneurysms. We used the following keywords: aneurysm, saccular aneurysm, intracranial aneurysm, experimental aneurysm, animal models, swine, rabbit, dog, primate, mice, and rat in different combinations (by using the Boolean operator OR in conjunction with the Boolean operator AND). We performed a hand search in journals not indexed in PubMed and in bibliographies of included articles and review articles for additional studies. This method of cross-reference checking was continued until no further publications were found. We included studies that met all of the following criteria: 1) peer-reviewed articles, 2) original studies (not a letter, review article, conference abstract, or editorial), 3) English or French language, 4) a treatment (stents, coils, embolic agent) performed, 5) minimum number of 5 aneurysms, 6) minimal follow-up of 2 weeks, and 7) angiographic or pathologic outcomes.

The second author (O.N.) independently assessed the reproducibility of the search strategy and the eligibility of studies. In the case of disagreement, both observers (F.B. and O.N.) reviewed the article in question together until a consensus was reached.

Data were extracted to a data sheet, including the following: publication (first author's name, URL), aim of the study, species, the number of animals/aneurysms, aneurysm induction, outcomes, follow-up, compliance with animal welfare regulations, and embolization technique.

General Characteristics and Angiographic and Pathologic Outcomes

The absence of standardized end points precluded a meta-analysis. We collected general characteristics of the models (aneurysm size, neck morphology, patency of untreated aneurysms, angiographic occlusion rates, histologic techniques, and surgical complications) as claimed by the authors of the various source articles, and the results are given as a narrative report with references, without an attempt at quantification and cross-comparison.

Quality Assessment

The methodologic quality of the studies was scored against the following criteria (1 point each)²⁵: 1) compliance with regulations, 2) inclusion of a control groups, 3) aneurysm size, 4) long-term patency, 5) rate of occlusion/recurrences, 6) correlation between angiography and histology, 7) blinding of outcome assessment; 8) randomization; and 9) procedural complications. Results expressed as means \pm standard error of the mean were compared with analysis of variance, by using $P < .05$ as significant.

Results

Description of Studies

The initial search retrieved 571 publications (Fig 1). After viewing abstracts, we excluded 292 (51%); 57 articles were identified on hand searching of bibliographies and review articles, giving a total of 336 articles that were fully evaluated. We found an increasing number of articles per year published between 1961 and 2008 (Fig 2). Canine models were the most frequent (105 publications), followed by rabbits (90), rodents (83), swine (49), and primates (9).

After examination of the full texts, we excluded 205 articles (61%) on the basis of selection criteria, yielding 131 studies.

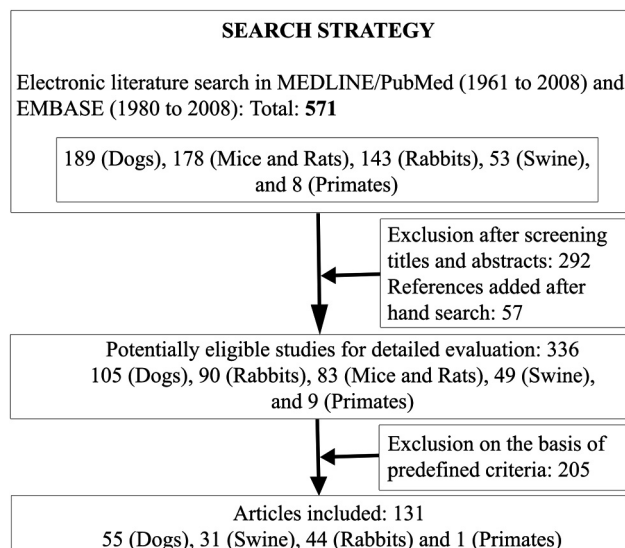


Fig 1. Search strategy and selection process for identifying articles on aneurysm models.

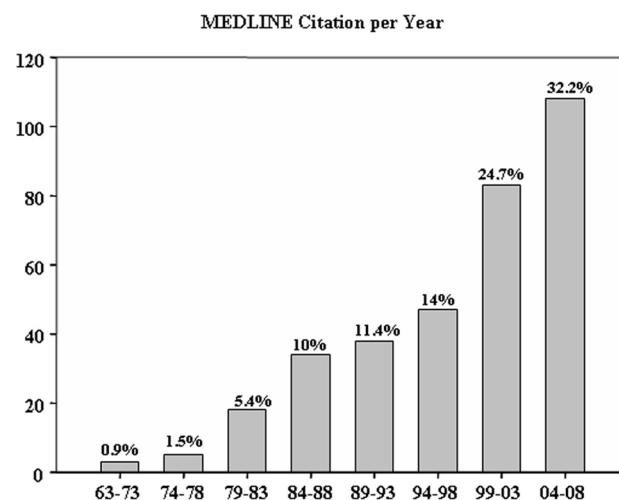


Fig 2. Number of publications per year reporting intracranial aneurysm models, as recruited in electronic data bases, showing a threefold output in the 2000s compared with the decades from 1963 to 1993.

There were 55 reports on dogs, 31 on swine, 44 on rabbits, and 1 study on primates (Table 1). We identified 3 different models: lateral wall (82 studies; 2143 aneurysms, 57%); bifurcation (31 studies; 848 aneurysms, 22%); and elastase-induced aneurysms (27 studies; 794 aneurysms, 21%). In the 55 reports on dogs, 34 (62%) were on the lateral wall aneurysms, 12 (22%) were on bifurcation aneurysms, and 9 (16%) studies were about both. In the 44 studies on rabbits, 27 (61%) were on elastase-induced aneurysms, 7 (16%) on lateral wall aneurysms, and 10 (23%) on bifurcation aneurysms.

Quality of Studies

Methodologic domains that were most frequently respected are summarized in Table 2. Few studies used randomization or blind assessment of outcomes or attempted to correlate pathologic end points with angiographic outcomes. There was no difference in quality scores among the lateral wall (5.07 ± 0.19 ; $n = 72$), bifurcation (5.41 ± 0.22 ; $n = 31$), and elastase-induced aneurysm models (5.56 ± 0.27 ; $n = 27$) ($P = .38$).

Table 1: Number of each type of aneurysm per species in the final set of included studies (n = 131)

Species (No. of Studies)	Lateral Wall, No. of Aneurysms (No. of Studies)	Bifurcation, No. of Aneurysms (No. of Studies)	Elastase, No. of Aneurysms (No. or % of Studies)
Dogs (55)	1051 (43) ^a	442 (21) ^a	0
Swine (31)	756	0	0
Rabbits (44)	313 (7)	406 (10)	794 (27)
Primates (1)	23	0	0
Total	2143 (57%)	848 (22%)	794 (21%)

^a Nine studies reported both lateral wall and bifurcation aneurysms.

Table 2: Results of quality assessment of included studies

Criteria	No. of Studies (%) (Total, 131)
Compliance with animal welfare regulations	90 (70)
Control groups included	91 (70)
Aneurysm size and morphology	95 (73)
Patency of untreated aneurysms	76 (59)
Angio and/or histologic scores	117 (90)
Correlation between angio and histologic results	34 (26)
Blinding of outcome assessment	27 (20)
Randomization of the experiment	19 (15)
Surgery and procedural complications	105 (81)

General Characteristics

The main characteristics of the 5 most frequent models are summarized in on-line Table 1.

Surgical constructions were most frequent: 2991 (57%) of 3785 aneurysms were surgically created, while 794 (21%) elastase-induced aneurysms were recorded. Canine models were most frequently used; many desirable features, including reliable anesthesia, large vessels, and excellent long-term survival, were reported.^{7,11,17,19,26}

Claims in favor of swine models included similarities to human physiology and coagulation systems; these models are commonly used in the field of cardiovascular research.²⁷

Surgical constructions have been used in all species, most frequently the lateral wall model ($n = 2143$, 57%). Bifurcation models included many variants.^{19,28,29} The ability to vary the anatomy of the bifurcation, aneurysm size, neck size, and fundus-to-neck ratio is a potential advantage of surgical models.²⁰

Creation of bifurcation aneurysms in rabbits is more difficult and perhaps less reliable, with a higher incidence of anesthesia-related deaths, procedure-related mortality, and parent vessel occlusions.^{8,30,31}

The elastase model was limited to rabbits, with advantages of easy in handling and long-term survival with aneurysms followed for >5 years.³² With some training, the results seem to be reproducible. Limitations included the uncontrolled size of aneurysms (potentially corrected by adjusting the ligation site),³³ sacrifice of the arterial access for each angiogram, and the small size of the animal, limiting the evaluation of large devices or techniques requiring multiple catheters.³⁴

Angiographic Outcomes

The lack of spontaneous thrombosis is an important criterion for a good model. Natural history studies of untreated aneurysms are few, however. In swine, the patency of untreated

aneurysm was evaluated in only 7/31 studies (22%); swine aneurysms, when left untreated, have a tendency for postoperative rupture or spontaneous thrombosis. This drawback necessitated immediate embolization following aneurysm construction.^{9,35} We noted spontaneous thrombosis between 2 and 7 weeks of lateral wall swine aneurysms ($n = 5$).^{13,35} Aneurysm patency was reported in 5/12 canine bifurcation aneurysm studies, in 6/9 on bifurcation and lateral wall aneurysms, and in 21/34 on lateral wall aneurysms. Canine lateral wall aneurysms may infrequently undergo thrombosis ($<10\%$), but most aneurysms (90%) remained patent during a follow-up period of ≥ 7 months.³⁶⁻³⁸ Bifurcation aneurysms enlarge during the first several weeks.³⁸ The incidence of spontaneous thrombosis is influenced by the ratio of the volume of the aneurysm to the area of the neck (ostium).¹² Modifications of the ostium can also minimize thrombosis.^{26,39}

In rabbits, 1/7 studies assessed the patency of surgically created lateral wall aneurysms, and 8/10, of bifurcation aneurysms. In the elastase model, 20/27 studies confirmed aneurysm patency. A long-term study showed neither spontaneous thrombosis nor significant change in dimensions 24 months after aneurysm creation.⁴⁰

The standard method for the evaluation of the treatment is angiography. The occlusion and recanalization rates found after embolization will, of course, depend on the device targeted by the publication.

Coil embolization of swine aneurysms is routinely followed by progressive and finally complete occlusion.³⁵ Residual necks and recurrences after coil embolization have been demonstrated in lateral wall aneurysms in dogs; however, recurrences are rare when lateral wall aneurysms are completely occluded initially.²⁸ Bifurcation aneurysm models in dogs and rabbits can show recurrences with variable frequencies, despite treatment with various types of devices.^{12,13,16,28,41-43} Angiographic recurrences are rarely documented after coil embolization of the rabbit elastase aneurysms, but "microrecurrences" (only seen at pathology) are common.^{10,44,45}

Pathologic Analyses of Treated Aneurysms

The pathology of aneurysms treated by embolization or stent placement is hampered by the presence of metallic or polymeric materials. One option is to embed the specimen in resins with the device in situ and resort to cutting-grinding thick sections, a method that precludes standard processing methods and advanced techniques such as immunohistochemistry or immunofluorescence. An alternative is to carefully dissect the device from the tissues and use standard paraffin embedding. A variety of ways to circumvent these difficulties have been published,⁴⁶⁻⁴⁸ but high-quality analyses of early fragile specimens (within days after embolization), which could have been mechanistically revealing, often remain problematic. Studies reporting pathologic techniques are summarized in Table 3. In the 120 studies that included histologic findings, only 23 (19%) used a qualitative or semiquantitative grading system developed by the authors.

A correlation between pathologic and angiographic outcomes has rarely been documented, except for canine bifurcation aneurysms.^{29,42,43} Among the 10 studies performed in bifurcation aneurysms in rabbits, 5 revealed a considerable discrepancy between radiologic and pathologic findings, with

Table 3: Number of studies reporting the different histologic techniques (tissue-processing procedure, conventional staining, IHC, SEM) and the use of scoring technique

Species, Aneurysm (No. of studies)	Studies with Histology, No. (%) ^a	Plastic Inclusion, No. (%) ^b	Paraffin Inclusion, No. (%) ^b	Stains, No. (%) ^b	IHC, No. (%) ^b	SEM, No. (%) ^b	Scoring Technique, No. (%) ^b
Canine, lateral wall (43) ^c	37 (86)	15 (40)	24 (65)	37 (100)	9 (24)	12 (32%)	4 (11)
Canine, bifurcation (21) ^c	19 (90)	8 (42)	11 (58)	19 (100)	8 (42)	4 (21%)	2 (10)
Swine, lateral wall (31)	26 (84)	10 (38)	19 (73)	25 (96)	8 (30)	4 (15%)	7 (27)
Rabbit, lateral wall (7)	4 (57)	0	4 (100)	4 (100)	3 (75)	1 (25%)	0
Rabbit, bifurcation (10)	8 (80)	2 (25)	6 (75)	8 (100)	1 (12)	3 (37%)	0
Rabbit, elastase (27)	26 (96)	14 (54)	17 (65)	26 (100)	5 (19)	4 (15%)	10 (38)
Total (139)	120 (86)	49 (41)	81 (67.5)	119 (99)	34 (28)	27 (22.5)	23 (19)

^a Percentage of studies in aneurysms of each animal species.^b Percentage of studies with histology.^c Nine studies reported on both lateral wall and bifurcation aneurysms.**Table 4: Primary histopathologic findings after coil embolization of experimental aneurysms**

Species, Aneurysm	Neck	Dome	Reaction Around and Inside Coils	Inflammation Reaction
Canine, lateral wall ^{7,41,52,57}	Neointima in the obliterated and recanalized aneurysms (170.00 ± 42.43 μm)	Variably organized connective tissue	Neointima and connective tissue tightly adhering to the coils	Minimal inflammatory reaction
Canine, bifurcation ^{28,29,43,52}	Very thin neointima (29.80 ± 24.35 μm); frequent endothelialized clefts	Little-to-no residual thrombus; organized vascularized fibrocellular tissue	Coils apparently bare or covered by a neointimal membrane of variable thickness	Minimal degree of inflammation
Swine, lateral wall ^{35,41,55,56}	Thick fibrous neointimal membrane >1000 μm	Dense fibrotic tissue	Neointima and connective tissue tightly adhering to the coils	Attenuated inflammatory reaction
Rabbit, bifurcation ^{8,15,18,30,31}	No endothelialization	No evidence of organized thrombus	Uncovered coils in the lumen and no fibrosis between the loops of coils	Severe chronic or no major inflammatory reaction
Rabbit, elastase ^{10,44-46,72}	Partial or complete thin neointimal coverage	Loose hypocellular vascularized connective tissue; no contractile cells and no collagen deposition	No endothelial layer covering the coils close to the aneurysm neck	Marked-to-mild localized chronic inflammatory infiltration

angiography overestimating the degree of occlusion in all cases.^{8,18,30,31} Microscopic recurrences have been described with the rabbit elastase model, but a correlation with angiographic recurrences is rarely possible.^{45,49,50}

Nevertheless, characteristics of the healing of experimental aneurysms, mainly gleaned from cross-species comparisons, have been described. These studies have uniformly revealed organization of the thrombus within the aneurysmal sac within 6 months of platinum coil embolization of surgically created aneurysms in canines and swine, as well as neointimal coverage of the ostium of occluded aneurysms.^{7,51-53} The most frequent descriptions of coil embolization of aneurysms are summarized in Table 4.

Aneurysms in swine have a tendency to heal, with or without embolization and with any material.^{9,13,35,54,55} Early inflammatory changes^{39,54,55} were followed by a centripetal myofibroblast infiltration, robust collagen deposition, and a thick neointima, which completely sealed the neck.⁴⁸ When all specimens showed permanent occlusions and favorable healing characteristics at some delayed time points, as seen in swine aneurysms, then some earlier timeframe was chosen, in hopes of detecting accelerated healing with some new material.⁵⁶

In the long term, treated canine lateral wall aneurysms are completely occluded in most cases.^{7,41,51-53,57} Canine bifurcation aneurysms have a propensity for recurrences, however; therefore, they can reproduce the problem found in human aneurysms.^{41,54} Nevertheless, organization of thrombus was found at 4 weeks,⁵⁷ whether aneurysms were occluded or showed early recurrences. At 3 months,^{28,29} recurrences were

found, despite the presence of a neointimal layer that was continuous with the endothelialized clefts, which are thought to be involved in the recanalization process.

Rabbits seem to have a lesser propensity for healing. There was no endothelialization across the aneurysm orifice and no evidence of organized thrombus, but a thickened wall was associated with chronic³¹ or no major inflammatory reaction⁸ in bifurcation aneurysms.

Platinum coil embolization of rabbit elastase-induced aneurysms was followed by relatively poor healing, through thrombus formation, granulated tissue organization, and, finally, loose connective-tissue formation with no contractile cell infiltration. Deposition of collagen was absent, and tissue coverage at the neck was relatively poor.^{10,44,57}

Discussion

The general ability of animal studies to predict future clinical success remains controversial. The debate has resurfaced recently with the repeated disappointments encountered in the development of drug therapies for stroke.⁵⁸ It remains difficult to choose which promising device should be prioritized and which should be subjected to long and expensive clinical trials, without good quality preclinical studies. We have focused our study on models designed to assess endovascular devices, a modest objective compared with more fundamental research on the growth and rupture of aneurysms, for which most recent models would be deficient or inappropriate.

We believe that animal models remain grossly underused. Many endovascular devices in current clinical use have never

been studied in published preclinical studies, and in the absence of randomized trials, many remain of unknown benefit. Animal studies have been more frequent in recent years, but there is no standardized testing required of an aneurysm device to be considered ready for a clinical application. Preclinical testing schemes typically include single-laboratory studies with various poorly validated outcomes and measures and small sample sizes. The main purpose of this review was to identify the best potential models that could serve such a standardizing role.

Some ideal features of a good aneurysm model include the following: minimal surgical and endovascular morbidity; similarity to human aneurysm shear stresses, hemodynamic forces, physical dimensions, perianeurysmal environment, and tissue responses; and stability without spontaneous thrombosis when untreated.⁵⁹ None of the available models offer all these features.

Different animal models target different purposes. For example, swine models, widely used in cardiology circles, have been thoroughly validated to assess the risk of in-stent stenosis. The propensity of this species for thrombosis and neointima formation, particularly useful to assess arterial stenoses after device implantation,²⁷ makes this model ill-fitted for discriminating the capacity of aneurysm-treatment devices to prevent recanalization or induce permanent occlusions. Reproducing the clinical problem that innovations are designed to address is a fundamental requirement of a model. Hence, lateral wall models in general, and swine models in particular, cannot predict the ability of new coils to improve long-term angiographic results.^{28,35}

Two models have repeatedly proved valuable in the assessment of aneurysmal devices: the canine bifurcation model and the rabbit elastase model. These models have not been thoroughly evaluated for repeatability of results between centers however. Future collaborative work should concentrate on this issue.

Any meaningful research must involve comparison between 2 treatments by using some end point criteria. Angiographic outcomes at 3 or 6 months can be compared in the canine bifurcation aneurysm models, and with such end points, the value of 1 embolic device over another was shown in the past.^{19,43,60} We must remember that even if better angiographic results in animals could be shown to predict better angiographic results in humans, these end points are surrogates for long-term efficacy in the prevention of ruptures.

The investigation of histologic phenomena that follow embolization has often been purely descriptive, but the meaning of these observations and their clinical relevance remain obscure when findings are not correlated, if not causally related, to a comparison between aneurysms that recur and those that do not.⁶¹ In the absence of an explanatory correlation between angiographic evolutions and pathologic findings, researchers have relied on a priori principles inspired from analogies with other vascular research fields. For example, incorporation into the vessel wall and neointima formation on stents, grafts, and stent-grafts⁶² have played a dominant role in the concept of healing after device implantation. Another potential justification for using pathologic findings as surrogate end points is human autopsy findings. Data on humans are quite limited.⁶³⁻⁶⁷ However, enduring occlusions after platinum coiling

have been associated with neointimal closure of the neck and organization of the clot within the aneurysmal sac,⁶⁷ while incomplete occlusions and recanalized aneurysms have been associated with unorganized thrombus and poor cellularity at the implant/parent artery interface.⁶⁸ Many investigators have published scales that offer a rational, if not a validated basis, for assessing the performance of an implant, by giving points for features considered favorable for healing, including neck coverage with neointima, thickness of neointimal layer, or clot organization at the dome.^{11,19,43-47,56,69} These semiquantitative scales have largely been confined to single-center experiences; they have not been reproduced or validated. No one, to our knowledge, has been able to show that these criteria predict better in vivo clinical performances, however. Finally, the absence of a proved link between the biologic responses of these models with those found in human aneurysms must be emphasized.

Conclusions

Currently, available animal models present multiple weaknesses, but preclinical testing of new devices remains a necessity. Candidate models most appropriate for this preclinical evaluation include the rabbit elastase and the canine bifurcation aneurysm models.

References

1. Hashimoto N, Handa H, Nagata I, et al. **Animal model of cerebral aneurysms: pathology and pathogenesis of induced cerebral aneurysms in rats.** *Neurol Res* 1984;6:33-40
2. Morimoto M, Miyamoto S, Mizoguchi A, et al. **Mouse model of cerebral aneurysm: experimental induction by renal hypertension and local hemodynamic changes.** *Stroke* 2002;33:1911-15
3. Frosen J, Marjamaa J, Myllarniemi M, et al. **Contribution of mural and bone marrow-derived neointimal cells to thrombus organization and wall remodeling in a microsurgical murine saccular aneurysm model.** *Neurosurgery* 2006;58:936-44
4. Raymond J, Lebel V, Ogoudikpe C, et al. **Recanalization of arterial thrombus, and inhibition with beta-radiation in a new murine carotid occlusion model: MRNA expression of angiopoietins, metalloproteinases, and their inhibitors.** *J Vasc Surg* 2004;40:1190-98
5. Abrahams JM, Forman MS, Grady MS, et al. **Biodegradable polyglycolide endovascular coils promote wall thickening and drug delivery in a rat aneurysm model.** *Neurosurgery* 2001;49:1187-93
6. Yasuda H, Kuroda S, Nanba R, et al. **A novel coating biomaterial for intracranial aneurysms: effects and safety in extra- and intracranial carotid artery.** *Neuropathology* 2005;25:66-76
7. Mawad ME, Mawad JK, Cartwright J, et al. **Long-term histopathologic changes in canine aneurysms embolized with Guglielmi detachable coils.** *AJNR Am J Neuroradiol* 1995;16:7-13
8. Reul J, Weis J, Spetzger U, et al. **Long-term angiographic and histopathologic findings in experimental aneurysms of the carotid bifurcation embolized with platinum and tungsten coils.** *AJNR Am J Neuroradiol* 1997;18:35-42
9. Murayama Y, Vinuela F, Suzuki Y, et al. **Development of the biologically active Guglielmi detachable coil for the treatment of cerebral aneurysms. Part II. An experimental study in a swine aneurysm model.** *AJNR Am J Neuroradiol* 1999;20:1992-99
10. Kallmes DF, Fujiwara NH, Yuen D, et al. **A collagen-based coil for embolization of saccular aneurysms in a New Zealand white rabbit model.** *AJNR Am J Neuroradiol* 2003;24:591-96
11. Raymond J, Guilbert F, Metcalfe A, et al. **Role of the endothelial lining in recurrences after coil embolization: prevention of recanalization by endothelial denudation.** *Stroke* 2004;35:1471-75
12. German W, Black S. **Experimental production of carotid aneurysms.** *N Engl J Med* 1954;250:104-06
13. Guglielmi G, Ji C, Massoud TF, et al. **Experimental saccular aneurysms. II. A new model in swine.** *Neuroradiology* 1994;36:547-50
14. Bavinszki G, al-Schameri A, Killer M, et al. **Experimental bifurcation aneurysm: a model for in vivo evaluation of endovascular techniques.** *Minim Invasive Neurosurg* 1998;41:129-32
15. Bavinszki G, Richling B, Binder BR, et al. **Histopathological findings in experimental aneurysms embolized with conventional and thrombogenic/anti-thrombolytic Guglielmi coils.** *Minim Invasive Neurosurg* 1999;42:167-74

16. Forrest MD, O'Reilly GV. Production of experimental aneurysms at a surgically created arterial bifurcation. *AJNR Am J Neuroradiol* 1989;10:400–02
17. Strother CM, Graves VB, Rappe A. Aneurysm hemodynamics: an experimental study. *AJNR Am J Neuroradiol* 1992;13:1089–95
18. Spetzger UJ, Reul J, Weis J. Endovascular coil embolization of microsurgically produced experimental bifurcation aneurysms in rabbits. *Surg Neurol* 1998;49:491–504
19. Raymond J, Salazkin I, Georganos S, et al. Endovascular treatment of experimental wide neck aneurysms: comparison of results using coils or cyanoacrylate with the assistance of an aneurysm neck bridge device. *AJNR Am J Neuroradiol* 2002;23:1710–16
20. Massoud TF, Guglielmi G, Ji C, et al. Experimental saccular aneurysms. I. Review of surgically-constructed models and their laboratory applications. *Neuroradiology* 1994;36:537–46
21. Cloft HJ, Altes TA, Marx WF, et al. Endovascular creation of an in vivo bifurcation aneurysm model in rabbits. *Radiology* 1999;213:223–28
22. Altes TA, Cloft HJ, Short JG, et al. 1999 ARRS Executive Council Award: creation of saccular aneurysms in the rabbit—a model suitable for testing endovascular devices. *AJR Am J Roentgenol* 2000;174:349–54
23. Krings T, Moller-Hartmann W, Hans FJ, et al. A refined method for creating saccular aneurysms in the rabbit. *Neuroradiology* 2003;45:423–29
24. Hoh BL, Rabinov JD, Pryor JC, et al. A modified technique for using elastase to create saccular aneurysms in animals that histologically and hemodynamically resemble aneurysms in humans. *Acta Neurochir (Wien)* 2004;146:705–11
25. Macleod MR, O'Collins T, Howels DW, et al. Pooling of animal experimental data reveals influence of study design and publication bias. *Stroke* 2004;35:1203–08
26. Shin YS, Niimi Y, Yoshino Y, et al. Creation of four experimental aneurysms with different hemodynamics in one dog. *AJNR Am J Neuroradiol* 2005;26:1764–67
27. Kantor B, Ashai K, Holmes DR Jr, et al. The experimental animal models for assessing treatment of stenosis. *Cardiovasc Radiat Med* 1999;1:48–54
28. Raymond J, Berthelet F, Desfaits AC, et al. Cyanoacrylate embolization of experimental aneurysms. *AJNR Am J Neuroradiol* 2002;23:129–38
29. Raymond J, Darsaut T, Salazkin Y, et al. Mechanisms of occlusion and recanalization in canine carotid bifurcation aneurysms embolized with platinum coils: an alternative concept. *AJNR Am J Neuroradiol* 2008;29:745–52
30. Spetzger U, Reul J, Weis J, et al. Microsurgically produced bifurcation aneurysms in a rabbit model for endovascular coil embolization. *J Neurosurg* 1996;85:488–95
31. Böcher-Schwarz HG, Ringel K, Bohl J, et al. Histological findings in coil-packed experimental aneurysms 3 months after embolization. *Neurosurgery* 2002;50:379–85
32. Lewis DA, Ding YH, Dai D, et al. Morbidity and mortality associated with creation of elastase-induced saccular aneurysms in a rabbit model. *AJNR Am J Neuroradiol* 2009;30:91–94
33. Ding YH, Dai D, Danielson MA, et al. Control of aneurysm volume by adjusting the position of ligation during creation of elastase-induced aneurysms: a prospective study. *AJNR Am J Neuroradiol* 2007;28:857–59
34. Ding YH, Danielson MA, Kadirvel R, et al. Modified technique to create morphologically reproducible elastase-induced aneurysms in rabbits. *Neuroradiology* 2006;48:528–32
35. Byrne JV, Hope JK, Hubbard N, et al. The nature of thrombosis induced by platinum and tungsten coils in saccular aneurysms. *AJNR Am J Neuroradiol* 1997;18:29–33
36. Kallmes DF, Altes TA, Vincent DA, et al. Experimental side-wall aneurysms: a natural history study. *Neuroradiology* 1999;41:338–41
37. Turk AS, Aagaard-Kienitz B, Niemann D, et al. Natural history of the canine vein pouch aneurysm model. *AJNR Am J Neuroradiol* 2007;28:531–32
38. Tsumoto T, Song JK, Niimi Y, et al. Interval change in size of venous pouch canine bifurcation aneurysms over a 10-month period. *AJNR Am J Neuroradiol* 2008;29:1067–70
39. Yoshino Y, Niimi Y, Song JK, et al. Preventing spontaneous thrombosis of experimental sidewall aneurysms: the oblique cut. *AJNR Am J Neuroradiol* 2005;26:1363–65
40. Ding YH, Dai D, Lewis DA, et al. Long-term patency of elastase-induced aneurysm model in rabbits. *AJNR Am J Neuroradiol* 2006;27:139–41
41. Raymond J, Desfaits AC, Roy D. Fibrinogen and vascular smooth muscle cell grafts promote healing of experimental aneurysms treated by embolization. *Stroke* 1999;30:1657–64
42. Song JK, Niimi Y, Yoshino Y, et al. Assessment of Matrix coils in a canine model of a large bifurcation aneurysm. *Neuroradiology* 2007;49:231–35
43. Turk AS, Luty CM, Carr-Brendel V, et al. Angiographic and histological comparison of canine bifurcation aneurysms treated with first generation Matrix and standard GDC coils. *Neuroradiology* 2008;50:57–65. Epub 2007 Sep 27
44. Dai D, Ding YH, Lewis DA, et al. A longitudinal immunohistochemical study of the healing of experimental aneurysms after embolization with platinum coils. *AJNR Am J Neuroradiol* 2006;27:736–41
45. Krings T, Busch C, Sellhaus B, et al. Long-term histological and scanning electron microscopy results of endovascular and operative treatments of experimentally induced aneurysms in the rabbit. *Neurosurgery* 2006;59:911–23
46. Dai D, Ding YH, Danielson MA, et al. Histopathologic and immunohistochemical comparison of human, rabbit, and swine aneurysms embolized with platinum coils. *AJNR Am J Neuroradiol* 2005;26:2560–68
47. Dai D, Ding YH, Danielson MA, et al. Modified histologic technique for processing metallic coil-bearing tissue. *AJNR Am J Neuroradiol* 2005;26:1932–36
48. Kadirvel R, Dai D, Ding YH, et al. Endovascular treatment of aneurysms: healing mechanisms in a swine model are associated with increased expression of Matrix metalloproteinases, vascular cell adhesion molecule-1, and vascular endothelial growth factor, and decreased expression of tissue inhibitors of Matrix metalloproteinases. *AJNR Am J Neuroradiol* 2007;28:849–56
49. Hans FJ, Moller-Hartmann W, Brunn A, et al. Treatment of wide-necked aneurysms with balloon-expandable polyurethane-covered stentgrafts: experience in an animal model. *Acta Neurochir (Wien)* 2005;147:871–76. Epub 2005 Mar 17
50. Dai D, Ding YH, Lewis DA, et al. A proposed ordinal scale for grading histology in elastase-induced, saccular aneurysms. *AJNR Am J Neuroradiol* 2006;27:132–38
51. Tenjin H, Fushiki S, Nakahara Y, et al. Effect of Guglielmi detachable coils on experimental carotid artery aneurysms in primates. *Stroke* 1995;26:2075–80
52. Macdonald RL, Mojtahedi S, Johns L, et al. Randomized comparison of Guglielmi detachable coils and cellulose acetate polymer for treatment of aneurysms in dogs. *Stroke* 1998;29:478–85
53. Guglielmi G, Vinuela F, Sepetka I, et al. Electrothrombosis of saccular aneurysms via endovascular approach. Part 1. Electrochemical basis, technique, and experimental results. *J Neurosurg* 1991;75:1–7
54. Raymond J, Venne D, Allas S, et al. Healing mechanisms in experimental aneurysms. I. Vascular smooth muscle cells and neointima formation. *J Neuro-radiol* 1999;26:7–20
55. Lee D, Yuki I, Murayama Y, et al. Thrombus organization and healing in the swine experimental aneurysm model. Part I. A histological and molecular analysis. *J Neurosurg* 2007;107:94–108
56. Murayama Y, Tateshima S, Gonzalez NR, et al. Matrix and bioabsorbable polymeric coils accelerate healing of intracranial aneurysms: long-term experimental study. *Stroke* 2003;34:2031–37
57. Cruise GM, Shum JC, Plenck H. Hydrogel-coated and platinum coils for intracranial aneurysm embolization compared in three experimental models using computerized angiographic and histologic morphometry. *J Mater Chem* 2007;17:3965–73
58. Donnan GA, Davis SM. Stroke drug development: usually, but not always, animal models. *Stroke* 2005;36:23–26
59. Massoud TF, Ji C, Guglielmi G, et al. Experimental models of bifurcation and terminal aneurysms: construction techniques in swine. *AJNR Am J Neuroradiol* 1994;15:938–44
60. Yoshino Y, Niimi Y, Song J, et al. Endovascular treatment of intracranial aneurysms: comparative evaluation in a terminal bifurcation aneurysm model in dogs. *J Neurosurg* 2004;101:996–1003
61. Raymond J, Salazkin I, Gevry G, et al. Interventional neuroradiology: the role of experimental models in scientific progress. *AJNR Am J Neuroradiol* 2007;28:401–05
62. Lerouge S, Raymond J, Salazkin I, et al. Endovascular aortic aneurysm repair with stent-grafts: experimental models can reproduce endoleaks. *J Vasc Interv Radiol* 2004;15:971–79
63. Molyneux A, Ellison D, Morris J, et al. Histological findings in giant aneurysms treated with Guglielmi detachable coils: report of two cases with autopsy correlation. *J Neurosurg* 1995;83:129–32
64. Castro E, Fortea F, Villoria F. Long-term histopathologic findings in two cerebral aneurysms embolized with Guglielmi detachable coils. *AJNR Am J Neuroradiol* 1999;20:549–52
65. Ishihara S, Mawad M, Ogata K. Histopathologic findings in human cerebral aneurysms embolized with platinum coils: report of two cases and review of the literature. *AJNR Am J Neuroradiol* 2002;23:970–74
66. Groden C, Hagel C, Delling G, et al. Histological findings in ruptured aneurysms treated with GDCs: six examples at varying times after treatment. *AJNR Am J Neuroradiol* 2003;24:579–84
67. Bavinszki GT, Killer M, Talazoglu V, et al. Gross and microscopic histopathological findings in aneurysms of the human brain treated with Guglielmi detachable coils. *J Neurosurg* 1999;91:284–96
68. Molyneux A, Stratton I, Sandercock P, et al, for the International Subarachnoid Aneurysm Trial (ISAT) Collaborative Group. International Subarachnoid Aneurysm Trial (ISAT) of neurosurgical clipping versus endovascular coiling in 2143 patients with ruptured intracranial aneurysms: a randomised trial. *Lancet* 2002;360:1267–74
69. Hino K, Konishi Y, Shimada A, et al. Morphologic changes in neo-intimal proliferation in an experimental aneurysm after coil embolization: effect of factor XIII administration. *Neuroradiology* 2004;46:996–1005. Epub 2004 Nov 5
70. Moller-Hartmann W, Krings T, Stein KP, et al. Aberrant origin of the superior thyroid artery and the tracheoesophageal branch from the common carotid artery: a source of failure in elastase-induced aneurysms in rabbits. *AJR Am J Roentgenol* 2003;181:739–41
71. Thix R, Hans FJ, Krings T, et al. Haemorrhagic tracheal necrosis as a lethal complication of an aneurysm model in rabbits via endoluminal incubation with elastase. *Acta Neurochir (Wien)* 2004;146:285–89. Epub 2004 Jan 22
72. Roth C, Struffert T, Grunwald I, et al. Long-term results with Matrix coils vs. GDC: an angiographic and histopathological comparison. *Neuroradiology* 2008;50:693–99. Epub 2008 May 29